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**STUDY OF  
INTER-LIVER FUNCTIONAL COMPETITION IN  
HETEROTOPIC AUXILIARY LIVER TRANSPLANTATION  
USING A RAT MODEL**

**Faculty of Medicine  
University of Ghent**



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LIVER TRANSPLANTATION  
USING A RAT MODEL**

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## **CHAPTER 1**

### **GENERAL INTRODUCTION**

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### Liver Transplantation: successes and challenges

On the first of March 1963, Dr. Starzl and associates performed the first **liver transplantation** in human at the University of Colorado.<sup>1</sup> Despite the extensive and highly successful four-year laboratory experience with dogs, Starzl's team failed this initial clinical attempt. It took another 4 years before the first recorded success in the history of human liver transplantation was realized. On July 23<sup>rd</sup>, 1967, a 1.5 years old girl with primary hepatocellular carcinoma underwent a liver transplantation and lived for 13 months before dying from diffuse metastases. Three decades later, more than 60,000 liver transplantations have been performed worldwide,<sup>2</sup> which has indeed revolutionized the care for patients with end-stage liver disease: patients, who otherwise were doomed to die within months to years, now have the opportunity for extended survival with excellent quality of life after transplantation.<sup>3</sup> Around 80 to 90% of the patients who undergo liver transplantation today are expected to be able to survive for more than a year, and most centres report a five-year survival rate above 70%.<sup>4-6</sup> The drastically improved survival is mainly attributed to the development of effective immunosuppression agents, especially the introduction of cyclosporine and tacrolimus,<sup>7</sup> to the use of better solutions for graft preservation<sup>8, 9</sup> and to advances in anaesthetic and surgical techniques. Nowadays, liver transplantation is indicated for acute or chronic liver failure from more than sixty different diseases, amongst which cholestatic disorders, chronic hepatitis, alcoholic liver disease, metabolic diseases, fulminant hepatic failure and hepatocellular carcinoma are the most frequent indications.<sup>10</sup> The age of the patients ranges from the first week of life to adults over 75 years of age.<sup>11, 12</sup>

Despite the recorded successes of liver transplantation, the procedure still faces potential **complications**. *Poor immediate graft function* and *infection* are the major causes of graft and patients loss early after transplantation.<sup>13, 14</sup> Although immunosuppression can safely be withdrawn in about 20% of the recipients,<sup>15</sup> the majority of the long-term survivors requires life-long use of those drugs and are confronted with the chronic exposure to the *toxic effects of immunosuppressive agents*. Besides metabolic abnormalities such as diabetes mellitus, hyperlipidemia, hypertension and nephrotoxicity, even at the lower therapeutic doses of the calcineurin inhibitors (cyclosporine and tacrolimus), 80% of recipients have some degree of impaired renal function and 4% may develop renal failure with half of them requiring renal transplantation.<sup>16</sup> In addition, the growing susceptibility to infection and the rising risk of malignancy add an increasing toll to the outcome. While more promising medications will become available, efforts have been made to induce recipient tolerance specific to the transplanted donor organ. Future

applications of the donor-specific-tolerance induction in human may eventually lead to complete elimination of toxic immunosuppressive drugs. Another major problem facing transplant clinicians is the ***recurrence of the original diseases***, which may occur in the patients with autoimmune diseases (such as primary biliary cirrhosis, primary sclerosing cholangitis and autoimmune hepatitis) and viral hepatitis (such as hepatitis B and C). It has been found that there is a direct correlation between overall immunosuppression, particularly steroids and antilymphocyte preparation, and viral replication. Although with regular administration of anti-hepatitis B specific immunoglobulins, hepatitis B relapse has become rare, almost all patients transplanted for hepatitis C will relapse despite the use of ribavirin. In some of them, fluctuating transaminases, cholestasis and fibrosis will eventually develop and progress to liver failure.<sup>13</sup>

The issue of utmost importance, however, is the widening gap between the rising number of potential recipients and the limited number of donors. Till 1999, the number of patients waiting for liver transplantation has reached over 11,000 in the United States, and, tragically, many of them would have lost their lives before a proper donor is available, as figure shows that more than 1,000 patients on the waiting list died in 1997 alone.<sup>17</sup> In the Euro-Transplant countries as well as in the United Kingdom, the mortality rate of patients on the waiting list is about 15 to 18%.<sup>4</sup> To address the problem of **donor shortage**, transplant surgeons are looking at new techniques to make better use of those organs that are offered. With ***marginal donor***, such as older donors (> 60 years), history of alcoholism, elevated transaminase, steatosis, and prolonged reamnation, the uncertainty stays at whether there will be early graft function after transplantation. These donors who were turned down in the past are now being routinely utilized with success. In rare situations, even a liver with a metabolic defect, such as familial amyloidotic polyneuropathy, has been implanted into patients who would otherwise die. Further efforts are required to better define the donor criteria, so that the usable marginal organs can be distinguished from those that are unlikely to function. Other targets remain to make better match of donors and recipients, and to have greater understanding of the mechanism of ischemia-reperfusion injury.<sup>11, 18, 19</sup> The technique of ***splitting livers*** allows two patients to benefit from one donor, one with the right and the other with the left liver lobe. Besides that two transplant teams are needed to complete the recipients' operations, expertise and skill to divide a liver properly are essential for successful use of this technique.<sup>20, 21</sup> With the advantage of short cold ischemia time, the use of a ***living related donation*** is another way to expand the donor pool. The elective nature of this procedure allows it to be performed at the best time for the recipient.<sup>22-25</sup> As the donor operation is not done for the physical benefit of the healthy donor and faces a complication

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rate of 13-20% and even mortality,<sup>26</sup> moral and ethical considerations are raised on this approach. These innovative techniques have predominantly been employed in children, and up to 40% of the paediatric patients may be candidates for one of these modified procedures.<sup>27, 28</sup> The increasing shortage of donors has also encouraged the attempt to use animals as donors of transplant organs. Up to date, eight whole-organ liver transplants have been carried out using livers from chimpanzee, baboon and pig.<sup>29-31</sup> Only one recipient was able to survive for more than 70 hours.<sup>32</sup> Hyperacute rejection is the major cause of death in the immediate post-transplant period after a **xeno-transplantation**.<sup>33</sup> Using transgenic animals bearing human complement regulatory proteins or modulating the antigenic target on the donor organ will be the first step to successful xeno-transplantation.<sup>34</sup> Further problems still remain to be solved on acute and chronic rejection; uncertainty of the physiological effects of the animal proteins produced by xenograft; and the risk of contamination of humans by animal viruses. Although recent clinical studies suggest that **hepatocyte transplantation** may be useful for providing metabolic support during liver failure to bridge patients to whole organ transplantation, and for replacing whole organ transplantation in metabolic liver diseases of certain type, shortage of human livers available for hepatocyte isolation and the necessitate for long-term immunosuppression are two major hurdles to widespread application of this procedure.<sup>35</sup> Current researches in experimental models are aimed especially at developing a tightly regulated human hepatocyte cell line that grows economically in culture and exhibits differentiated liver functions.<sup>36</sup> Similarly, a **bio-artificial liver support system** capable of sustaining the life of the patients with acute liver failure could lead to improved survival and an effective device may even be able to support the recovery of the native liver without liver transplantation. However, systems as such are still under investigation.<sup>37, 38</sup>

#### Auxiliary Liver Transplantation: specificities and difficulties

In comparison to **orthotopic liver transplantation** (OLTx), in which the diseased native liver is totally removed and replaced by a liver graft, **auxiliary liver transplantation** (ALTx) is characterised by leaving a whole or a part of the native liver *in situ* in parallel to an implanted hepatic graft. This concept is originated from an experimental work of Welch in 1955.<sup>39, 40</sup> Despite the fact that dramatically improved survival rate has made OLTx the standard treatment for the patients with end-stage liver diseases, theoretically, ALTx has several **advantages** over OLTx. Firstly, in case the graft is damaged by severe post-transplant rejection or vascular complications, which would otherwise lead to a fatal consequence of hepatic failure in OLTx, the remnant native liver in ALTx may provide some degree of synthetic and

clearing liver function. The imminent death of the recipient or the necessity for immediate re-transplantation may thus be obviated.<sup>28, 41-43</sup> Secondly, liver grafts obtained by innovative surgical techniques such as split and living related donor are often relatively small in size and may not always provide sufficient function for OLTx. These grafts can still be useful for ALTx, where matching of the donor and the recipient body size is not required.<sup>44-46</sup> Thirdly, an auxiliary liver may enable patients with terminal hepatic failure to be “bridged” for a few days or possibly weeks until a proper liver donor becomes available, and in this case a heterotopic procedure could preserve the orthotopic site for subsequent transplantation. An efficient auxiliary graft may even “bridge” the patient till the native liver recovers. For this purpose, concordant species could be potential donors of auxiliary “bridging” livers for selected patients when the need for transplantation is extremely urgent.<sup>34, 47, 48</sup>

The first auxiliary liver transplantation in human was performed by Absolon in 1964,<sup>49</sup> and it was till 1972 when an auxiliary transplantation truly prolonged a human life.<sup>50</sup> During the following two decades, ALTx was done solely in a heterotopic manner - **heterotopic auxiliary liver transplantation** (HALTx), where a graft (usually partial) is placed below the un-resected native liver. The initial clinical results of HALTx were rather disappointing. Amongst the reported fifty HALTx during the period 1964-1984, only two patients survived longer than a year, and one of them lived fourteen and half years after transplantation with a well functioning graft.<sup>51-53</sup> One of the major problems associated with HALTx is the limited space in the abdominal cavity for an additional large liver graft and the gravity gradient of the venous outflow caused by implanting the engrafted liver at a non-physiological position. This may lead to insufficient venous drainage of the engrafted liver and inadequate portal perfusion of the engrafted and even the native liver.<sup>54</sup> An insufficient reperfusion of the liver graft could cause primary non-function immediately after transplantation, and, at the long-term, graft atrophy may occur due to inter-liver competition.<sup>55-59</sup> Though the portal “banding” technique is a common practice to secure a sufficient amount of portal flow to the graft,<sup>56-59</sup> the difficulty often remains on the proportional distribution of portal blood to the two livers according to the purpose of the transplantation.<sup>60</sup> Vascular complications especially thrombosis of the portal vein are also more likely to develop after HALTx than OLTx. The high morbidity and mortality caused by these disadvantages have led to temporary abandonment of HALTx in the early 70s. Many efforts have been made ever since to improve post-transplant survival. Most notably, based upon the experiences in animal studies,<sup>53, 61-66</sup> the contributions of Terpstra’s group have improved the surgical techniques of HALTx with markedly increased post-HALTx survival rate.<sup>54, 58, 66-71</sup> Since 1980s’, the concept of ALTx has further been extended by the introduction of a new approach - **auxiliary**

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**partial orthotopic liver transplantation** (APOLTx), where the left or the right lobe of the native liver is resected and replaced by an auxiliary graft.<sup>28, 41, 42, 72-78</sup>

The physiological position of the hepatic graft by this approach results in an optimal outflow pressure. Accumulating clinical results have shown a reduced incidence of post-transplant portal thrombosis,<sup>41, 77, 79</sup> and banding of the portal vein to the native liver is also less required except in cases of non-cirrhotic metabolic liver diseases.<sup>42, 78, 80, 81</sup> Since the use of only segments 2 and 3 of the graft usually provides sufficient function, this technique is commonly performed with reduced-size, split or living related donor livers.<sup>28, 43, 81, 81</sup>

Up to date, ALTx, particularly APOLTx, has been considered as a successful alternative to OLTx in specific clinical situations. HALTx, which avoids the surgical trauma of more extensive liver dissection, has been used to treat selected patients with end-stage *chronic liver diseases* at high risk for OLTx.<sup>52, 67-69, 82-84</sup> Currently, most of the alleged difficulties of native hepatectomy are no longer relevant for experienced surgeons.<sup>54</sup> The major concern for using HALTx to treat patients in this category lies on the original disease in the remaining native liver, which may further affect the engrafted liver, and may promote secondary tumour development.

Children suffering from liver diseases represent about 15% to 20% of the liver transplant registrants.<sup>85</sup> They have a specific spectrum of indications with mainly hepatic-based *inborn errors of metabolism*.<sup>86</sup> Since many of these diseases cause structural hepatic damage and eventually lead to cirrhosis, whole-liver replacement by OLTx is the preferred procedure. For certain types of non-cirrhotic metabolic disorders, such as type 1 Crigler-Najjar syndrome, urea cycle enzyme deficiencies, disorders of fatty acid metabolism, familial hypercholesterolemia, hemophilia and ornithine transcarbamylase deficiency, an auxiliary liver may correct the partial enzymatic deficiency responsible for the disease without the need to remove the otherwise normal native liver.<sup>41, 42, 78, 81, 86-89</sup> ALTx also preserves the patient's native liver, which remains accessible for future gene transfer therapy.<sup>27, 90, 91</sup> Difficulties stay on how to determine the optimal graft volume required to correct the metabolic deficiency, to appreciate the necessary portal supply to the two livers and to avoid the possible damaging effects of the retaining diseased native liver. The long-term prospect of this procedure needs more time to adjudge.

It has been shown that even with optimal intensive care, less than 20% of the patients with *acute liver failure* and *sub-acute liver failure* may survive without a liver transplantation.<sup>92, 93</sup> Even though the failing liver may have the potential to regenerate with time, most patients will suffer fatal neurologic damage before the liver recovery actually occurs. To date, liver

transplantation is still the best life saving procedure for many patients with acute liver failure. Although ALTx was initially proposed as a “bridge” to “tide the patient over” the period of acute hepatic insufficiency,<sup>40</sup> currently, most of the lives of the patients with acute liver failure are saved by OLTx with a one-year survival rate of over 60%.<sup>94-96</sup> Evidently, the potential recovery of the failing liver is completely negated by the total removal of the native liver in the OLTx procedure, and the majority of the patients who survive the OLTx surgery have to depend on life-long use of immunosuppressive drugs. In contrast, an auxiliary liver graft may provide temporary functional support for the patient to allow the native liver to recover. Following a sufficient recuperation of the diseased native liver, the auxiliary liver graft can then be either removed or left to atrophy. The toxic immunosuppressive agents can be tapered or eventually withdrawn, and its consequent complications will thus be avoided.<sup>57-59, 72, 75-77, 97-109</sup> Recently, Erhard et al. have demonstrated the feasibility of using arterialised portal supply to sustain the liver graft in ALTx for acute liver failure patients. The main advantage of this modified approach is to leave the hilum and the portal vein of the native liver untouched, which may reduce the incidence of possible complications such as arterial vascular stenosis or portal vein thrombosis caused by performing intrahilar vascular dissection and anastomosis. It also avoids disturbance of the native liver recovery due to “portal flow steal phenomenon”.<sup>110, 111</sup> Current difficulties of using ALTx to treat patients with acute liver failure are mainly twofold: (1) There is still a lack of reliable pre-operative measurements to assess regenerating potential of the native liver for selecting appropriate candidates for the procedure. Indeed, histological evaluation of the presence of viable hepatocytes, fibrosis or regeneration in the diseased liver does not appear to have clear predictive value;<sup>99, 112, 113</sup> (2) The remaining, severely diseased native liver may put the newly engrafted liver at risk and lead to functional failure, or may also cause development of the so-called “toxic liver syndrome” with subsequent multi-organ failure.<sup>104, 114, 115</sup>

### **Rat Model of Auxiliary Liver Transplantation: evolution and questions**

The first animal liver transplantation was performed in the dog by Welch in 1955<sup>39</sup> and later studies described this attempt in other animal species including the baboon,<sup>116</sup> the monkey,<sup>117</sup> the pig,<sup>118</sup> the mouse<sup>119</sup> and the rat.<sup>120</sup> The use of genetically well-defined inbred strains of rats allows indeed a variety of experiments to be set up with controlled immunological features. This, in addition to other advantages, has made the rat the most commonly used experimental animal for liver transplantation studies, since large animals such as the pig or the dog are of limited use for immunological studies due to

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the difficulty of obtaining inbred strains.

In 1966, Lee and Edgington introduced the first rat model of ALTx. In this initial model, 32% of the donor liver was implanted at a heterotopic position; the blood supply of the engrafted liver was exclusively arterial from the recipient's coeliac axis; and the venous return of the graft went through the suprahepatic inferior donor vena cava anastomosed to the infrahepatic native cava. The portal vein, the infrahepatic inferior vena cava and the common bile duct of the liver graft were ligated. Apart from a liver mass reduction of approximate 68%, the portal vein and the hepatic arterial supply as well as the bile drainage of the recipient's native liver remained intact.<sup>120</sup> Further modification was added to this model by the same authors two years later. At this time, the engrafted liver was re-vascularized with both the portal venous and the hepatic arterial supply, while the native liver was deprived of portal blood. The bile drainage of the engrafted liver was not yet reconstructed. Serum aspartate aminotransferase activity and histology were used as indicators of allogeneic liver rejection. In addition to the description of histopathological changes during the rejection process, it was also observed that re-establishing portal supply to the engrafted liver would prompt graft regeneration. It was therefore concluded that the portal blood flow was the major factor influencing liver regenerative capacity.<sup>121</sup>

Kort et al. reported a technique, in which the infrahepatic donor vena cava was used to drain the engrafted liver in the rat HALTx model. It appeared that this modification could drastically reduce the incidence of post-transplant graft congestion, which was frequently seen in the original model. Consequently, the survival rate after HALTx was improved. However, neither the hepatic artery nor the bile drainage of the engrafted liver was reconstructed.<sup>122</sup>

Further improvement of survival was obtained by performing the procedure of re-connecting the graft common bile duct to the recipient's duodenum.<sup>123</sup> A delicate method to share portal blood supply between the two livers was later described by Hess et al. The engrafted liver was perfused with mesentericosplenic venous blood, while the native liver retained the pancreaticoduodenal portal flow along with hepatic arterial supply. Due to technical difficulties, hepatic arterial supply to the engrafted liver was not re-established.<sup>124</sup> By examining the effects of different surgical manipulations of the native liver on the graft regeneration, their studies showed that the fate of the auxiliary liver graft was directly influenced by the functional state of the recipient's native liver independent of the amount of portal blood flow.<sup>124, 125</sup>

Müller reported his version of rat HALTx in 1983, in which the liver graft was implanted in the left loin following a left nephrectomy. The auxiliary liver was supplied with sole arterial blood through an anastomosis between



the recipient's left renal artery and the graft's portal vein, while the hepatic artery of the graft was ligated. This made it the first rat model of ALTx with an arterialised portal supply. Venous outflow of the engrafted liver was drained into the recipient's vena cava through an anastomosis between the inferior vena cava of the graft and the left renal vein of the recipient. The common bile duct of the engrafted liver was connected to the recipient's left ureter. No surgical manipulation was done on the native liver. Instead of conventional hand-sutured technique, an intra-vascular polyethylene "stent" was used to connect the graft's portal vein to the recipient's renal artery. This simplified technique shortened the total ischemic time of the graft and led to an improved two-week survival rate of 77% after transplantation. In this specific setting of a portal-arterialised liver graft functioning in parallel with a normal native liver, the isogenic auxiliary liver graft showed gradual atrophy, but its histological microstructure was preserved even six months after transplantation.<sup>126</sup>

The "cuff" technique, or extra-vascular stent, is another approach to simplify the method of micro-vascular anastomosis. It was applied in rat HALTx surgery by Marni et al. to perform both portal vein and vena cava anastomoses. With this method, occlusion time of the recipient's portal vein needed to perform the portal anastomosis and the total ischemic time of the liver graft were considerably decreased, which contributed to a two-week survival rate as high as 90%. To favour the competitive position of the engrafted liver, the recipient's native liver was handicapped with both a partial hepatectomy and a common bile duct ligation, while no re-arterialisation of the graft was attempted. Two months after the transplantation, the engrafted livers enlarged to a size twofold that of the original ones, whereas the recipient's native livers appeared atrophic. The authors mentioned the interest of using this model for transplantation immunology studies.<sup>127</sup>

Lee et al. described two specific variations of rat HALTx in 1986. The first one was called the "double-liver" model, in which the suprahepatic inferior vena cava of the engrafted liver was anastomosed to the infrahepatic portion of the recipient inferior vena cava; the graft's portal vein was anastomosed more distally to the recipient's inferior vena cava at the level of the renal vein. The inferior vena cava between the two anastomoses was then ligated, which led to the perfusion of the engrafted liver with systemic venous blood. The graft's hepatic artery was anastomosed to the recipient's infrarenal aorta. No manipulation was done on the native liver. Along with its later arterial-venous fistula version,<sup>128-130</sup> this model was designed to study liver regeneration following transplantation. The second variation introduced by Lee was the so-called "reversed circulation" model. The recipient's portal vein was connected to the suprahepatic vena cava of the engrafted liver, and

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the engrafted liver was drained through an anastomosis between the graft's portal vein and the recipient's infrahepatic vena cava. The native liver was thus deprived of its portal supply. This model was developed to study the effects of a reversed circulation on the hepatic physiology.<sup>131</sup>

Schleimer et al. recently reported a new approach in the rat HALTx, in which the portal vein of the engrafted liver was arterIALIZED via the recipient's renal artery.<sup>132</sup> The difference with Müller's technique is that the engrafted liver's outflow was made possible through an anastomosis performed directly between the donor and the recipient vena cava, and the bile duct was re-connected to the recipient's duodenum. The recipient's native liver was subjected to an 85% partial hepatectomy in order to induce acute liver failure. With the support of a portal arterIALIZED auxiliary liver, the failing native liver regenerated to about 2.5% of the body weight and 83% of the rats did survive an observation period of three months.<sup>133</sup>

To date, two different types of APOLTx models have been described in the rat. The partial liver graft was implanted at an orthotopic site either above - in a reversal fashion<sup>134, 135</sup> or at the left side of the partial hepatectomised native liver.<sup>136</sup>

The rat model of ALTx has been used for various studies such as to evaluate new surgical techniques,<sup>120, 122, 123, 126, 127, 131, 135, 137, 138</sup> and related physiological<sup>128, 129, 139, 140</sup> or morphological changes of the liver;<sup>141, 142</sup> to test its feasibility for correcting metabolic deficiency<sup>143-146</sup> or for maintaining hepatic function and supporting native liver regeneration at the time of acute liver failure;<sup>133</sup> to study transplantation immunology<sup>126, 147-149, 149-152</sup> and to investigate the mechanisms of inter-liver functional competition.<sup>120, 121, 124, 125, 133, 153-160</sup>

In addition to technical difficulties, there are other problems with the use of the rat model of HALTx. Comparing to OLTx, the rat model of HALTx is characterized by the coexistence of the engrafted liver and the recipient's native liver after the transplantation. The commonly used parameters such as animal survival and serum biochemical tests could thus not discriminate the functionality of each individual liver. The question remains on how to evaluate the post-transplant viability of an individual liver, especially the engrafted organ. A possible approach is to handicap the native liver to such a degree that it will be insufficient to guarantee survival. The post-transplant animal survival and liver biology will then necessarily reflect the viability of the newly engrafted liver. In the previous studies, different surgical manipulations of the native liver have been used for this purpose either with a 68% partial hepatectomy alone<sup>121, 122</sup> or with a 68% partial hepatectomy plus a common bile duct ligation.<sup>123, 127</sup> Up to date, there has been no report on the possible differential effects of these various approaches on post-transplant

animal and graft survival, as well as the animal serum biochemical values. This was investigated in the current work. Furthermore, as a non-invasive measurement, magnetic resonance imaging (MRI) was tested for its capability to monitor the post-transplant hepatic viability in this rat model.

After HALTx, the interaction of the two livers also influences transplantation immunology<sup>126, 147-149, 149-152</sup> and may play a role in the inter-liver competition. Starzl et al. were among the first to observe that an auxiliary liver graft would atrophy in the presence of a healthy recipient's native liver after HALTx.<sup>161, 162</sup> The term of "functional competition", which was originally used to describe the compensatory hyperplasia of one liver lobe after occlusion of the bile duct of the other lobe,<sup>163</sup> was applied to depict this phenomenon. This competitive behaviour of the two livers has also been noticed later in the APOLTx.<sup>42, 78, 80</sup> Despite many efforts made over the past three decades, the precise mechanisms of this phenomenon are still not fully understood. Some investigators claim that atrophy or hypertrophy of the liver graft is the result of the functional competition between the two livers governed by the body demand for liver function. Shalm<sup>164</sup> and Hess et al.<sup>123</sup> showed that atrophy of the auxiliary liver together with functional deterioration of the graft was due to the presence of the recipient's un-handicapped native liver. Van der Heyde et al. demonstrated in the dog that by increasing a certain degree of functional impairment on the native liver, for instance through performing a common bile duct ligation, the graft atrophy could be prevented.<sup>165</sup> Hess confirmed these results in a rat model and showed that the auxiliary engrafted liver would be replaced by connective tissue if no manipulations on the native liver were done.<sup>123</sup> Van der Heyde et al. also reported that a host portacaval shunt would enhance the function of the graft and prevent its atrophy.<sup>166</sup> In contrary, other investigators dispute this theory and emphasize the importance of portal blood itself for maintenance of hepatic graft integrity by showing that the graft atrophy can be prevented by diversion of all portal flow through the graft.<sup>162, 167</sup> Using a "split transposition" dog model, Marchioro et al. reported that the liver fragments receiving splanchnic blood were always hypertrophic compared with the hepatic lobes perfused by systemic venous blood. Their studies suggested that only the splanchnic portal inflow, which carried certain substances, was essential for the preservation of hepatocytes integrity and could boost liver regeneration.<sup>162, 168, 169</sup> In a series of studies using this "double fragment model", in which the separated two liver fragments were provided with blood of various sources, Starzl et al. claimed that the main splanchnic venous "hepatotrophic factors" were endogenous hormones, amongst which the single most important was insulin.<sup>170-173</sup> While new substances with hepatotrophic effect continue to emerge, to date, it has generally been accepted that hormone hepatotrophic factors are the key elements for liver regeneration. Therefore, after HALTx, the liver that

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receives the major part of portal supply will eventually regenerate, and the final outcome of the inter-liver competition actually depends upon the distribution of the “hepatotrophic factors” between the two livers.<sup>42, 56-59, 80, 153, 154, 162, 170, 171, 174-178</sup>

No consensus exists so far on whether the physiological status of each individual liver may influence the inter-liver competition. This was investigated in the current work using a rat model of HALTx, in which the physiological balance was favourably slanted towards either the engrafted liver or the recipient’s native liver through surgical manipulations. In addition, an effort was also made to re-examine the controversy on whether changing the content of the portal supply would alter hepatic regeneration capacity following a standard partial hepatectomy.

## REFERENCES

1. Starzl TE, Marchioro TL, Porter KA. Experimental and clinical observations after homotransplantation of the whole liver. *Rev Int Hepatol* 1965; 15(8):1447-1480.
2. Wall WJ. Liver transplantation: past accomplishments and future challenges. *Can J Gastroenterol* 1999; 13(3):257-263.
3. Belle SH, Porayko MK, Hoofnagle JH, Lake JR, Zetterman RK. Changes in quality of life after liver transplantation among adults. National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK) Liver Transplantation Database (LTD). *Liver Transpl Surg* 1997; 3(2):93-104.
4. Neuberger J. Liver transplantation. *QJM* 1999; 92(10):547-550.
5. Pirenne J, Koshiha T. Present status and future prospects in liver transplantation. *Int Surg* 1999; 84 (4):297-304.
6. Wong WW, Bain VG. Update in liver transplantation. *Can Fam Physician* 1999; 45:1241-1249.
7. Starzl TE, Iwatsuki S, Van Thiel DH, Gartner JC, Zitelli BJ, Malatack JJ, Schade RR et al. Evolution of liver transplantation. *Hepatology* 1982; 2(5):614-636.
8. Kalayoglu M, Sollinger HW, Stratta RJ, D'Alessandro AM, Hoffmann RM, Pirsch JD, Belzer FO. Extended preservation of the liver for clinical transplantation. *Lancet* 1988; 1 (8586):617-619.
9. Todo S, Nery J, Yanaga K, Podesta L, Gordon RD, Starzl TE. Extended preservation of human liver grafts with UW solution. *JAMA* 1989; 261(5):711-714.
10. Carithers RLJ. Liver transplantation. American Association for the Study of Liver Diseases. *Liver Transpl* 2000; 6(1):122-135.
11. Detre KM, Lombardero M, Belle S, Beringer K, Breen T, Daily OP, Ascher NL. Influence of donor age on graft survival after liver transplantation--United Network for Organ Sharing Registry. *Liver Transpl Surg* 1995; 1(5):311-319.
12. Woodle ES, Millis JM, So SK, McDiarmid SV, Busuttil RW, Esquivel CO,

## CHAPTER 1 General Introduction 13

- Whittington PF et al. Liver transplantation in the first three months of life. *Transplantation* 1998; 66 (5):606-609.
13. Pirenne J, Koshiha T. Present status and future prospects in liver transplantation. *Int Surg* 1999; 84 (4):297-304.
  14. Wong WW, Bain VG. Update in liver transplantation. *Can Fam Physician* 1999; 45:1241-1249.
  15. Devlin J, Doherty D, Thomson L, Wong T, Donaldson P, Portmann B, Williams R. Defining the outcome of immunosuppression withdrawal after liver transplantation. *Hepatology* 1998; 27(4):926-933.
  16. Fisher NC. Renal failure after lung transplantation. *Lancet* 1998; 352 (9121):69-70.
  17. Penko ME, Tirbaso D. An overview of liver transplantation. *AACN Clin Issues* 1999; 10(2):176-184.
  18. Pruim J, Klompmaker IJ, Haagsma EB, Bijleveld CM, Slooff MJ. Selection criteria for liver donation: a review. *Transpl Int* 1993; 6 (4):226-235.
  19. Mirza DF, Gunson BK, Da Silva RF, Mayer AD, Buckels JA, McMaster P. Policies in Europe on "marginal quality" donor livers. *Lancet* 1994; 344(8935):1480-1483.
  20. Busuttill RW, Goss JA. Split liver transplantation. *Ann Surg* 1999; 229(3):313-321.
  21. Sindhi R, Rosendale J, Mundy D, Taranto S, Baliga P, Reuben A, Rajagopalan PR et al. Impact of segmental grafts on pediatric liver transplantation--a review of the United Network for Organ Sharing Scientific Registry data (1990-1996). *J Pediatr Surg* 1999; 34 (1):107-110.
  22. Raia S, Nery JR, Mies S. Liver transplantation from live donors. *Lancet* 1989; 2 (8661):497.
  23. Strong RW, Lynch SV, Ong TH, Matsunami H, Koido Y, Balderson GA. Successful liver transplantation from a living donor to her son. *N Engl J Med* 1990; 322 (21):1505-1507.
  24. Tanaka K, Uemoto S, Honda K, Morimoto T, Tanaka A, Shimabara Y, Mori K et al. Living related liver transplantation. *Nippon Rinsho* 1991; 49(11):2725-2731.
  25. Testa G, Malago M, Broelsch CE. Living-donor liver transplantation in adults. *Langenbecks Arch Surg* 1999; 384 (6):536-543.
  26. Grewal HP, Thistlewaite JRJ, Loss GE, Fisher JS, Cronin DC, Siegel CT, Newell KA et al. Complications in 100 living-liver donors. *Ann Surg* 1998; 228(2):214-219.
  27. Pappas SC, Rouch DA, Stevens LH. New techniques for liver transplantation: reduced-size, split-liver, living-related and auxiliary liver transplantation. *Scand J Gastroenterol Suppl* 1995; 208:97-100.
  28. Uemoto S, Yabe S, Inomata Y, Nishizawa H, Asonuma K, Egawa H, Kiuchi T et al. Coexistence of a graft with the preserved native liver in auxiliary partial orthotopic liver transplantation from a living donor for ornithine transcarbamylase deficiency. *Transplantation* 1997; 63 (7):1026-1028.
  29. Starzl TE, Fung J, Tzakis A, Todo S, Demetris AJ, Marino IR, Doyle H et al. Baboon-to-human liver transplantation. *Lancet* 1993; 341(8837):65-71.

## 14 CHAPTER 1 General Introduction

30. Fujioka H, Cramer DV, Yasunaga C, Tusso PJ, Wu GD, Middleton Y, Moscioni AD et al. Differential patterns of reaction of human natural antibodies to pig hepatocytes and vascular endothelium. *Liver Transpl Surg* 1995; 1(2):85-93.
31. Lambrechts D, Sachs DH, Cooper DK. Discordant organ xenotransplantation in primates: world experience and current status. *Transplantation* 1998; 66(5):547-561.
32. Cardoso J. Utilization of xenogenic liver in human clinics. Tests and perspectives. *Pathol Biol* 2000; 48 (4):362-364.
33. Makowka L, Cramer DV, Hoffman A, Breda M, Sher L, Eiras-Hreha G, Tusso PJ et al. The use of a pig liver xenograft for temporary support of a patient with fulminant hepatic failure. *Transplantation* 1995; 59 (12):1654-1659.
34. Fung J, Rao A, Starzl T. Clinical trials and projected future of liver xenotransplantation. *World J Surg* 1997; 21(9):956-961.
35. Strom SC, Chowdhury JR, Fox JJ. Hepatocyte transplantation for the treatment of human disease. *Semin Liver Dis* 1999; 19(1):39-48.
36. Kobayashi N, Noguchi H, Watanabe T, Matsumura T, Totsugawa T, Fujiwara T, Westerman K et al. Establishment of a tightly regulated human cell line for the development of hepatocyte transplantation. *Hum Cell* 2000; 13(1):7-13.
37. Koebe HG, Schildberg FW. The artificial liver--an interim report. *Wien Klin Wochenschr* 1998; 110 (16):551-563.
38. Dowling DJ, Mutimer DJ. Artificial liver support in acute liver failure. *Eur J Gastroenterol Hepatol* 1999; 11(9):991-996.
39. Welch CS. A note on the transplantation of the whole liver in dogs. *Transplant Bull* 1955; 2:54-56.
40. Goodrich EOJ, Welch HF, Nelson JA, Beecher TS, Welch CS. Homotransplantation of the canine liver. *Surgery* 1956; 39:244-251.
41. Broelsch CE, Emond JC, Whittington PF, Thistlethwaite JR, Baker AL, Lichtor JL. Application of reduced-size liver transplants as split grafts, auxiliary orthotopic grafts, and living related segmental transplants. *Ann Surg* 1990; 212(3):368-375.
42. Whittington PF, Emond JC, Heffron T, Thistlethwaite JR. Orthotopic auxiliary liver transplantation for Crigler-Najjar syndrome type 1. *Lancet* 1993; 342 (8874):779-780.
43. Kaibori M, Uemoto S, Fujita S, Inomata Y, Egawa H, Asonuma K, Kiuchi T et al. Native hepatectomy after auxiliary partial orthotopic liver transplantation. *Transpl Int* 1999; 12(5):383-386.
44. Emond JC, Heffron TG, Kortz EO, Gonzalez-Vallina R, Contis JC, Black DD, Whittington PF. Improved results of living-related liver transplantation with routine application in a pediatric program. *Transplantation* 1993; 55(4):835-840.
45. Rogiers X, Malago M, Habib N, Knoefel WT, Pothmann W, Burdelski M, Meyer-Moldenhauer WH et al. In situ splitting of the liver in the heart-beating cadaveric organ donor for transplantation in two recipients. *Transplantation* 1995; 59 (8):1081-1083.

46. Egawa H, Tanaka K, Inomata Y, Uemoto S, Okajima H, Satomura K, Kiuchi T et al. Auxiliary partial orthotopic liver transplantation from a living related donor: a report of two cases. *Transplant Proc* 1996; 28 (2):1071-1072.
47. Starzl TE, Marchioro TL, Faris TD, McCardle RJ, Iwaski Y. Avenues of future research in homotransplantation of the liver with particular reference to hepatic supportive procedures, antilymphocyte serum, and tissue typing. *Am J Surg* 1966; 112 (3):391-400.
48. Mieles L, Ye Y, Luo Y, Kobayashi T, Li SF, Niekrasz M, Kosanke S et al. Auxiliary liver allografting and xenografting in the nonhuman primate. *Transplantation* 1995; 59 (12):1670-1676.
49. Absolon KB, Hagihara PF, Griffen WOJ, Lillehei RC. Experimental and clinical heterotopic liver homotransplantation. *Rev Int Hepatol* 1965; 15(8):1481-1490.
50. Fortner JG, Kinne DW, Shiu MH, Howland WS, Kim DK, Castro EB, Yeh SD et al. Clinical liver heterotopic (auxiliary) transplantation. *Surgery* 1973; 74(5):739-751.
51. Fortner JG, Yeh SD, Kim DK, Shiu MH, Kinne DW. The case for and technique of heterotopic liver grafting. *Transplant Proc* 1979; 11(1):269-275.
52. Houssin D, Franco D, Berthelot P, Bismuth H. Heterotopic liver transplantation in end-stage HBsAg-positive cirrhosis. *Lancet* 1980; 1(8176):990-993.
53. Terpstra OT, Schalm SW, Reuvers CB, Baumgartner D, Groenland TH, ten KF, Stibbe J et al. The role of auxiliary liver transplantation. *Transplant Proc* 1987; 19(5):4370-4372.
54. Blankensteijn JD, Schalm SW, Terpstra OT. New aspects of heterotopic liver transplantation. *Transpl Int* 1992; 5 (1):43-50.
55. Provoost AP, Madern GC, Sinaasappel M, Terpstra OT, Molenaar JC. Successful prolonged correction of an inborn metabolic defect by heterotopic auxiliary liver transplantation in a dog model. *Transplant Proc* 1993; 25(2):1950-1951.
56. Nagashima I, Bergmann L, Schweizer R. How can we share the portal blood inflow in auxiliary partial heterotopic liver transplantation without portal hypertension? *Surgery* 1994; 116(1):101-106.
57. Moritz MJ, Jarrell BE, Munoz SJ, Maddrey WC. Regeneration of the native liver after heterotopic liver transplantation for fulminant hepatic failure. *Transplantation* 1993; 55(4):952-954.
58. Metselaar HJ, Hesselink EJ, de Rave S, ten Kate FJ, Lameris JS, Groenland TH, Reuvers CB et al. Recovery of failing liver after auxiliary heterotopic transplantation. *Lancet* 1990; 335(8698):1156-1157.
59. van Hoek B, Ringers J, Kroes AC, van Krieken JH, van Schelven WD, Masclee AA, van Krieken-Hogenberk LG et al. Temporary heterotopic auxiliary liver transplantation for fulminant hepatitis B. *J Hepatol* 1995; 23(2):109-118.
60. de Jonge J, Madern GC, Terpstra OT, Sinaasappel M, Molenaar JC, Provoost AP, Tilanus HW. Directing portal flow is essential for graft

## 16 CHAPTER 1 General Introduction

- survival in auxiliary partial heterotopic liver transplantation in the dog. *J Pediatr Surg* 1999; 34(8):1265-1268.
61. Reuvers CB, Terpstra OT, de Groot GH, Kooy PP, ten Kate FW, Schalm SW, Molenaar JC et al. Auxiliary transplantation of a partially hepatectomized liver in pigs with fulminant hepatic failure. *Transplant Proc* 1984; 16(5):1236-1239.
  62. Reuvers CB, Terpstra OT, ten Kate FW, Kooy PP, Molenaar JC, Jeekel J. Long-term survival of auxiliary partial liver grafts in DLA-identical littermate beagles. *Transplantation* 1985; 39(2):113-118.
  63. Reuvers CB, Terpstra OT, Boks AL, de Groot GH, Jeekel J, ten Kate FW, Kooy PP et al. Auxiliary transplantation of part of the liver improves survival and provides metabolic support in pigs with acute liver failure. *Surgery* 1985; 98(5):914-921.
  64. Reuvers CB, Terpstra OT, ten Kate FJ, Kooy PP, Provoost AP, Molenaar JC, Jeekel J. Rejection and survival of auxiliary partial liver grafts in non-tissue-typed pigs. *Eur Surg Res* 1986; 18(2):86-95.
  65. Reuvers CB, Terpstra OT, Groenland TH, Boks AL, Faithfull NS, ten KF. Hemodynamics and coagulation in experimental auxiliary liver transplantation during fulminant hepatic failure. *Ann Surg* 1986; 204(5):552-558.
  66. Terpstra OT, Reuvers CB, Schalm SW. Auxiliary heterotopic liver transplantation. *Transplantation* 1988; 45(6):1003-1007.
  67. Terpstra OT, Schalm SW, Weimar W, Willemse PJ, Baumgartner D, Groenland TH, ten Kate FW et al. Auxiliary partial liver transplantation for end-stage chronic liver disease. *N Engl J Med* 1988; 319 (23):1507-1511.
  68. Metselaar HJ, Hesselink EJ, de Rave S, Groenland TH, Bakker CM, Weimar W, Schalm SW et al. A comparison between heterotopic and orthotopic liver transplantation in patients with end-stage chronic liver disease. *Transplant Proc* 1991; 23(1 Pt 2):1531-1532.
  69. Willemse PJ, Ausema L, Terpstra OT, Krenning EP, ten Kate FW, Schalm SW. Graft regeneration and host liver atrophy after auxiliary heterotopic liver transplantation for chronic liver failure. *Hepatology* 1992; 15(1):54-57.
  70. ten Kate FJ, Schalm SW, Willemse PJ, Blok AP, Heijtkink RA, Terpstra OT. Course of hepatitis B and D virus infection in auxiliary liver grafts in hepatitis B-positive patients. A light-microscopic and immunohistochemical study. *J Hepatol* 1992; 14 (2-3):168-175.
  71. Terpstra OT. Auxiliary liver grafting: a new concept in liver transplantation. *Lancet* 1993; 342(8874):758.
  72. Bismuth H, Houssin D. Partial resection of liver grafts for orthotopic or heterotopic liver transplantation. *Transplant Proc* 1985; 17:279-283.
  73. Then PK, Feldman L, Broelsch CE. Flow and vascular resistance measurements in auxiliary liver segments transplanted in orthotopic position. *Transplant Proc* 1989; 21(1 Pt 2):2378-2380.
  74. Broelsch CE, Whittington PF, Emond JC. Evolution and future perspectives for reduced-size hepatic transplantation. *Surg Gynecol Obstet* 1990; 171(4):353-360.



75. Gubernatis G, Pichlmayr R, Kemnitz J, Gratz K. Auxiliary partial orthotopic liver transplantation (APOLT) for fulminant hepatic failure: first successful case report. *World J Surg* 1991; 15(5):660-665.
76. Boudjema K, Jaeck D, Simeoni U, Becmeur F, Schieffer F, Bientz J, Brunot P et al. Temporary auxiliary orthotopic transplantation of a reduced liver for fulminant hepatitis. A successful case in a child. *Chirurgie* 1993; 119(5):257-262.
77. Boudjema K, Cherqui D, Jaeck D, Chenard-Neu MP, Steib A, Freis G, Becmeur F et al. Auxiliary liver transplantation for fulminant and subfulminant hepatic failure. *Transplantation* 1995; 59(2):218-223.
78. Rela M, Muiesan P, Andreani P, Gibbs P, Mieli-Vergani G, Mowat AP, Heaton ND. Auxiliary liver transplantation for metabolic diseases. *Transplant Proc* 1997; 29(1-2):444-445.
79. van Hoek B, de Boer J, Boudjema K, Williams R, Corsmit O, Terpstra OT. Auxiliary versus orthotopic liver transplantation for acute liver failure. EURALT Study Group. European Auxiliary Liver Transplant Registry. *J Hepatol* 1999; 30(4):699-705.
80. Rogiers X, Malago M, Habib N, Knoefel WT, Pothmann W, Burdelski M, Meyer-Moldenhauer WH et al. In situ splitting of the liver in the heart-beating cadaveric organ donor for transplantation in two recipients [see comments]. *Transplantation* 1995; 59(8):1081-1083.
81. Kaibori M, Egawa H, Inomata Y, Uemoto S, Asonuma K, Kiuchi T, Varela-Fascineto G et al. Selective portal blood flow diversion in auxiliary partial orthotopic liver transplantation to induce regeneration of the graft. *Transplantation* 1998; 66(7):935-937.
82. Rinkes IH, Van der Hoop AG, Hesselink EJ, Metselaar H, de Rave S, Zonderland HM, Schalm SW et al. Does auxiliary heterotopic liver transplantation reverse hypersplenism and portal hypertension? [see comments]. *Gastroenterology* 1991; 100(4):1126-1128.
83. Metselaar HJ, Hesselink EJ, Schalm SW, Terpstra OT. Favorable results of auxiliary heterotopic liver transplantation in patients with end-stage chronic liver insufficiency. *Ned Tijdschr Geneesk* 1991; 135(27):1221-1226.
84. Haberal M, Karakayali H, Boyacioglu S, Gur G, Baysal C, Arslan G, Moray G et al. Successful living-related heterotopic auxiliary liver transplantation for chronic Budd-Chiari syndrome. *Transplant Proc* 1999; 31(7):2902-2903.
85. Busuttil RW, Goss JA. Split liver transplantation. *Ann Surg* 1999; 229(3):313-321.
86. Burdelski M, Rogiers X. Liver transplantation in metabolic disorders. *Acta Gastroenterol Belg* 1999; 62(3):300-305.
87. Inui A, Fujisawa T, Komatsu H, Tanaka K, Inui M. Histological improvement in native liver after auxiliary partial liver transplantation for ornithine transcarbamylase deficiency. *Lancet* 1996; 348(9029):751-752.
88. Kiuchi T, Edamoto Y, Kaibori M, Uryuhara K, Kasahara M, Uemoto S, Egawa H et al. Auxiliary liver transplantation for urea-cycle enzyme deficiencies: lessons from three cases. *Transplant Proc* 1999; 31(1-2):528-529.

## 18 CHAPTER 1 General Introduction

89. Rela M, Muiesan P, Vilca-Melendez H, Dhawan A, Baker A, Mieli-Vergani G, Heaton ND. Auxiliary partial orthotopic liver transplantation for Crigler-Najjar syndrome type I. *Ann Surg* 1999; 229(4):565-569.
90. Rela M, Muiesan P, Heaton ND, Corbally M, Hajj H, Mowat AP, Williams R et al. Orthotopic liver transplantation for hepatic-based metabolic disorders. *Transpl Int* 1995; 8(1):41-44.
91. Davern TJ, Scharschmidt BF. Gene therapy for liver disease. *Dig Dis* 1998; 16(1):23-37.
92. O'Grady JG, Schalm SW, Williams R. Acute liver failure: redefining the syndromes. *Lancet* 1993; 342(8866):273-275.
93. Bernuau J, Benhamou JP. Classifying acute liver failure. *Lancet* 1993; 342(8866):252-253.
94. Makowka L, Chapman FA, Cramer DV, Qian SG, Sun H, Starzl TE. Platelet-activating factor and hyperacute rejection. The effect of a platelet-activating factor antagonist, SRI 63-441, on rejection of xenografts and allografts in sensitized hosts. *Transplantation* 1990; 50(3):359-365.
95. Lee WM. Acute liver failure. *N Engl J Med* 1993; 329(25):1862-1872.
96. Bismuth H, Samuel D, Castaing D, Adam R, Saliba F, Johann M, Azoulay D et al. Orthotopic liver transplantation in fulminant and subfulminant hepatitis. The Paul Brousse experience. *Ann Surg* 1995; 222(2):109-119.
97. Letoublon C, Guignier M, Barnoud D, Magne JL, Martin-Barbaz F, Zarski JP, Faure H et al. Heterotopic hepatic transplantation in fulminating hepatitis. *Chirurgie* 1989; 115(1):30-35.
98. Stampfl DA, Munoz SJ, Moritz MJ, Rubin R, Armenti VT, Jarrell BE, Maddrey WC. Heterotopic liver transplantation for fulminant Wilson's disease. *Gastroenterology* 1990; 99(6):1834-1836.
99. Chenard-Neu MP, Boudjema K, Bernuau J, Degott C, Belghiti J, Cherqui D, Costes V et al. Auxiliary liver transplantation: regeneration of the native liver and outcome in 30 patients with fulminant hepatic failure--a multicenter European study. *Hepatology* 1996; 23(5):1119-1127.
100. Erhard J, Lange R, Giebler R, Rauen U, de Groot H, Eigler FW. Auxiliary liver transplantation in urgent and emergency indications. *Langenbecks Arch Chir Suppl Kongressbd* 1996; 113:422-424.
101. Bismuth H, Samuel D, Castaing D, Williams R, Pereira SP. Liver transplantation in Europe for patients with acute liver failure. *Semin Liver Dis* 1996; 16(4):415-425.
102. Hellinger A, Rauen U, de Groot H, Erhard J. Auxiliary liver transplantation for acute liver failure after intake of 3,4-methylenedioxymethamphetamine ("Ecstasy"). *Dtsch Med Wochenschr* 1997; 122(22):716-720.
103. Roll C, Ballauff A, Lange R, Erhard J. Heterotopic auxiliary liver transplantation in a 3-year-old boy with acute liver failure and aplastic anemia. *Transplantation* 1997; 64 (4):658-660.
104. Pereira SP, McCarthy M, Ellis AJ, Wendon J, Portmann B, Rela M, Heaton N et al. Auxiliary partial orthotopic liver transplantation for acute liver failure. *J Hepatol* 1997; 26(5):1010-1017.
105. Sudan DL, Shaw BWJ, Fox IJ, Langnas AN. Long-term follow-up of auxiliary orthotopic liver transplantation for the treatment of fulminant

- hepatic failure. *Surgery* 1997; 122 (4):771-777.
106. Rosenthal P, Roberts JP, Ascher NL, Emond JC. Auxiliary liver transplant in fulminant failure. *Pediatrics* 1997; 100(2):E10.
107. van Hoek B, de Boer J, Boudjema K, Williams R, Corsmit O, Terpstra OT. Auxiliary versus orthotopic liver transplantation for acute liver failure. EURALT Study Group. European Auxiliary Liver Transplant Registry. *J Hepatol* 1999; 30(4):699-705.
108. Fischer L, Sterneck M, Rogiers X. Liver transplantation for acute liver failure. *Eur J Gastroenterol Hepatol* 1999; 11(9):985-990.
109. Maille F, Duvoux C, Cherqui D, Radier C, Zafrani ES, Dhumeaux D. Auxiliary hepatic transplantation in iproniazid-induced subfulminant hepatitis. Should iproniazid still be sold in France? *Gastroenterol Clin Biol* 1999; 23(10):1083-1085.
110. Erhard J, Lange R, Giebler R, Rauen U, de Groot H, Eigler FW. Arterialization of the portal vein in orthotopic and auxiliary liver transplantation. A report of three cases. *Transplantation* 1995; 60(8):877-879.
111. Erhard J, Lange R, Rauen U, Scherer R, Friedrich J, Pietsch M, de Groot H et al. Auxiliary liver transplantation with arterialization of the portal vein for acute hepatic failure. *Transpl Int* 1998; 11(4):266-271.
112. Shaw BWJ. Auxiliary liver transplantation for acute liver failure. *Liver Transpl Surg* 1995; 1(3):194-200.
113. Hanau C, Munoz SJ, Rubin R. Histopathological heterogeneity in fulminant hepatic failure. *Hepatology* 1995; 21(2):345-351.
114. Ringe B, Lubbe N, Kuse E, Frei U, Pichlmayr R. Total hepatectomy and liver transplantation as two-stage procedure. *Ann Surg* 1993; 218(1):3-9.
115. Rozga J, Podesta L, LePage E, Hoffman A, Morsiani E, Sher L, Woolf GM et al. Control of cerebral oedema by total hepatectomy and extracorporeal liver support in fulminant hepatic failure. *Lancet* 1993; 342(8876):898-899.
116. Calne RY, White HJ, Herbertson BM, Millard PR, Davis DR, Salaman JR, Samuel JR. Pig-to-baboon liver xenografts. *Lancet* 1968; 1(7553):1176-1178.
117. Calne RY, Davis DR, Pena JR, Balner H, Vries Md, Herbertson BM, Millard PR et al. Hepatic allografts and xenografts in primates. *Lancet* 1970; 1(7638):103-106.
118. Calne RY, White HJ, Yoffa DE, Binns RM, Maginn RR, Herbertson RM, Millard PR et al. Prolonged survival of liver transplants in the pig. *Br Med J* 1967; 4(580):645-648.
119. Qian S, Fung JJ, Demetris AJ, Starzl TE. Allogeneic orthotopic liver transplantation in mice: a preliminary study of rejection across well-defined MHC barriers. *Transplant Proc* 1991; 23(1 Pt 1):705-706.
120. Lee S, Edgington TS. Liver transplantation in the rat. *Surg Forum* 1966; 17:220-222.
121. Lee S, Edgington TS. Heterotopic liver transplantation utilizing inbred rat strains. I. Characterization of allogeneic graft rejection and the effects of biliary obstruction and portal vein circulation on liver regeneration. *Am J Pathol* 1968; 52(3):649-669.

## 20 CHAPTER 1 General Introduction

122. Kort WJ, Wolff ED, Eastham WN. Heterotopic auxiliary liver transplantation in rats. Use of the infrahepatic vena cava as the efferent vessel. *Transplantation* 1971; 12(6):415-420.
123. Hess F, Jerusalem C, van der Heyde MN. Advantages of auxiliary liver homotransplantation in rats. *Arch Surg* 1972; 104:76-80.
124. Hess F, Willemen A, Jerusalem C. Survival of auxiliary rat liver grafts with decreased portal blood flow. *Eur Surg Res* 1978; 10(6):444-455.
125. Hess F, Willemen A, Jerusalem C. Auxiliary liver transplantation in the rat, influence of the condition of the recipient's liver on the fate of the graft. *Eur Surg Res* 1977; 9(4):270-279.
126. Müller G. A simple technique for heterotopic auxiliary liver transplantation in the rat. *Transplantation* 1983; 36(2):221-222.
127. Marni A, Ferrero ME. Heterotopic liver grafting in the rat. A simplified method using cuff techniques. *Transplantation* 1985; 39(3):329-331.
128. Lee S, D'Silva M, Glassford E, Dennis R, Moossa AR. Arterialization of liver transplants in rats: I. Application of the tissue perfusion monitor in hemodynamic studies. *Microsurgery* 1989; 10(1):15-20.
129. Glassford E, D'Silva M, Ghorab H, Bai S, Lee SU, Moossa AR, Lee S. Arterialization of the liver. II. Systemic pressure gradients in rats following variously sized arteriovenous fistulae. *Microsurgery* 1990; 11(2):177-183.
130. D'Silva M, Pirenne J, Glassford E, Mayer D, Bai S, Gittes RF, Lee S. Arterialization of the liver. III. Influence of systemic and portal pressure gradients following heterotopic partial liver transplantation. *Microsurgery* 1990; 11(2):184-187.
131. Lee S, Scott MH. Six models of heterotopic rat liver transplantation: introducing a reverse circulation model. *Microsurgery* 1986; 7(2):91-94.
132. Schleimer K, Lange R, Rauen U, Erhard J. Auxiliary liver transplantation in acute liver failure in the rat -- an illustrated description of a new surgical approach. *Langenbecks Arch Surg* 1999; 384(2):204-208.
133. Schleimer K, Lange R, Rauen U, Nowak B, Brandt-Mainz K, de Groot H, Erhard J. Auxiliary rat liver transplantation with portal vein arterialization in acute hepatic failure. *Transplantation* 2000; 70 (1):73-78.
134. Luo M, Yu Y, Tang Z. Auxiliary partial orthotopic liver transplantation in rats. *Zhonghua Wai Ke Za Zhi* 1997; 35(9):561-563.
135. Nishio K, Nakajima Y, Kanehiro H, Hisanaga M, Kin T, Ohyama T, Ohashi K et al. Development of auxiliary partial orthotopic liver transplantation model in the rat. *Transplant Proc* 1998; 30(7):3237-3238.
136. Ikegami T, Nishizaki T, Yanaga K, Ohta R, Hiroshige S, Sugimachi K. A novel auxiliary partial orthotopic liver transplantation model in rats. *Eur Surg Res* 2000; 32 (5):267-273.
137. Capizzi FD, Alonzo A, Abo-Rahmy E, Prodi G. The survival of an auxiliary transplanted liver in the inbred rat. *J Chir* 1977; 114(1-2):107-110.
138. Hong HQ, Asahara T, Ito H, Watanabe H, Kimura A, Urushihara T, Marubayashi S et al. Study of portal arterialization with auxiliary liver in rats. *Hiroshima J Med Sci* 1991; 40(1):29-33.
139. Zelder O, Paidlick A, Bode C, Bode JC, Jerusalem CR, Hamelmann H. Changes in the activity of enzymes with various cell location following

- heterotopic auxiliary liver transplantation in the rat. *Langenbecks Arch Chir* 1974; Suppl:337-341.
140. Thermann M, Zelder O, Hess F, Jostarndt L, Jerusalem CR, Hamelmann H. Studies on the oxygen supply of heterotopic auxiliary liver grafts in rats. *Eur Surg Res* 1975; 7(3):156-163.
  141. Fisher ER, Fisher B. Histopathologic and ultrastructural study of allogeneic hepatic transplantation in isogenic rats. *Lab Invest* 1970; 23(3):318-326.
  142. Andrzejewski W, Orkisz S, Bartel H. Electron microscopic study of the rat liver after its orthotopic and auxiliary syngeneic transplantation. *Exp Pathol* 1985; 27(1):49-54.
  143. Jansen PL, Hess F, Peters WH, Koenders E, Jerusalem C, Corstens FH. Auxiliary liver transplantation in jaundiced rats with UDP-glucuronyltransferase deficiency and defective hepatobiliary transport. *J Hepatol* 1989; 8(2):192-200.
  144. Asonuma K, Gilbert JC, Stein JE, Takeda T, Vacanti JP. Quantitation of transplanted hepatic mass necessary to cure the Gunn rat model of hyperbilirubinemia. *J Pediatr Surg* 1992; 27(3):298-301.
  145. Wang YN, Luk JM, Chung S, Fan ST. In situ gene transfer into rat auxiliary liver transplant. *Transplantation* 1997; 64(11):1537-1541.
  146. Kobayashi E, Yoshida Y, Nozawa M, Hishikawa S, Yamanaka T, Miyata M, Fujimura A. Auxiliary heterotopic liver transplantation in the rat: a simplified model using cuff technique and application for congenitally hyperbilirubimemic Gunn rat. *Microsurgery* 1998; 18(2):97-102.
  147. Gugenheim J, Houssin D, Tamisier D, Franco D, Martin E, Lang P, Bismuth H. Spontaneous long-term survival of liver allografts in inbred rats: influence of the hepatectomy of the recipient's own liver. *Transplantation* 1981; 32(5):445-450.
  148. Icard P, Sawyer GJ, Houssin D, Fabre JW. Marked differences between orthotopic and heterotopic auxiliary liver allografts in the induction of class II MHC antigens on hepatocytes. *Transplantation* 1990; 49(5):1005-1007.
  149. Astarcioglu I, Gugenheim J, Gigou M, Amorosa L, Fabiani B, Reynes M, Bismuth H. Immunosuppressive properties of auxiliary liver allografts into sensitized rats. *Transplantation* 1990; 49(6):1186-1188.
  150. Kamada N, Muller GH, Katami M, Tamaki T, Baguerizo A, Shinomiya T. Sensitization of rats for rejection of heart allografts by heterotopic auxiliary liver grafting or administration of liver cell suspensions. *Transplantation* 1985; 40(1):106-108.
  151. Kamada N, Yamaguchi A, Yoshimura S, Sumimoto R, Teramoto K, Baquerizo A. Mechanisms of immunosuppression by serum from liver grafted rats: evidence for functional clonal deletion in vivo. *Transplant Proc* 1990; 22(4):1971-1972.
  152. Bestian JM, Janin A, Zenner L, Pruvot FR, Zelus D, Courtade A, Dessaint JP et al. Allogenic microchimerism following auxiliary heterotopic liver transplantation in rat and swine. *Transplant Proc* 1995; 27(2):1675.
  153. Chandler JG, Lee S, Krubel R, Rosen H, Nakaji NT, Orloff MJ. The inter-liver competition and portal blood in regeneration of auxiliary liver transplants. *Surg Forum* 1971; 22:341-343.

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154. Broelsch CE, Lee S, Charters AC, Chandler JG, Grambort DE, Orloff MJ. Regeneration of liver isografts transplantation in continuity with splanchnic organs. *Surg Forum* 1974; 25(0):394-396.
155. Lorente L, Arias J, Aller MA, Ispizua JI, Rodriguez J, Duran H. Heterotopic auxiliary liver transplantation with portal flow. Gradual development of the collateral circulation. *HPB Surg* 1990; 2 (4):281-291.
156. Yu W, Wan X, Wright JRJ, Coddington D, Bitter-Suermann H. The effect of intrahepatic islet isografts on liver integrity following heterotopic auxiliary liver is transplantation in rats. *Transplant Proc* 1992; 24 (6):2920-2921.
157. Yamadera H, Yamaguchi A, Shinomiya T, Matsuda M, Sakurai K, Kimura H. Immunologic enhancement mediated by vascularized auxiliary liver transplantation in the rat. *Transplant Proc* 1994; 26(4):2242.
158. Yu W, Wan X, Wright JRJ, Coddington D, Bitter-Suermann H. Heterotopic liver transplantation in rats: effect of intrahepatic islet isografts and split portal blood flow on liver integrity after auxiliary liver is transplantation. *Surgery* 1994; 115 (1):108-117.
159. Sauvanet A, Yang S, Bernuau D, Beyne P, Denninger MH, Farges O, Lebrech D et al. Auxiliary liver transplantation: how to improve regeneration of the native liver by surgery. *Transpl Int* 1999; 12 (6):433-438.
160. Yoo CH, Wang Y, Ha J, Mao L, Kim S, Tarin T, Wolf P et al. Long-term fate of heterotopic liver transplants in rats with portal vein inflow only. *Microsurgery* 1999; 19(2):78-82.
161. Starzl TE, Marchioro TL, Huntley RT. Experimental and clinical homotransplantation of the liver. *Amer NY Acad Sci* 1964; 120:739-765.
162. Marchioro TL, Porter KA, Dickins TC, Faris TD, Starzl TE. Physiologic requirements for auxiliary liver transplantation. *Surg Gynecol Obstet* 1965; 121:17-31.
163. Schalm L, Bax HR, Mansens J. Atrophy of the liver after occlusion of the bile ducts or portal vein and compensatory hypertrophy of the unoccluded portion and its clinical importance. *Gastroenterology* 1955; 31:131-155.
164. Schalm L. Heterotopic auxiliary liver transplantation. I. A physiological concept of functional competition. *Arch Chir Neerl* 1966; 18(4):283-284.
165. van der Heyde MN, Schalm L, Vink M. The role of functional competition in auxiliary liver transplantation. *Transplantation* 1967; 5:78-80.
166. Heyde MN, Schalm L. Auxiliary liver-graft without portal blood. Experimental autotransplantation of left liver lobes. *Br J Surg* 1968; 55(2):114-118.
167. Halgrimson CG, Marchioro TL, Faris TD, Porter KA, Peters GN, Starzl TE. Auxiliary liver transplantation: effect of host portacaval shunt. Experimental and clinical observations. *Arch Surg* 1966; 93(1):107-118.
168. Marchioro TL, Porter KA, Brown BI, Faris TD, Herrmann TJ, Sudweeks A, Starzl TE. The specific influence of nonhepatic splanchnic venous blood flow on the liver. *Surg Forum* 1965; 16:280-282.
169. Marchioro TL, Porter KA, Brown BI, Otte JB, Starzl TE. The effect of partial portacaval transposition on the canine liver. *Surgery* 1967; 61(5):723-732.

170. Starzl TE, Francavilla A, Halgrimson CG, Francavilla FR, Porter KA, Brown TH, Putnam CW. The origin, hormonal nature, and action of hepatotrophic substances in portal venous blood. *Surg Gynecol Obstet* 1973; 137(2):179-199.
171. Starzl TE, Porter KA, Kashiwagi N. Portal hepatotrophic factors, diabetes mellitus and acute liver atrophy, hypertrophy and regeneration. *Surg Gynecol Obstet* 1975; 141(6):843-858.
172. Starzl TE, Porter KA, Putnam CW. Intraportal insulin protects from the liver injury of portacaval shunt in dogs. *Lancet* 1975; 2(7947):1241-1242.
173. Starzl TE, Watanabe K, Porter KA, Putnam CW. Effects of insulin, glucagon, and insulinglucagon infusions on liver morphology and cell division after complete portacaval shunt in dogs. *Lancet* 1976; 1(7964):821-825.
174. Wexler MJ, Farkouh EF, Farrer PA, Slapak M, MacLean LD. Auxiliary liver transplantation: a successful model without portal blood or portacaval shunt. *Ann Surg* 1972; 175 (3):357-374.
175. Sgro JC, Charters C, Chandler JG, Grambort DE, Orloff MJ. Site of origin of the hepatotrophic portal blood factor involved in liver regeneration. *Surg Forum* 1973; 24:377-379.
176. Hagiwara PF, Griffen WOJ. Homologous heterotopic porcine liver transplantation. *J Surg Res* 1975; 19(2):89-97.
177. Francavilla A, Hagiya M, Porter KA, Polimeno L, Ihara I, Starzl TE. Augmenter of liver regeneration: its place in the universe of hepatic growth factors. *Hepatology* 1994; 20(3):747-757.
178. Whittemore AD, Voorhees ABJ, Price JBJ. Hepatic blood flow and pancreatic hormones as modifiers of hepatic regeneration. *Surg Forum* 1976; 27(62):363-365.

## **CHAPTER 2**

### **POST-TRANSPLANT EVALUATION OF HEPATIC VIABILITY IN THE RAT MODEL OF HETEROTOPIC AUXILIARY LIVER TRANSPLANTATION**



## **26 2.1. Post-HALTx Evaluation of Graft Viability**

### **2.1. Evaluation of Graft Viability in Heterotopic Auxiliary Liver Transplantation in the Rat**

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#### **SUMMARY**

In the rat model of heterotopic auxiliary liver transplantation, the coexistence of the engrafted liver and the recipient's native liver makes it difficult to evaluate the post-transplant graft viability. In this study, auxiliary liver transplantation was performed in Wistar rats, in which the recipient's native liver was handicapped with a 68% partial hepatectomy and a common bile duct ligation. Serum biochemistry of the liver was analysed and compared with that of the selected control group. The surgical handicap of the liver showed severe damaging effects: the handicapped native livers appeared atrophic at autopsy and no long-term animal survival could be achieved without an auxiliary liver transplantation. As the engrafted liver corrected the cholestasis of the handicapped native liver, significant differences of serum biochemistry were found between the transplanted group and the control group: for bilirubin concentration and gamma glutamyl transferase activity from post-operative day 3 to 28 ( $p < 0.05$ ); alkaline phosphatase on day 3, 7, 14 and 28 ( $p < 0.05$ ); alanine aminotransferase activity on day 3 and 14 ( $p < 0.05$ ) and aspartate aminotransferase activity on day 14 ( $p < 0.05$ ). The efficiency to induce hepatic failure and to hamper its regeneration capacity in the native liver makes animal survival and liver biology as reliable parameters to evaluate the post-transplant graft viability in this rat model.

CHAPTER 2 Post-HALTx Evaluation of Hepatic Viability **27**

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## **INTRODUCTION**

Although the first attempt of liver transplantation in human was a heterotopic auxiliary liver transplantation (HALTx) (1), orthotopic liver transplantation (OLTx) has long been accepted as the standard treatment for end-stage liver diseases. This is mainly due to the poor results of clinical HALTx. However, HALTx has always been theoretically attractive with advantages over OLTx in some specific cases (2-4). HALTx avoids the surgical trauma of more extensive liver dissection and the an-hepatic phase needed for the transplantation. For the patients with end-stage chronic liver disease at high risk for OLTx, HALTx certainly represents a life-saving alternative (5). In addition, a heterotopic auxiliary liver may correct some inborn errors of metabolism with enzymatic deficiency without removing the otherwise normal liver (6). Above all, HALTx is particularly useful for treating patients with fulminant and subfulminant hepatic failure in whom the function of the native liver may still recover. Following a successful recuperation of the native liver, the auxiliary liver can then be removed, thus avoiding the life-long use of immunosuppressive drugs and its potential complications. This has been proven by encouraging clinical results from different organ-transplantation centres in recent years (7-10).

However, the engrafted liver and the recipient's native liver may behave competitively to each other. As a consequence, the atrophy of the engrafted liver is the major cause of the failure in HALTx. Even though this "inter-liver competition" phenomenon was recognized as early as at the beginning of clinical and experimental HALTx (11, 12), its mechanisms still remain incompletely understood. Amongst other experimental protocols, the rat HALTx is an ideal model to perform studies related to this more exceptional but important clinical situation (13, 14).

As the engrafted liver coexists with the recipient's native liver in this rat model, how to evaluate the post-transplant graft viability was an open question. In the current study, the function of the native liver was surgically handicapped. The most commonly used serum biochemical parameters were tested as possible indicators to evaluate the post-transplant evolution of the engrafted liver.

## **MATERIALS AND METHODS**

### **Experimental Animals**

All animals of this study received care in accordance with institutional, national and European guidelines. Male out bred Wistar rats (B & K

Universal Limited, England) weighing 240-320g were used. The rats had free access to food and water before surgery.

### Experimental Groups (*Table 1*)

HALTx group ( $n = 12$ ): HALTx was performed in all rats of this group, in which the recipient's native liver was handicapped with both a 68% partial hepatectomy (68% PH) and a common bile duct ligation (CBDL).

In order to evaluate post-transplant liver biology, a proper control group with long-term animal survival was selected from four experimental models:

- Group 1 ( $n = 10$ ). The liver handicapped in the same way as for the recipient's native liver in the HALTx group: a portacaval shunt (PCS), a 68% PH and a CBDL.
- Group 2 ( $n = 10$ ). A PCS and a 68% PH.
- Group 3 ( $n = 10$ ). A PCS and a CBDL.
- Group 4 ( $n = 10$ ). A 68% PH and a CBDL.

### Surgical Techniques

All operative procedures were carried out under ether anaesthesia with clean but non-sterile instruments, and x 12.5 magnification. During the operation, the body temperature of the rat was maintained at approximately 36°C by using heating pads.

- (a) *HALTx*: The HALTx was performed according to the technique described elsewhere (15). Briefly, 32% of the donor liver was used for transplantation after perfusion with cold (4° C) heparinized physiological saline solution via the portal vein. The native liver of the recipient rat was handicapped with both a 68% PH and a CBDL, while the hepatic artery remained intact. Following a right nephrectomy, the engrafted liver was implanted in the right paravertebral gutter under the native liver of the recipient. The cuff technique was used for the anastomoses of both the vena cava and the portal vein. The engrafted liver received its blood supply from the portal vein of the recipient. The venous drainage was made through the right renal vein of the recipient to the vena cava. Restoration of the bile drainage was done by insertion of the intubated common bile duct of the donor liver into the recipient duodenum. No re-arterialisation of the engrafted liver was attempted in this study. The graft fixation was performed to avoid twisting and kinking of the donor vena cava as previously reported (15) (See photo illustration at the end of this section).

### 30 2.1. Post-HALTx Evaluation of Graft Viability

During the operation, cold ischemia time of the engrafted livers (starting from saline perfusion via the portal vein and ending at portal re-vascularization of the engrafted liver) was kept between 72 and 79 minutes.

- (b) *PCS*: The method described by Lee (16) was modified with the cuff technique to perform a PCS. After a left nephrectomy, the left renal vein was freed up to the vena cava, which was then cross-clamped. An incision was made into the anterior wall of the stump of the left renal vein. The portal vein was divided from the pyloric vein to the liver hilus, tied, transected and cuffed with a 0.4 cm length polyethylene tube. The cuffed portal vein was then inserted into the left renal vein and secured with a 5-0 ligature.
- (c) *68%PH*: The median lobe and the left lobe of the liver were ligated and then excised according to the standard method introduced by Higgins and Anderson (17).
- (d) *CBDL*: The common bile duct was double ligated with non-absorbable sutures and the segment in between was then removed.

#### Post-operative Care and Follow-up

All rats were given intra-muscular injection of 10 mg Na Cefazolin immediately after surgery and this was repeated daily for three days. No immunosuppressive agent was administered to the rats. During the following twenty-four hours, rats were given 5% glucose to drink, after which the pre-operative diet of food pellets and water was resumed.

All rats were observed daily and survival time of each rat was recorded.

For the serum biochemical analysis, a blood sample of 0.4 - 0.5 ml was collected from the rat's tail vein pre-operatively, on the post-operative day 3, 7, 14, 21, 28, 56 and 84. Serum bilirubin concentration (TBil), gamma glutamyl transferase activity (GGT), alkaline phosphatase (ALP), alanine aminotransferase activity (ALAT), aspartate aminotransferase activity (ASAT), lactate dehydrogenase (LDH) and total protein (TP) were determined by using BM / Hitachi System 747 (Boehringer Mannheim GmbH, Diagnostica). The reference intervals (range) used in this laboratory were for serum TBil 0.00 – 0.20 ml/dl, GGT 0 U/l, ALP 58.18 – 328.14 U/l, ALAT 17.38 – 59.55 U/l and ASAT 51.73 – 107.89 U/l.

An autopsy was performed whenever a rat died. The long-term survivors were sacrificed at the end of the 3-month observation period. The livers were excised, cleaned of gross adhesions, and fixed in 10% formalin. Four-micron thick sections were stained with hematoxylin and eosin (HE) or trichrome

and examined under light microscopy.

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**Table 1.** Experimental groups and post-operative animal survival

Experimental groups	Survival time (days, mean ± SD)			
Control groups				
G1 ( <i>n</i> = 10): PCS + 68% PH + CBDL	0.5 ± 0.3			
G2 ( <i>n</i> = 10): PCS + 68% PH	0.8 ± 0.6			
	Survival rates			
	1 week	1 month	2 months	3 months
Control groups				
G3 ( <i>n</i> = 10): PCS + CBDL	100%	80%	20%	20%
G4 ( <i>n</i> = 10): 68% PH + CBDL	100%	100%	80%	50%
HALTx group ( <i>n</i> = 12)	100%	66%	50%	50%

Note: PCS, portacaval shunt; 68% PH, 68% partial hepatectomy; CBDL, common bile duct ligation; HALTx, heterotopic auxiliary liver transplantation.

### Statistical Analysis

The data were expressed as the mean  $\pm$  standard deviation (SD). The distribution of the data was checked by Kolmogorov test. The Student's *t*-test was applied for the normally distributed data, while the not-normally-distributed data were evaluated by the Mann-Whitney rank sum test. *P* values equal to or less than 0.05 were considered statistically significant.

## RESULTS

### Post-operative Animal Survival (Table 1)

*HALTx group.* All 12 rats survived the first post-transplant week. The 1-month and the 3-month survival rate were 66% and 50%, respectively. The causes of post-operative deaths were mainly due to liver abscess (3/6), cholangitis (1/6), peritonitis (1/6) and rejection (1/6). The biochemical data of the six long-term survival rats were compared with that of the control group.

*Control groups.* Despite the comparable vascular cross-clamping time, all rats in the group 1 died of liver failure within 24 hours after surgery and in the group 2 reduced liver handicap could not significantly improve animal survival. In the group 3, two out of ten rats died of infectious complications (cholangitis and peritonitis) during the 1<sup>st</sup> post-operative month and six rats that died between the 1<sup>st</sup> and the 2<sup>nd</sup> month showed signs of biliary congestion with ascites. The 2-month survival rate was only 20% in this group. All 10 rats of the group 4 survived the first post-operative month. One rat died of pneumonia later on and cholangitis caused the other 4 animal losses. The 3-month survival rate of this group was 50%.

### **Serum Biochemical Analysis**

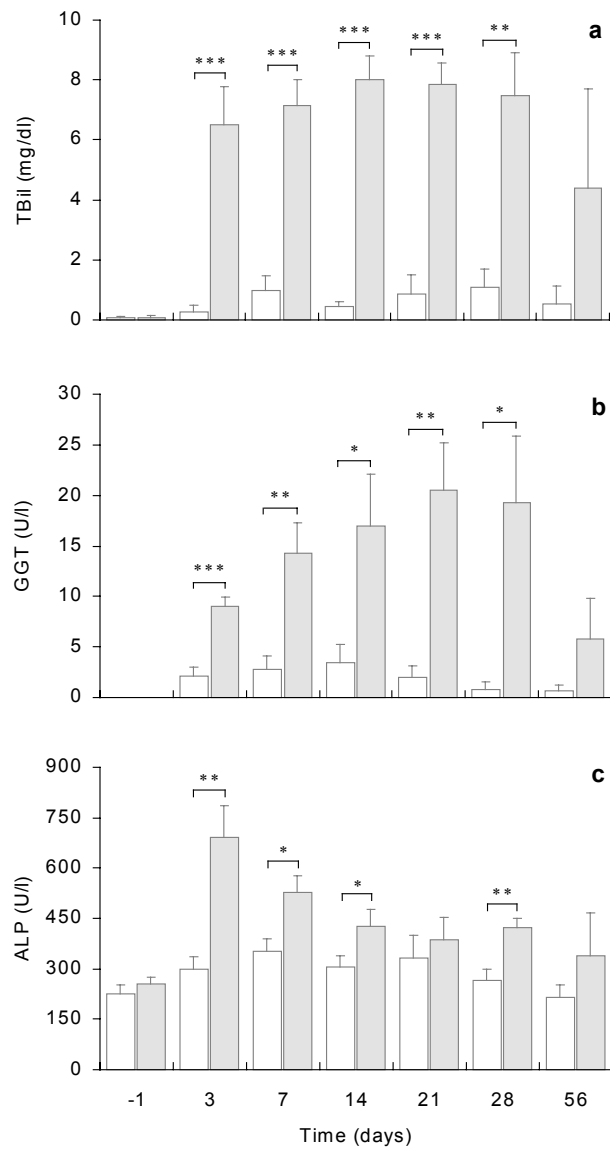
Since none of the rats in the group 1 and 2 survived longer than two days and the 2-month survival rate of the group 3 was as low as 20%, only the data of the group 4 were available to compare with those of the HALTx group.

The most commonly used serum biochemical parameters (TBil, GGT, ALP, ALAT, ASAT, LDH and TP) were evaluated as possible indicators of liver viability for this rat model. No significant differences between the two groups were found for LDH and TP.

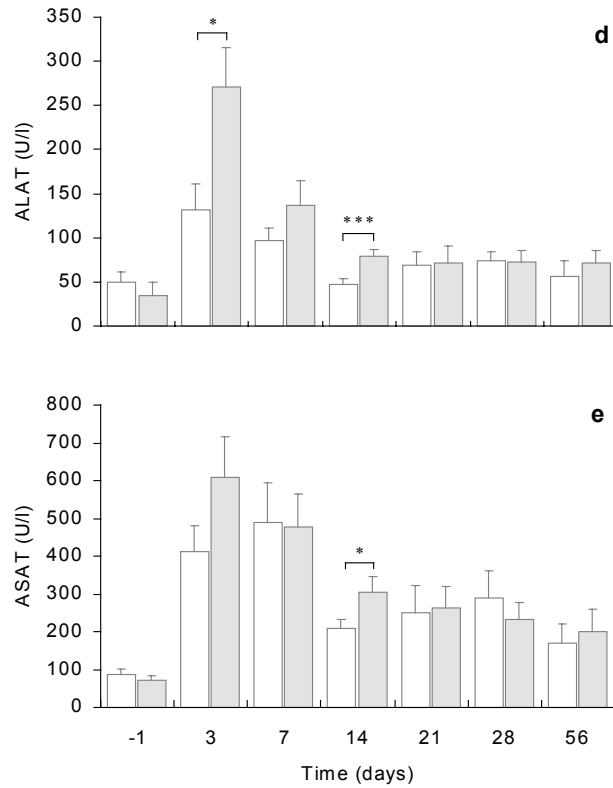
Serum TBil, GGT, ALP, ALAT and ASAT levels were elevated in the rats of two groups after surgery. However, the rats in the HALTx group had significantly lower values of TBil than that in the group 4 on day 3 ( $p < 0.001$ ), 7 ( $p < 0.001$ ), 14 ( $p < 0.001$ ), 21 ( $p < 0.001$ ) and 28 ( $p < 0.005$ ) (*Figure 1a*). The same profile appeared for GGT on day 3 ( $p < 0.001$ ), 7 ( $p < 0.01$ ), 14 ( $p < 0.05$ ), 21 ( $p < 0.005$ ) and 28 ( $p < 0.05$ ) (*Figure 1b*). Serum ALP also showed a pronounced difference between the two groups on day 3 ( $p < 0.01$ ), 7 ( $p < 0.05$ ), 14 ( $p < 0.05$ ) and 28 ( $p < 0.01$ ) (*Figure 1c*). In the HALTx group, serum ALAT was significantly lower than those of the group 4 on day 3 ( $p < 0.05$ ) and 14 ( $p < 0.001$ ) (*Figure 1d*) while ASAT differed significantly between the two groups on day 14 ( $p < 0.05$ ) (*Figure 1e*).



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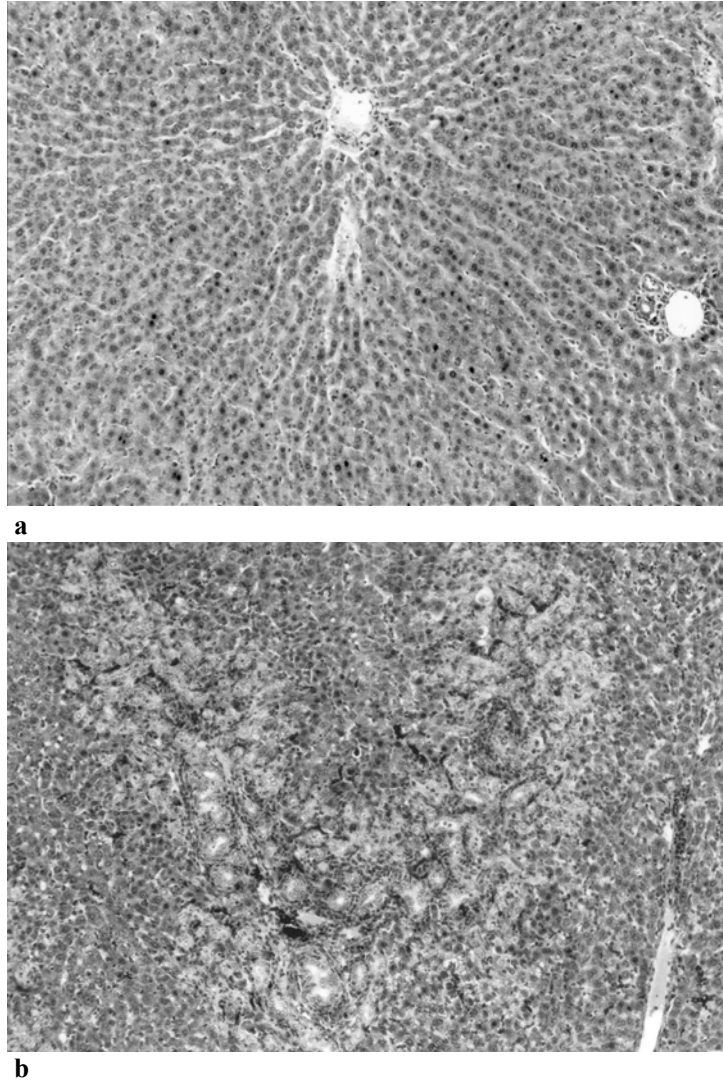


**Figure 1. Continued**



**Figure 1.** Comparison of serum biochemical tests (mean  $\pm$  SD) between the HALTx group (□) and the control group 4 (■): (a) bilirubin concentration (TBil), (b) gamma-glutamyl transferase activity (GGT), (c) alkaline phosphatase (ALP), (d) alanine aminotransferase activity (ALAT) and (e) aspartate aminotransferase activity (ASAT). Asterisk indicates statistically significant differences between the two groups. \*  $p \leq 0.05$ , \*\*  $p < 0.01$  and \*\*\*  $p < 0.001$ .

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**Figure 2.** Histological findings of 3-month survival rats: **(a)** HALTx group: The engrafted liver preserved its normal lobular architecture without major histopathological alterations (original magnification x 125). **(b)** Control group 4: The liver completely lost its general lobular architecture with the proliferation of the cholangioles lined by swollen cuboidal epithelium (original magnification x 125)

### Autopsy and Histology

Three months after the transplantation, the 6 long-term survivors of the HALTx group were sacrificed for histological analysis. At autopsy, the engrafted livers of these rats appeared normal in size and colour and the cuffed portal vein and vena cava anastomoses remained patent. These engrafted livers revealed a normal lobular architecture without major histopathological alterations (*Figure 2a*), whereas the general lobular architecture of the handicapped native livers was lost and replaced by ductular metaplasia in the portal tracts accompanied by fibrosis.

In the group 4, the 5 long-term survival rats were also sacrificed 3 months after the surgery. A bile duct cyst of 1-3 cm diameter was found in the hepatic hilus of all animals. The livers were macronodular with disappearance of the general lobular architecture characterised by the proliferation of the cholangioles lined by swollen cuboidal epithelium (*Figure 2b*). There was no substantial difference with the native liver of the recipient in HALTx group.

### DISCUSSION

Since the technique of HALTx in rat was first described in 1966 (13), this rat model has been used in experimental researches of various subjects such as liver regeneration, hepatotrophic factors, allogenic microchimerism and graft rejection (18-20). As the two-liver state in this model makes it difficult to evaluate the viability of the engrafted liver after the transplantation, a possible approach is to handicap the recipient's native liver to such a degree that it will eventually cease its function. In this study, liver failure was induced surgically by means of handicapping the native liver with both a 68% PH and a CBDL. Corresponding to a previous observation (14), it appeared that this handicap could damage the organ so severely that not only the liver failure was assured causing animal death, but also the hepatic regeneration capacity was completely impaired leading to atrophy in long-term survivors. Therefore, the post-transplant animal survival relies primarily upon the function of the engrafted liver, and the serum biochemistry of the liver reflects the viability of that engrafted organ despite the presence of the recipient's native liver.

In evaluating the post-transplant graft viability, the combination of a PCS, a 68% PH and a CBDL should be the most appropriate control since the native liver in the HALTx group was handicapped in the same way. However, without an auxiliary liver, this approach was always lethal. A reduced handicap with a PCS and a 68% PH also failed to significantly improve

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animal survival. Although the rats subjected to a PCS plus a CBDL did survive longer, a 2-month survival rate of 20% could not provide sufficient data for a long-term comparative study. Most rats of this group died between the 1<sup>st</sup> and 2<sup>nd</sup> post-operative month with signs of cholestasis and ascites, which indicated that this handicap could induce a subacute liver failure. In the absence of long-term survival, no real control or comparison could be done and even less traumatic surgical procedures had to be tested. A 68% PH plus a CBDL without a PCS eventually appeared to be the only viable control of this model, which facilitated a long-term comparison of serum biochemistry with the HALTx group.

Along with autopsy and pathological examination, post-operative animal survival is indeed a commonly used parameter to evaluate the viability of the engrafted liver after HALTx. However, survival rate does not allow to evaluate, for instance, the degree of the liver injury caused by preservation and re-perfusion, and to appreciate the post-operative complications of the engrafted liver. Serum biochemistry of the liver is an un-invasive measurement, which can be used for systematic evaluation of graft viability by simple means. In contrary to previous publications (18, 21), in this study, LDH was not significantly different between the HALTx group and the control group. This could be due to the fact that LDH is an enzyme present in a wide variety of tissues including heart, skeletal muscle, kidney and brain in addition to the liver. Thus muscle trauma induced by the operative procedures can interfere and influence LDH value mainly in the early observation phase. Since TP measures different sorts of proteins, which may be affected positively or negatively by the experimental interventions, it lacks specificity for the liver function as shown in this study.

Serum TBil, GGT and ALP are the parameters directly linked to hepatobiliary cholestasis. In the present study, the serum levels of TBil, GGT and ALP increased drastically in the rats of the control group after a 68% PH and a CBDL, but not in the HALTx group despite the fact that the recipient's native liver was subjected to the same handicap. This demonstrated that the function of bile secretion was mainly taken over by the engrafted liver from the recipient's native liver, which was consequently less cholestatic. During the first post-operative month, the significantly lower levels of serum TBil, GGT and ALP in the transplanted rats were thus a sign of proper function of the engrafted liver. The elevation of serum ALAT and ASAT immediate after the operation indicated hepatocellular damage in both the transplanted and the control groups. The lower peak values and faster normalisation of both parameters in the rats of the HALTx group suggested that the engrafted liver reduced the deleterious effect of cholestasis on the native liver.

As the high success rates of the HALTx procedure can be achieved with a sole portal blood supply, the re-arterialisation of the engrafted liver has not been considered as a necessity for this rat model (14, 22, 23). However, using OLTx model, the recent studies have demonstrated the beneficial effects of the re-arterialisation on improving micro-vascular perfusion (24), modifying the expression of class I and II MHC antigens (25, 26), maintaining an adequate oxygen supply and tissue ATP (27), reducing the post-transplant biliary complications (28-30) and increasing animal survival rate (31, 32). Whether equal mechanisms may be implicated in the HALTx has not yet been addressed. As the interaction between the engrafted and the native liver added an important element to this specific rat model, this question indeed warrants further attention.

In conclusion, the data of present study show that in this rat model of HALTx, handicap of the native liver with a 68% PH and a CBDL can efficiently induce liver failure and hamper its regeneration capacity. The post-transplant animal survival and normalization of the liver biology reflect well the viability of the engrafted liver.

## REFERENCES

1. Starzl TE, Marchioso TL, Huntley RT. Experimental and clinical homotransplantation of the liver. *Am NY Acad Sci* 1964; 120: 739-765.
2. Starzl TE, Iwatsuki S, Van Thiel DH. Evolution of liver transplantation. *Hepatology* 1982; 2: 614-636.
3. Starzl TE, Demetris AJ, Van Thiel D. Liver transplantation. *N Engl J Med* 1989; 321-16: 1092-1099.
4. Terpstra OT, Reuvers CB, Schalm SW. Auxiliary heterotopic liver transplantation. *Transplantation* 1988; 45-6: 1003-1007.
5. Terpstra OT, Schalm SW, Weimar W, Willemse P, Baumgartner D, Groenland T, ten Kate F, Porte RJ, De Rave S, Reuvers CB, Stibbe J, Terpstra JL. Auxiliary partial liver transplantation for end-stage chronic liver disease. *N Engl J Med* 1988; 319: 1507-1511.
6. Madern GC, Terpstra OT, Sinaasappel M, Provoost AP, Rothuizen J, Molenaar JC. Heterotopic liver transplantation corrects the inborn error of hepatic metabolism in a dog model. *Transplant Proc* 1991; 23-1: 716-717.
7. Van Hoek B, Ringers J, Kroes AC, van Krieken JH, van Schelven WD, Masclee Ad, van Krieken-Hogenberk LG, Haak HR, Lamers CB, Terpstra OT. Temporary heterotopic auxiliary liver transplantation for fulminant hepatitis B. *J Hepatol* 1995; 23: 109-118.
8. Boudjema K, Cherqui D, Jaeck D, Chenard-Neu MP, Steib A, Freis G, Becmeur F, Brunot B, Simeoni U, Bellocq JP, Tempe JD, Wolf P, Cinqualbre J. Auxiliary liver transplantation for fulminant and subfulminant hepatic failure. *Transplantation* 1995; 59-2: 218-223.

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9. Chenard-Neu MP, Boudjema K, Bernuau J, Degott C, Belghiti J, Cherqui D, Costes V, Demergue J, Durand F, Erhard J, de Hemptinne B, Gubernatis G, Hadengue A, Kemnitz J, McCarthy M, Masschek H, Mentha G, Oldhafer K, Portmann B, Praet M, Ringers J, Rogiers X, Rubbia L, Schalm S, Bellocq JP. Auxiliary liver transplantation: regeneration of the native liver and outcome in 30 patients with fulminant hepatic failure – a multicenter European study. *Hepatology* 1996; 23-5: 1119-1127.
10. Moritz WJ, Jarrell BE, Armenti V, Radomski J, Carabasi RA, Zeitoun G, Columbus K, Rubin R, Munoz Smaddrey W. Heterotopic liver transplantation for fulminant hepatic failure -- a bridge to recovery. *Transplantation* 1990; 50: 524-526.
11. Marchioro TZ, Poster KA, Dickinson TC, Faris TD, Starzl TE. Physiologic requirements for auxiliary liver transplantation. *Surg Gynecol Obst* 1965; 121: 17-31.
12. Van der Heyde MN, Schalm L, Vink M. The role of functional competition in auxiliary liver transplant. *Transplantation* 1967; 5: 78-80.
13. Lee S, Edgington TS. Liver transplantation in the rat. *Surg Forum* 1966; 17: 220-222.
14. Hess F, Jerusalem C, Van der Heyde MN. Advantages of auxiliary liver homo-transplantation in rats. *Arch Surg* 1972; 104: 76-80.
15. Fan YD, Hesse UJ, Praet M, de Hemptinne B. Inbred PVG rats are not suitable for the model of heterotopic auxiliary liver transplantation (HALTx). *Euro Surg Res* 1996; 28-S1: 33.
16. Lee S, Fisher B. Portacaval shunt in the rat. *Surgery* 1961; 50: 668-672.
17. Higgins GM, Anderson RM. Experimental pathology of the liver: I. Restoration of the liver of the white rat following partial surgical removal. *Arch Pathol* 1931; 12: 186-202.
18. Lee S, Edgington TS. Heterotopic liver transplantation utilising inbred rat strains: characterisation of allogeneric graft rejection and the effect of biliary obstruction and portal vein circulation on liver regeneration. *Am J Pathol* 1968; 52: 649-669.
19. Yu WY, Wan XY, Wright JR, Coddington D, Bitter-Suermann H. Heterotopic liver transplantation in rats: Effects of intrahepatic islet isografts and split portal blood flow on liver integrity after auxiliary liver isotransplantation. *Surgery* 1994; 115-1: 108-117.
20. Bestian JM, Janin A, Zenner L, Pruvot FR, Zelus D, Courtade A, Dessaint JP, Capron A. Allogenic microchimerism following auxiliary heterotopic liver transplantation in rat and swine. *Transplantation Proceedings* 1995; 27-2: 1675.
21. Miyata M, Fischer JH, Fuhs M, Isselhard W, Kasai Y. A simple method for orthotopic liver transplantation in the rat – cuff technique for three vascular anastomoses. *Transplantation* 1980; 30-5: 335-338.
22. Kort WJ, Wolff ED, Eastham WN. Heterotopic auxiliary liver transplantation in rats: use of the infrahepatic vena cava as the efferent vessel. *Transplantation* 1971; 12-6: 415-420.
23. Marni A, Ferrero ME. Heterotopic liver grafting in the rat. *Transplantation* 1985; 39-3: 329-331.

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24. Post S, Menger MD, Rentsch M, Gonzalez AP, Herfarth C, Messmer K. The impact of arterialization on hepatic microcirculation and leukocyte accumulation after liver transplantation in the rat. *Transplantation* 1992; 54: 789-794.
25. Ulrichs K, Engemann R, Thiede A, Müller-Ruchholtz W. Allograft tolerance in rats with orthotopic liver transplants: Advantages of rearterialization. *Eur.Surg.Res.* 1981; 79: 13.
26. Sumimoto R, Shinomiya T, Yamaguchi A. Influence of hepatic arterial blood flow in rats with liver transplants. Examination of donor liver-derived serum class I MHC antigen in rats with liver transplants with or without hepatic arterial reconstruction. *Transplantation* 1991; 51: 1138-1139.
27. Reck T, Steinbauer F, Steinbauer M, et al. Impact of arterialization on hepatic oxygen supply, tissue energy phosphates, and outcome after liver transplantation in the rat. *Transplantation* 1996; 62: 582-587.
28. Steffen R, Ferguson DM, Krom RA. A new method for orthotopic rat liver transplantation with arterial cuff anastomosis to the recipient common hepatic artery. *Transplantation* 1989; 48: 166-168.
29. Howden B, Jablonski P, Grossman H, Marshall VC. The importance of the hepatic artery in rat liver transplantation. *Transplantation* 1989; 47: 428-431.
30. Zhao D, Zimmermann A, Wheatley AM. Morphometry of the liver after liver transplantation in the rat: significance of an intact arterial supply. *Hepatology* 1993; 17: 310-317.
31. Engemann R, Ulrichs K, Thiede A, Muller-Ruchholtz W, Hamelmann H. Value of a physiological liver transplant model in rats. Induction of specific graft tolerance in a fully allogeneic strain combination. *Transplantation* 1982; 33: 566-568.
32. Gao W, Lemasters JJ, Thurman RG. Development of a new method for hepatic rearterialization in rat orthotopic liver transplantation. Reduction of liver injury and improvement of surgical outcome by arterialization. *Transplantation* 1993; 56: 19-24.

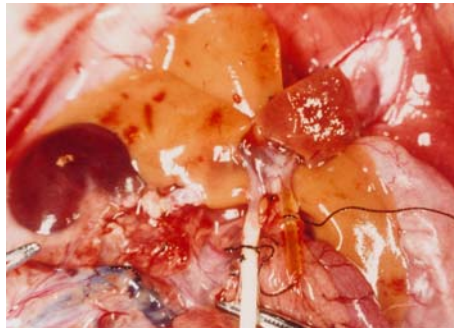


## 42 2.1. Post-HALTx Evaluation of Graft Viability



### ***Donor liver preparation***

After a 68% partial hepatectomy, the common bile duct was intubated; the portal vein and the vena cava were isolated.



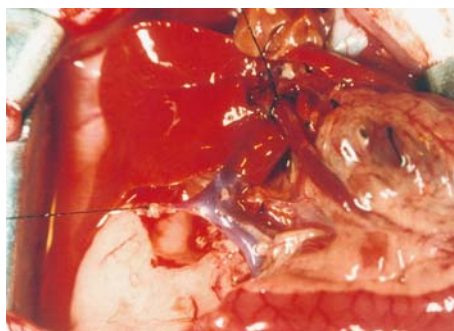
### ***Donor liver perfusion***

The donor liver was then perfused with cold saline solution through the portal vein.



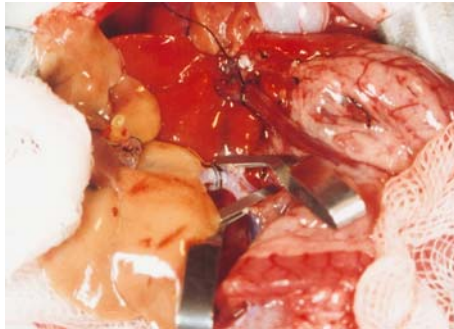
### ***Cuff preparation***

Both the portal vein and the vena cava of the graft were cuffed with a polyethylene tube.



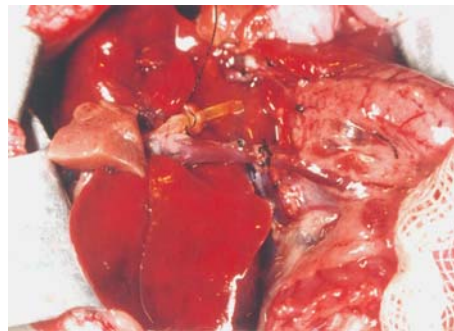
### ***Recipient preparation***

The native liver of the recipient was handicapped with a 68% PH and CBDL. The portal vein and the right renal vein were prepared for vascular anastomoses.



***Graft venous outflow***

The venous drainage of the engrafted liver was made through the right renal vein of the recipient to the vena cava.



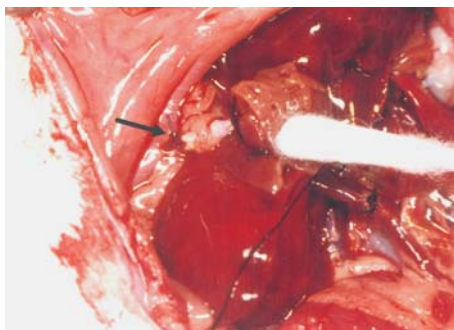
***Graft blood supply***

The engrafted liver received its blood supply from the portal vein of the recipient.



***Graft bile drainage***

The intubated common bile duct of the graft was inserted into the recipient duodenum.



***Graft fixation***

The remained donor diaphragm was fixed to the recipient's lateral abdominal wall.

**2.2.  $T_1$  Relaxation Times for Viability Evaluation of the Engrafted and the Native Liver in a Rat Model of Heterotopic Auxiliary Liver Transplantation: a pilot study**

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**SUMMARY**

Following a heterotopic auxiliary liver transplantation, commonly used measurements are either invasive or non-indicative of individual viability of the coexisting engrafted and native livers. Magnetic resonance imaging (MRI) was therefore tested for its potential to monitor the post-transplant hepatic viability in a rat model. Thirteen Wistar rats were systematically evaluated with MRI and serum biochemical liver parameters. Post-transplant complications and the causes of animal death were identified by autopsy and histo-pathological examinations. The data of the healthy survivors were compared with those of the rats that developed complications. On MRI, the hepatic complications could be depicted in the individual livers. A specific pattern of signal evolution was found in the livers of the healthy survivors: the mean  $T_1$  relaxation times of the engrafted livers increased immediately after transplantation ( $476 \pm 64$  ms, mean  $\pm$  standard deviation, pre-operative;  $730 \pm 48$  ms, week 1) and then declined steadily to a three month value of  $489 \pm 246$  ms; while, following a transient first rise ( $476 \pm 64$  ms, pre-operative;  $589 \pm 28$  ms, week 1), the mean  $T_1$  value of the native livers increased again four weeks after surgery and reached a three month value of  $859 \pm 43$  ms. However, in the rats with various complications, the mean  $T_1$  relaxation times of the engrafted livers continued to increase throughout the first post-operative month ( $760 \pm 48$  ms, week 1;  $922 \pm 76$  ms, week 4), while that of the native liver only varied mildly ( $546 \pm 25$  ms, week 1;  $473 \pm 25$  ms, week 4). After the first post-transplant week, the healthy engrafted livers could already be distinguished from those with complications by a significant decrease in  $T_1$  relaxation times. These data suggest that, besides demonstrating major complications, MRI may allow one to monitor the viability of each liver by analysing the relative signal intensity and  $T_1$  relaxation times after a heterotopic auxiliary liver transplantation.

## INTRODUCTION

In heterotopic auxiliary liver transplantation (HALTx), a donor liver graft is implanted into the recipient without complete removal of the diseased native liver. This operative procedure avoids the surgical trauma of more extensive liver dissection and the an-hepatic phase needed in case of orthotopic liver transplantation (OLTx). For the patients with end-stage chronic liver disease at high risk for OLTx, HALTx can indeed be a life saving alternative<sup>1</sup>. In addition, without removing the otherwise normal liver, an auxiliary liver may correct some inborn errors of metabolism with enzymatic deficiency<sup>2,3</sup>. Above all, HALTx has the advantage over OLTx in treating patients with fulminant and sub-fulminant hepatic failure in whom the function of the native liver may still recover. Following a successful recuperation of the native liver, the auxiliary liver can then be removed or left to atrophy, thus avoiding the life-long use of immunosuppressive drugs and its potential complications<sup>4-9</sup>.

Despite these advantages, HALTx has not widely been used. Comparing to OLTx, HALTx is technically more demanding with a higher incidence of post-transplant complications such as arterial and venous thrombosis. Moreover, as the remaining native liver is functionally competing with the newly engrafted liver, graft atrophy is another major cause of failure<sup>10,11</sup>. Since the so-called “hepatotrophic factors” are generally accepted as the prime factor for the preservation of hepatic integrity, a proper sharing of the portal blood between the two livers is important for the success of the procedure<sup>12</sup>. Up to date, a reliable method of dividing the portal blood flow proportionally between the two livers is still lacking. Careful monitoring of the post-transplant evolution of the two livers is therefore crucial, since an early identification of complications can lead to a possible surgical correction or other interventions that may reverse the otherwise failing procedure. In case of a HALTx for fulminant or sub-fulminant hepatic failure, an accurate assessment of the regeneration of the native liver is also important to define the exact time to taper and eventually to stop immunosuppression, and possibly to remove the engrafted liver<sup>13</sup>. Amongst the most commonly used measurements for the post-transplant evaluation, biological tests of the liver can only show the global hepatic function rather than the respective contributions of each individual liver. Although the individual function of the engrafted liver or the native liver is assessable by Hida scintigraphy<sup>14,15</sup>, various post-transplant complications that may lead to graft dysfunction, such as acute rejection, vascular occlusions, biliary obstruction and abscess formation, cannot be distinguished by this technique<sup>16</sup>; besides, the procedure uses radioactive tracers. Histological examination, which does show the state of the examined organ accurately, requires invasive biopsy procedures and cannot be repeated frequently without risk.

#### **44 2.2. $T_1$ Relaxation Times for Hepatic Evaluation in HALTx**

As a non-invasive technique, magnetic resonance imaging (MRI) may have the advantages of providing reliable information of each individual liver after a HALTx. Moreover, the post-transplant evolution of the two livers recorded by a systematic MRI monitoring may be useful for studies on the mechanisms of “inter-liver competition” with important impact on liver transplantation and hepatological research. This potential had not yet been explored so far and was tested in the current study using a rat model of HALTx.

### **MATERIALS AND METHODS**

#### **Experimental Animals**

Male outbred Wistar rats (B & K Universal Limited, England) weighing 280-350 g were used as donors and recipients. All the animals received humane care according to the European guidelines for animal care (Het Belgisch Staatsblad, Feb. 29<sup>th</sup>, 1992). The rats had free access to water and standard pellet food. Before surgery, food but not water was withdrawn from the recipient rats for 12 hours.

#### **Experimental Design**

The combination of a 68% partial hepatectomy (68% PH) and a common bile duct ligation (CBDL) does severely handicap the function of the native liver and cause atrophy. An engrafted liver in this condition can benefit from stimulating trophic factors circulating in the portal system to regenerate<sup>17</sup>. This operative procedure with drastic changes in the engrafted and the native liver was specifically chosen to test the potential of MRI in post-transplant monitoring.

MRI measurement and serum biochemical liver tests were carried out on a regular basis. An autopsy and subsequent histo-pathological examination were performed whenever a rat died or at the end of the 3-month follow-up period when all the long-term survivors were sacrificed.

Sixteen rats were subjected to HALTx. Only the rats that survived the surgical procedure longer than one week were used for systematic post-transplant evaluation, while those that died within the first post-operative week were regarded as surgical failure and excluded from the study. Based upon autopsy and histo-pathological evidence, the rats were subdivided into two groups: group 1 consisted of the rats free of any complications; the rats that developed post-transplant complications including infection, biliary obstruction, vascular thrombosis and rejection were included in group 2. The data of MRI, liver biology and histology of these two groups were compared.

**Surgical Procedure**

All operations were performed with ether anaesthesia, clean instruments and x 12.5 microscopic magnification. Body temperature of the rats was maintained at approximately 36°C by using heating pads during surgery.

HALTx was performed according to the technique described elsewhere<sup>18</sup>. Briefly, after a 68% PH, the donor liver was perfused with cold ( $\pm 4^{\circ}\text{C}$ ) heparinized Ringer's solution via the portal vein. Following a right nephrectomy, the liver graft was implanted in the right paravertebral gutter under the native liver of the recipient. The cuff technique was used for the anastomoses of both the vena cava and the portal vein. The engrafted liver received its portal blood supply from the portal vein of the recipient. The venous drainage was made by connecting the infrahepatic vena cava of the donor liver to the right renal vein of the recipient. No re-arterialisation of the engrafted liver was attempted in this study. Restoration of the bile drainage was done by insertion of the intubated common bile duct of the donor liver into the recipient's duodenum. As the portal flow of the recipient was completely shunted to the engrafted liver, the blood supply of the native liver remained solely from the hepatic artery. The native liver was further handicapped with both a 68% PH and a CBDL.

For all the operative procedure of HALTx in the rats of both groups, the cold ischemia time of the liver grafts was kept between 62 and 74 minutes. The duration of warm ischemia of the liver grafts (from the implantation to the blood re-circulation of the engrafted liver) was between 11 and 14 minutes.

At the end of the operation, the rats were transfused with 1-1.5 cc of Ringer's solution and an intramuscular injection of 10 mg Na Cefazolin was given. No immunosuppressive agent was administered. The pre-operative diet of food pellets was resumed 2 hours after the operation.

**MR Evaluation**

During MRI examination, the rat was anaesthetised with 1.2 vol % isoflurane through a gas mask. Examinations were performed on a 1.0 Tesla scanner (EXPERT, Siemens<sup>TM</sup> Germany) using a standard circular polarised knee-coil with a diameter of 25 cm. Each MR experiment lasted less than 30 minutes. To establish the reference value of  $T_1$  relaxation time in the normal rat liver, thirteen rats of the same strain free of any surgical manipulation were scanned. MRI examinations of the transplanted rats were performed pre-operatively and weekly during the evaluation period of three months. An extra five rats subjected only to a 68% PH and a CBDL without HALTx were also evaluated weekly during the first post-operative month to identify the effects of these surgical interventions on the status of the native liver.

## 46 2.2. $T_1$ Relaxation Times for Hepatic Evaluation in HALTx

For visual evaluation, the rats were scanned with 4 mm thick axial slices with  $T_1$ -weighted contrast ( $T_1W$ ) (TR/TE/NEX = 400 ms/12 ms/4), and 2 mm thick coronal slices with  $T_1W$  contrast (400 ms/12 ms/4).

For quantitative evaluation, proton relaxometry with calculation of  $T_1$  relaxation times in the engrafted and the native livers was carried out. For this, a spin-echo sequence was used and measurements were repeated with different repetition times (TR/TE = 2000,1000,500,250/12 ms). A  $T_1$  relaxation curve was obtained by plotting the signal intensities in a region of interest in the engrafted and the native livers as a function of the repetition time. The  $T_1$  relaxation time was extracted from the plots by fitting the data points against the theoretical  $T_1$  relaxation functions given by the equation:

$$S_{(TR)} = S_0 \cdot (e^{-TR/T_1} - 2 \cdot e^{-\frac{-(TR-TE/2)}{T_1}} + 1)$$

with  $S_0$  and  $T_1$  as the unknown variables. Fitting was performed by use of a home-written computer program based on the Levenberg-Marquardt algorithm, as this is proven to be preferable above other fitting methods with regard to signal-to-noise consideration<sup>19</sup>.

### Liver Biology

A blood sample of 0.4 - 0.5 ml was collected from the rat's tail vein for serum biochemical tests. Using routine photometric tests on a Hitachi 747 analyser, serum alanine aminotransferase activity (ALAT) and bilirubin concentration (TBil) were determined pre-operatively and further on the post-operative day 3, 7, 14, 21, 28, 56 and 84.

### Autopsy and Histology

During each autopsy, the examination was focused on the macroscopic appearance of the engrafted and the native liver, the patency of the vascular anastomoses and the common bile duct, the complications and the possible causes of animal death where premature death occurred.

The livers were excised, cleaned of gross adhesions, and fixed in a 10% formalin solution. Four-micron-thick sections were stained with hematoxylin and eosin (HE), and examined light-microscopically.

### Statistics

The data were expressed as the mean  $\pm$  standard deviation (SD). The distribution of the data was checked by Kolmogorov test. The Student's  $t$ -test or the Mann-Whitney rank sum test was applied for normally or not-normally distributed data, respectively.  $P$  values equal to or less than 0.05 were considered statistically significant.

## RESULTS

### Post-operative Animal Survival

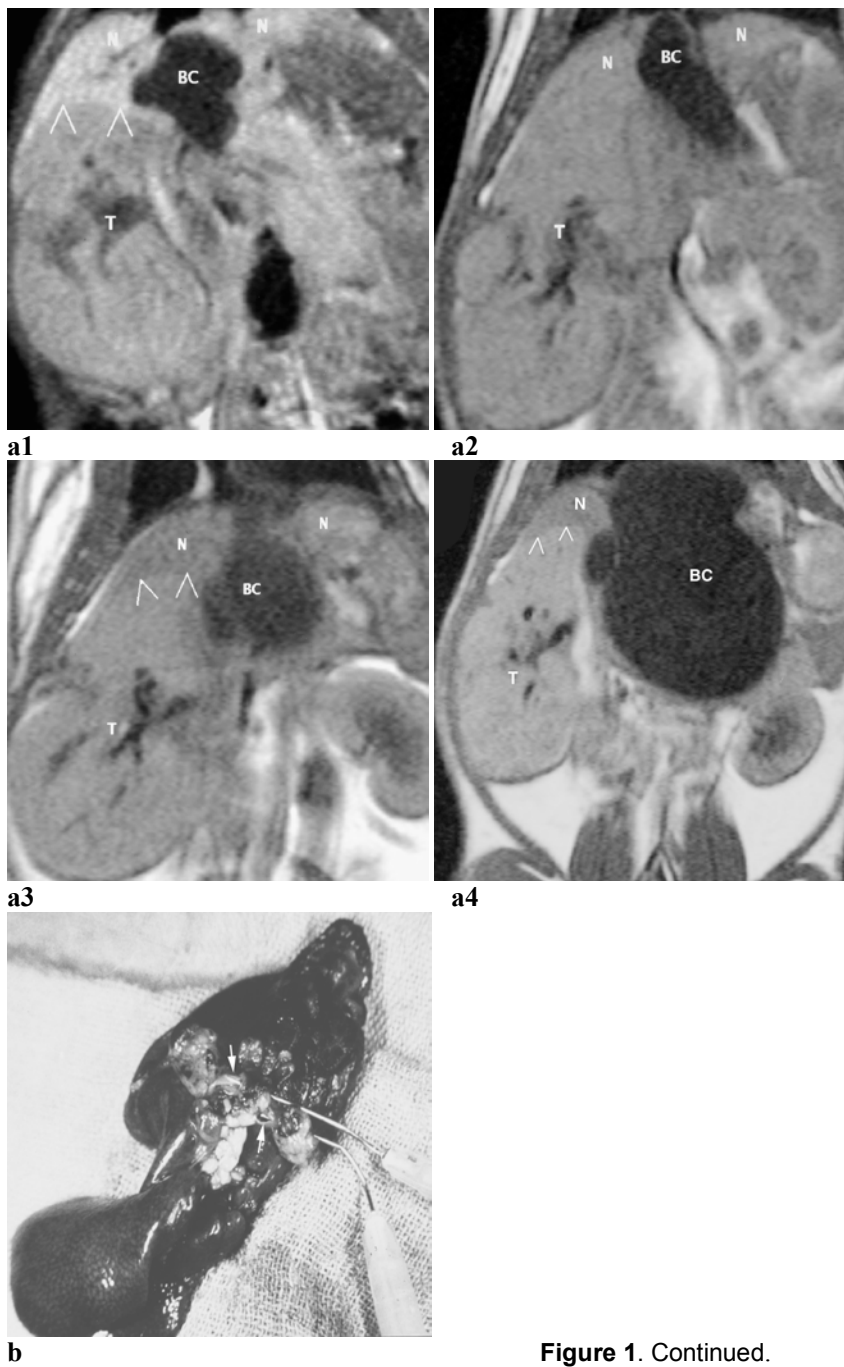
Three out of the sixteen rats were excluded from the study due to early post-operative death. One rat died of bleeding on the second day, while peritonitis secondary to bile leakage caused another two animal losses on the third and fourth day. Amongst the thirteen rats that survived longer than one week after HALTx, four appeared normal during the observation period of three-month and no signs of infection, biliary obstruction, vascular thrombosis and rejection were found at sacrifice. These four rats were included in group 1. Among the nine rats of group 2, eight developed various complications and died during the first seven weeks after transplantation (1 in week 2, 2 in week 3, 1 in week 4, 1 in week 5, 1 in week 6 and 2 in week 7). Cholangitis ( $n = 4$ ), liver abscess ( $n = 2$ ), vena cava thrombosis ( $n = 1$ ) and rejection ( $n = 1$ ) were found to be the major causes of animal death. Only one out of nine rats in group 2 survived longer than seven weeks, which was sacrificed at the end of the eighth week due to abnormal MRI data. Portal vein thrombosis of the engrafted liver was identified.

### MRI

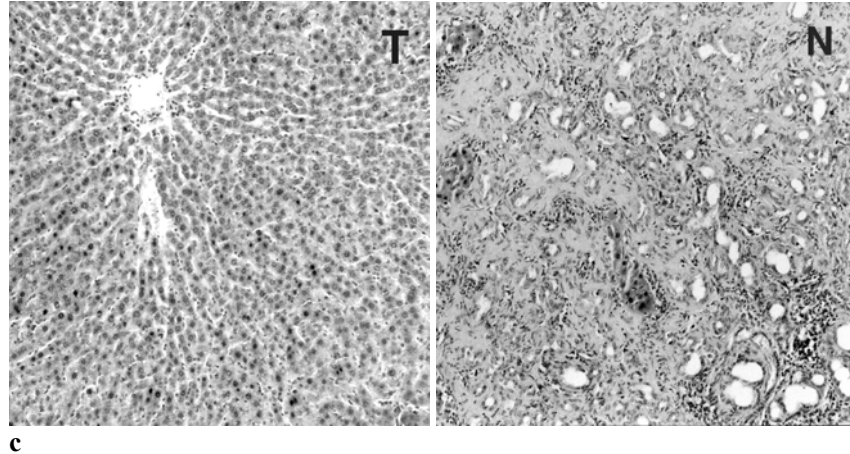
Using anatomical landmarks, the engrafted and the native liver could clearly be distinguished from each other on MR images. In the rats without complications (group 1), both livers remained homogeneous throughout the observation period. Notably, the signal intensity of the two livers showed a transition on the  $T_1W$  images during the observation period: the signal intensity of the engrafted livers evolved from hypo-intense to hyper-intense, while that of the native livers changed from hyper-intense to hypo-intense (*Figure 1a*). In comparison, in the rats with post-transplant complications (group 2), the signal of the engrafted livers was in-homogeneous and showed low signal areas on  $T_1W$  images at various times. These hypo-intense areas were later identified as complications such as abscess formation. Moreover, no transition of signal intensity was observed: the signal intensity of the engrafted liver remained hypo-intense, while that of the native liver was constantly hyper-intense even in the rat that was sacrificed two months after HALTx (*Figure 2a*).



**48** 2.2.  $T_1$  Relaxation Times for Hepatic Evaluation in HALTx

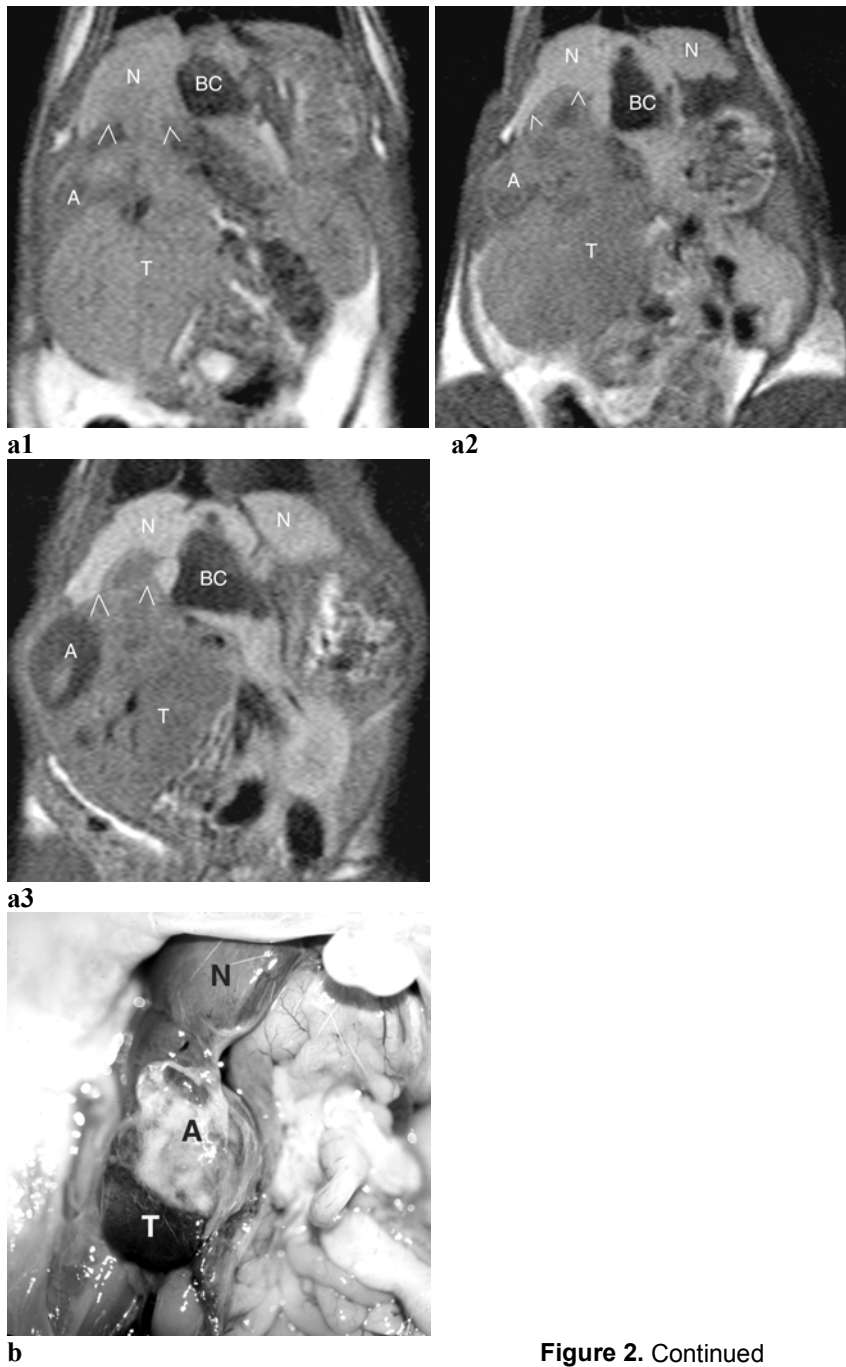


**Figure 1.** Continued.

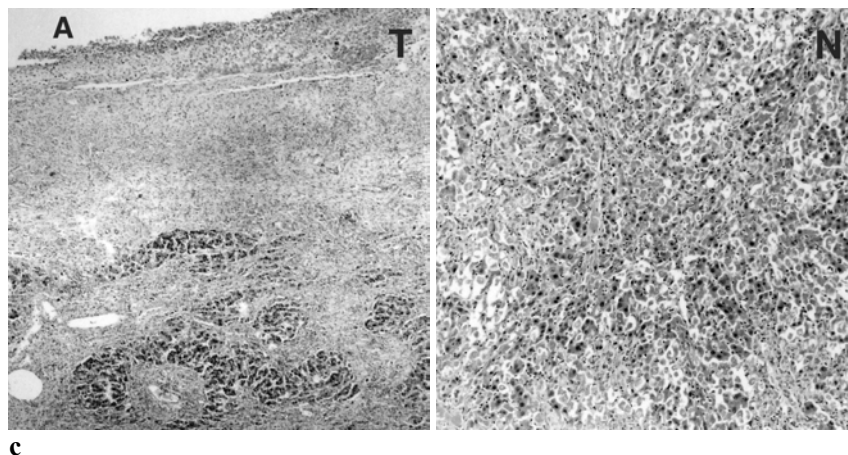


**Figure 1. (a1-4)** Post-HALTx evolution of signal intensity of a rat in group 1 (in a series of 2, 4, 6 and 12 weeks). Relative to that of the other liver, the signal intensity of the engrafted liver (T) evolved from hypo-intense to hyper-intense, while that of the native liver (N) changed in opposite direction during the 3 months observation period. '∧' shows the interface between the two livers. BC indicates a bile cyst. **(b)** Autopsy observation (3 months post-HALTx). The engrafted liver revealed a normal appearance with patent portal vein and vena cava anastomoses (indicated by '↑'); **(c)** Histological findings (3 months post-HALTx). The engrafted liver (T) preserved its normal lobular architecture without major histo-pathological alterations, while that of the native liver (N) was completely destroyed and replaced by ductular metaplasia in the portal tracts accompanied by fibrosis (original magnification x 125, HE staining).

**50** 2.2.  $T_1$  Relaxation Times for Hepatic Evaluation in HALTx



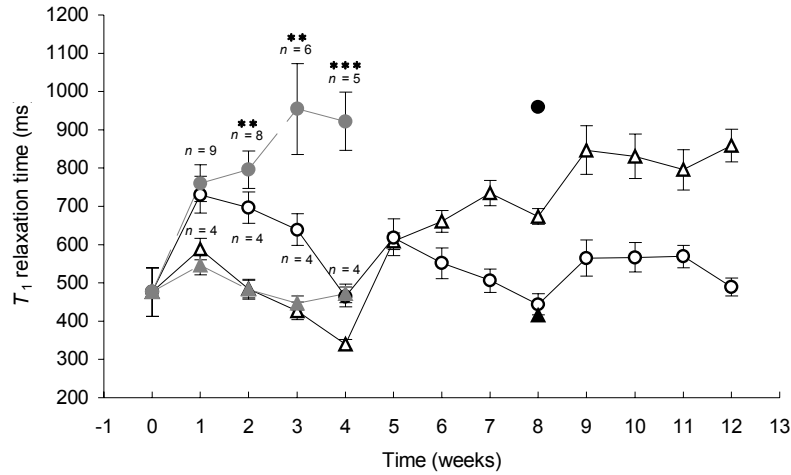
**Figure 2.** Continued



**Figure 2. (a1-3)** Post-HALTx evolution of signal intensity of a rat in group 2 (in a series of 1, 2 and 4 weeks). Relative to that of the other liver, the signal intensity of the engrafted liver (T) remained hypo-intense, while that of the native liver (N) became more hyper-intense. The hypo-intensity in the engrafted liver (A) was identified as an abscess at autopsy. '^' shows the interface between the two livers. BC indicates a bile cyst. **(b)** Autopsy observation (7 weeks post-HALTx). The engrafted liver (T) appeared atrophic and an abscess (A) was observed, while the size of the native liver (N) was less changed. **(c)** Histological findings (7 weeks post-HALTx). Besides remnants of hepatocytes, the general lobular architecture was hardly visible in the engrafted liver (T), which contained an abscess (A) (HE, original magnification  $\times 100$ ). The native liver (N) revealed ischemic necrosis of the hepatocytes (HE, original magnification  $\times 250$ ).

The evolution of the signal intensity was the visual reflection of changing  $T_1$  relaxation times (*Figure 3*). The reference  $T_1$  value of the liver established from thirteen normal rats was  $476 \pm 64$  ms. In group 1, the mean value of the  $T_1$  relaxation time of the engrafted livers showed a transient increase immediately after HALTx with a peak value in the first post-operative week ( $476 \pm 64$  ms, pre-operative;  $730 \pm 48$  ms, week 1). Thereafter, the value fell back to the pre-transplant level at the one-month time point ( $464 \pm 26$  ms) and remained around this same range till the end of observation period of three months ( $489 \pm 24$  ms). Parallel to the evolution of the engrafted livers, the mean  $T_1$  value of the native livers increased only slightly from a pre-operative value of  $476 \pm 64$  ms to  $589 \pm 28$  ms in the first week. The value increased again one month after the transplantation, kept on rising to another

## 52 2.2. $T_1$ Relaxation Times for Hepatic Evaluation in HALTx



**Figure 3.** Post-HALTx evolution of mean  $T_1$  relaxation time (ms) in group 1 (○ — ○, the engrafted livers; △ — △, the native livers) and in group 2 (● — ●, the engrafted livers; ▲ — ▲, the native livers; ●, the engrafted liver and ▲, the native liver of the rat that was sacrificed at the end of the second post-transplant month). Values are expressed as mean  $\pm$  SD. The asterisk indicates statistically significant differences between the engrafted livers of group 1 (○ — ○) and the engrafted livers of group 2 (● — ●). \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ .

peak in the seventh week ( $737 \pm 33$  ms), and remained high till the end of the third month ( $859 \pm 43$  ms).

In contrast to the pattern presented by group 1, group 2 showed a rather different profile (Figure 3). Instead of an initial increase followed by a decrease, the mean  $T_1$  relaxation time of the engrafted livers in this group continued to increase throughout the first post-operative month ( $476 \pm 64$  ms, pre-operative;  $922 \pm 76$  ms, week 4), while that of the native livers only varied mildly during the same period of time ( $476 \pm 64$  ms, pre-operative;  $473 \pm 25$  ms, week 4). This high  $T_1$  in the engrafted liver and low  $T_1$  in the native liver was again recorded in the rat that was sacrificed at the end of the second post-transplant month (959 ms, the engrafted liver; 415 ms, the native liver). Compared with the engrafted livers in group 1, the mean  $T_1$  relaxation times of the engrafted livers in group 2 were significantly higher in the second ( $p = 0.005$ ), the third ( $p = 0.001$ ) and the fourth post-transplant week ( $p < 0.001$ ).

**Table 1.** Comparison of post-operative  $T_1$  relaxation time (ms) between the native livers of experimental groups and the livers subjected only to 68% PH and CBDL

	68% PH/CBDL ( $n = 5$ )	NL-Group 1 ( $n = 4$ )	NL-Group 2
Pre-operative reference value: $476 \pm 64$			
week 1	$519 \pm 54$	$589 \pm 28$	$546 \pm 25$ ( $n = 9$ )
week 2	$537 \pm 81$	$485 \pm 25$	$483 \pm 24$ ( $n = 8$ )
week 3	$446 \pm 52$	$427 \pm 23$	$447 \pm 18$ ( $n = 6$ )
week 4	$602 \pm 50$	$340 \pm 13^{***}$	$473 \pm 25$ ( $n = 5$ ) <sup>***</sup>
week 8		$673 \pm 21$	
week 12		$859 \pm 43$	

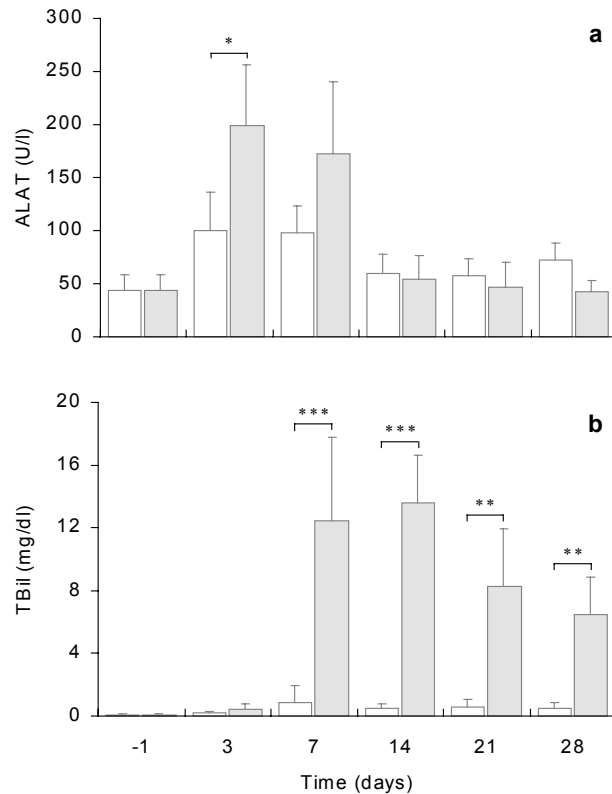
Notes: 68% PH: 68% partial hepatectomy; CBDL: common bile duct ligation; NL: native liver. Data are expressed as mean  $\pm$  SD. The asterisk indicates statistically significant differences between the values of experimental group and that of the rats subjected only to 68% PH and CBDL. \*\*\*  $p < 0.001$ .

During the first three post-operative weeks, as shown in *Table 1*, the mean  $T_1$  relaxation time of the native livers in both group 1 and 2 was similar to that of the livers subjected only to 68% PH and CBDL, all of which remained in or slightly above the normal range ( $476 \pm 64$  ms). This might indicate that the significantly higher  $T_1$  value of the livers subjected only to 68% PH and CBDL in the fourth week ( $p < 0.001$ ) as well as the constantly elevated  $T_1$  of the native livers in the rats of group 1 four weeks after HALTx were caused by the factors beyond surgical trauma.

### Liver Biology

The mean serum ALAT values elevated in both group 1 and group 2 after transplantation, reached peak levels on the third day and then returned to normal range in two-weeks time. The mean ALAT values of group 2 showed a more marked increase than that of group 1 on day 3 ( $199 \pm 57$  U/l vs.  $91 \pm 54$  U/l;  $p = 0.05$ ) (*Figure 4a*). Serum TBil revealed pronounced differences between the two groups as shown in *Figure 4b*. Compared with those of group 1, the mean TBil values of group 2 were significantly higher on the seventh ( $1.31 \pm 1.49$  mg/dl, group 1;  $12.52 \pm 4.94$  mg/dl, group 2;

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**Figure 4.** Post-HALTx evolution of **(a)** mean serum alanine aminotransferase activity (ALAT, U/l) and **(b)** mean bilirubin concentration (TBil, mg/dl) in group 1 (□) and group 2 (■). Values are expressed as mean  $\pm$  SD. The asterisk indicates statistically significant differences between the two groups. \*  $p \leq 0.05$ ; \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ .

$p < 0.001$ ), the fourteenth ( $0.48 \pm 0.34$  mg/dl,  $13.26 \pm 2.81$  mg/dl, respectively;  $p < 0.001$ ), the twenty-first ( $0.63 \pm 0.54$  mg/dl,  $8.26 \pm 2.79$  mg/dl, respectively;  $p = 0.005$ ) and the twenty-eighth day ( $0.53 \pm 0.34$  mg/dl,  $6.63 \pm 2.01$  mg/dl, respectively;  $p = 0.007$ ).

### Autopsy and Histology

At sacrifice, the four long-term survivors of group 1 showed enlarged liver grafts with normal appearance and patent anastomoses of the portal vein and the vena cava (*Figure 1b*), while their native livers were atrophic. The histological examination revealed that the general architecture of those

engrafted livers was well preserved, while that of the handicapped native livers was completely altered and replaced by ductular metaplasia in the portal tracts accompanied by fibrosis (*Figure 1c*).

In contrast, all the engrafted livers of the rats in group 2 appeared atrophic, while the size of the native livers was less changed (*Figure 2b*). As a result of common bile duct ligation, a bile cyst was observed at the hilus of the native livers in all the rats of group 1 and 2. Histological examination showed a drastically destructed lobular architecture of the engrafted livers due to cholangitis, rejection, portal vein thrombosis and liver abscess, while various degree of ischemic hepatocytes necrosis was observed in the native livers (*Figure 2c*).

## DISCUSSION

MRI seems an excellent technique to monitor changes in both the engrafted and the native liver after a HALTx. As the anatomical border of each liver is identifiable, the post-transplant complications, such as intra- or inter-hepatic abscess and liver necrosis, can visually be depicted. Moreover, the data of this study show that, in the absence of steatosis<sup>20</sup>, the better the hepatic viability, the higher the signal intensity on  $T_1W$  images. This finding corresponds well with a previous report by Siegelman and his colleagues, in which the similar signal pattern has been observed in a clinical case<sup>21</sup>. The signal intensity of the liver on each MR image indicates therefore the viability of the organ at that specific time point, especially in respect to its physiological position in the inter-liver competition, even when no obvious complications can yet be seen. MRI could thus have great clinical applications as an indirect and accurate measurement of hepatic viability. A technique as such is particularly needed in some clinical situations, for instance pre-surgical evaluation of transplant candidate in fulminant or sub-fulminant hepatic failure, post-transplant or post-hepatectomy assessment of liver viability.

Several experimental and clinical studies have demonstrated a close correlation between  $T_1$  relaxation time and the water content of the liver tissue: the lower the  $T_1$  value, the lower the water content<sup>22-26</sup>. In addition, a low  $T_1$  may also be indicative of a normal bile production and energy metabolism<sup>22</sup>. As normal water content can only be preserved with integrity of cell membranes, an elevated  $T_1$  could reflect the processes such as edema, cell death, disintegration and inflammation in the examined organ<sup>23</sup>. The transient post-transplant increase of  $T_1$  value in the engrafted and the native livers observed in this study may reveal the degree of hepatic trauma caused by surgery. Following a peak value, the constantly decreasing  $T_1$  relaxation



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times of the healthy hepatic grafts demonstrate the recuperating process of the engrafted livers from the preservation and re-perfusion injury. Meanwhile, the functional deterioration of the handicapped native livers caused by biliary obstruction and partial hepatectomy is represented by the elevation of the  $T_1$  relaxation time one month after HALTx. When the engrafted livers fully recover and eventually take over the hepatic function, the native livers enter a phase of progressive atrophy. MRI records this “take-over” as a cross-point around the fifth post-transplant week. As a functioning engrafted liver may reduce the toxic effects of the cholestasis of the handicapped native live, the discrepancy of the  $T_1$  values between the native livers of the transplanted group and the handicapped livers without HALTx may be interpreted as another sign of this recovery. On the other hand, if a complication such as biliary obstruction, vascular thrombosis and abscess formation occur in the engrafted livers, this normal pattern is no longer seen. With increasing inflammation, the  $T_1$  relaxation times of the engrafted livers, instead of declining, keep on rising after transplantation. Meanwhile, less altered  $T_1$  relaxation times recorded in the native livers reveal that the handicapped organs remain relatively normal to compensate the functional deterioration of the engrafted livers.

In addition, as early as after the first post-transplant week, a dramatic difference of the  $T_1$  relaxation times of the engrafted livers could be found between the healthy long-term survivors and the rats with complications. This observation may indicate that if no MRI sign of recovery from the operative trauma appears at this time, the engrafted liver is developing either an acute or a chronic complication. The eventual outcome of the engrafted liver and the native liver in the inter-liver competition can therefore be predicted at an early post-transplant stage through MRI analysis.

A previous study has shown that, in this specific rat model of HALTx, the functionality of the native liver is so severely damaged by surgical handicap that the rat will die unless a viable auxiliary liver is present. In case a post-operative complication occurs in the engrafted liver at a later stage, the animal is unable to survive, as the handicapped native liver is already atrophic at that time<sup>17</sup>. Despite a portal vein thrombosis developed in the engrafted liver, however, a rat of group 2 in this study survived longer than two months following transplantation. The  $T_1$  relaxation time of its native liver prior to sacrifice was in the normal range. The reason why this handicapped native liver is still able to maintain certain function for such a long period is unknown. The progressive nature of portal vein thrombosis after transplantation may explain part of this observation.

Although definitive conclusions can only be drawn from a larger sample study, the data of this pilot experiment do show the beneficial potential of using MRI in post-HALTx monitoring. Prospective clinical investigations

are indeed worthy to test the findings of this study that major post-transplant complications can be depicted individually by this measurement, and that the hepatic function is also assessable by analysis of the signal intensity and  $T_1$  relaxation times. These studies will demonstrate if the final outcome of each liver may be forecasted as early as after the first post-transplant week. As MRI allows documenting the evolution of the engrafted and the native liver systematically and non-invasively, this procedure can thus facilitate the studies on inter-liver functional competition.

## REFERENCES

1. Metselaar HJ, Hesselink EJ, Schalm SW, Terpstra OT. Favorable results of auxiliary heterotopic liver transplantation in patients with end-stage chronic liver insufficiency. *Ned Tijdschr Geneesk* 1991; **135**: 1221-6.
2. Rela M, Muiesan P, Andreani P, Gibbs P, Mieli-Vergani G, Mowat AP, Heaton ND. Auxiliary liver transplantation for metabolic diseases. *Transplant Proc* 1997; **29**: 444-5.
3. Burdelski M, Rogiers X. Liver transplantation in metabolic disorders. *Acta Gastroenterol Belg* 1999; **62**: 300-5.
4. Terpstra OT. Auxiliary liver grafting: a new concept in liver transplantation [comment]. *Lancet* 1993; **342**: 758.
5. Boudjema K, Cherqui D, Jaeck D, Chenard-Neu MP, Steib A, Freis G, Becmeur F, Brunot B, Simeoni U, Bellocq JP. Auxiliary liver transplantation for fulminant and subfulminant hepatic failure. *Transplantation* 1995; **59**: 218-23.
6. Shaw BWJ. Auxiliary liver transplantation for acute liver failure. *Liver Transpl Surg* 1995; **1**: 194-200.
7. van Hoek B, Ringers J, Kroes AC, van Krieken JH, van Schelven WD, Masclee AA, van Krieken-Hogenberk LG, Haak HR, Lamers CB, Terpstra OT. Temporary heterotopic auxiliary liver transplantation for fulminant hepatitis B. *J Hepatol* 1995; **23**: 109-18.
8. Chenard-Neu MP, Boudjema K, Bernuau J, Degott C, Belghiti J, Cherqui D, Costes V, Domergue J, Durand F, Erhard J, de Hemptinne B, Gubernatis G, Hadengue A, Kemnitz J, McCarthy M, Maschek H, Mentha G, Oldhafer K, Portmann B, Praet M, Ringers J, Rogiers X, Rubbia L, Schalm S, Bellocq JP. Auxiliary liver transplantation: regeneration of the native liver and outcome in 30 patients with fulminant hepatic failure--a multicenter European study. *Hepatology* 1996; **23**: 1119-27.
9. van Hoek B, de Boer J, Boudjema K, Williams R, Corsmit O, Terpstra OT. Auxiliary versus orthotopic liver transplantation for acute liver failure. EURALT Study Group. European Auxiliary Liver Transplant Registry. *J Hepatol* 1999; **30**: 699-705.
10. Starzl TE, Marchioro TL, Huntley RT. Experimental and clinical homotransplantation of the liver. *Amer NY Acad Sci* 1964; **120**: 739-65.

## 58 2.2. $T_1$ Relaxation Times for Hepatic Evaluation in HALTx

11. Marchioro TL, Porter KA, Dickins TC, Faris TD, Starzl TE. Physiologic requirements for auxiliary liver transplantation. *Surg Gynecol Obstet* 1965; **121**: 17-31.
12. Starzl TE, Halgrimson CG, Francavilla FR, Porter KA, Brown TH, Putnam CW. The origin, hormonal nature and action of hepatotrophic substances in portal venous blood. *Surg Gynecol Obstet* 1973; **137**: 179-99.
13. Erhard J, Lange R, Rauen U, Scherer R, Friedrich J, Pietsch M, de Groot H, Eigler FW. Auxiliary liver transplantation with arterialization of the portal vein for acute hepatic failure. *Transpl Int* 1998; **11**: 266-71.
14. Uemoto S, Yabe S, Inomata Y, Nishizawa H, Asonuma K, Egawa H, Kiuchi T, Okajima H, Yamaoka Y, Yamabe H, Inui A, Fujisawa T, Tanaka K. Coexistence of a graft with the preserved native liver in auxiliary partial orthotopic liver transplantation from a living donor for ornithine transcarbamylase deficiency. *Transplantation* 1997; **63**: 1026-8.
15. Sakahara H, Kiuchi T, Nishizawa S, Saga T, Nakamoto Y, Sato N, Higashi T, Tanaka K, Konishi J. Asialoglycoprotein receptor scintigraphy in evaluation of auxiliary partial orthotopic liver transplantation. *J Nucl Med* 1999; **40**: 1463-7.
16. Buyck D, Bonnin F, Bernuau J, Belghiti J, Bok B. Auxiliary liver transplantation in patients with fulminant hepatic failure: hepatobiliary scintigraphic follow-up. *Eur J Nucl Med* 1997; **24**: 138-42.
17. Fan YD, Praet M, de Hemptinne B. The need to handicap the recipient's native liver in the rat model of heterotopic auxiliary liver transplantation. *HPB Surg* 1999; **11**: 225-33.
18. Fan YD, Leroux-Roels G, Praet M, Hesse U, de Hemptinne B. Evaluation of graft viability in heterotopic auxiliary liver transplantation in the rat. *J Invest Surg* 1999; **12**: 327-34.
19. De Deene Y, Van de Walle R, Achten E. Mathematical analysis and experimental investigation of noise in quantitative magnetic resonance imaging applied to polymer gel dosimetry. *Signal processing* 1998; **70**: 85-101.
20. Siegelman ES. MR imaging of diffuse liver disease. Hepatic fat and iron. *Magn Reson Imaging Clin N Am* 1997; **5**: 347-65.
21. Siegelman ES, Mitchell DG, Rubin R, Moritz MJ, Munoz SJ, Palazzo JP, Rifkin MD. Recovery of native liver after heterotopic liver transplantation for fulminant hepatic failure: MR studies. *J Comput Assist Tomogr* 1992; **16**: 152-4.
22. Holzmüller P, Moser E, Reckendorfer H, Burgmann H, Sperlich M. Proton spin-lattice relaxation time as liver transplantation graft viability parameter. *Magn Reson Imaging* 1993; **11**: 229-39.
23. Holzmüller P, Moser E, Reckendorfer H, Burgmann H, Winklmayr E, Sperlich M. Proton spin-spin relaxation times as liver transplantation graft viability parameter. *Magn Reson Imaging* 1993; **11**: 749-59.
24. Thomsen C. Quantitative magnetic resonance methods for in vivo investigation of the human liver and spleen. Technical aspects and preliminary clinical results. *Acta Radiol Suppl* 1996; **401**: 1-34.

25. Tsybulevskii AI, Dubovaia TK, Sergeev AI. Use of antihypoxants and antioxidants for pharmaco-correction of disorders of water metabolism in the liver and small intestine after vagotomy. *Patol Fiziol Eksp Ter* 1996; **4**:33-6.
26. Kreft B, Dombrowski F, Block W, Bachmann R, Pfeifer U, Schild H. Evaluation of different models of experimentally induced liver cirrhosis for MRI research with correlation to histopathologic findings. *Invest Radiol* 1999; **34**: 360-6.

## **CHAPTER 3**

### **STUDY OF INTER-LIVER FUNCTIONAL COMPETITION FOLLOWING HETEROTOPIC AUXILIARY LIVER TRANSPLANTATION**

**3.1. The Need to Handicap the Recipient's Native Liver in the Rat Model of Heterotopic Auxiliary Liver Transplantation**

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**SUMMARY**

In the rat model of heterotopic auxiliary liver transplantation (HALTx), the opinion varies on whether and how the recipient's native liver should be handicapped. To avoid atrophy of the transplanted organ, in this study, two different handicaps were evaluated and their effects on post-operative animal survival and liver biology were described. With a sole portacaval shunt (group 1) all rats survived longer than 3 months. An additional handicap of the liver with either a 68% partial hepatectomy (68% PH) (group 2), or both a 68% PH and a common bile duct ligation (CBDL) (group 3) led to a 100% lethality within 2 days after surgery. When an auxiliary liver was transplanted to the rats handicapped with a 68% PH (group 4), serum TBil and ALAT values were significantly lower than those handicapped with both a 68% PH and a CBDL (group 5). Autopsy and histology of the long-term survivors revealed the atrophy of the engrafted livers and the regeneration of the native livers in group 4 whereas it showed the opposite in group 5. Thus the various manipulations of the native liver do influence differently the post-transplant animal survival, serum liver biochemistry and the outcome of the engrafted liver in this rat model of HALTx.

**INTRODUCTION**

In recent years, more and more reports from different organ-transplantation centres have been published about encouraging results of using heterotopic auxiliary liver transplantation (HALTx) to treat patients with end-stage liver diseases especially fulminant and subfulminant hepatic failure <sup>1-3</sup>. This has raised new attention to study the potential problems of this operative procedure with animal models <sup>4-5</sup>. HALTx in the rat has long been used for experimental research, since it was first described by Lee and Edgington in 1966 <sup>6</sup>. The investigators have used this model to study amongst others, liver regeneration, hepatotrophic factors, allogeneic microchimerism and graft rejection <sup>5,7,8</sup>. Several technique modifications of this model have been introduced in that respect <sup>9-12</sup>.

Although the observation of graft atrophy and the concept of “inter-liver competition” in HALTx were reported in the early 60's <sup>13-15</sup>, opinion still varies on whether in this rat model the native liver of the recipient should be handicapped and in this case which approach should be used. Some authors handicapped the native liver either with a 68% partial hepatectomy (68% PH) <sup>7,10</sup> or a 68% PH plus a common bile duct ligation (CBDL) <sup>9,11</sup>. Others, on the contrary, left the native liver intact <sup>5,8,12</sup>. Up to date, there has been no report about how these different manipulations of the native liver do influence the post-HALTx animal survival and serum biochemical values. This was investigated in the present study.

**MATERIALS AND METHODS****Experimental Design (Table 1)**

To evaluate the effects of different handicaps on the native liver, three experimental groups were studied, in which the livers were put in functionally similar situations as the native livers in the HALTx model: group 1 ( $n = 10$ ), an end-to-side portacaval shunt (PCS) (without further handicap of the liver); group 2 ( $n = 10$ ), a PCS plus a 68% PH as additional handicap of the liver; and group 3 ( $n = 10$ ), a PCS, a 68% PH and a CBDL to handicap the liver.

Two experimental groups were tested later on, since no long-term survivor and thus serum biochemical evolution could be obtained from the rats of group 2 and 3. In group 4 ( $n = 12$ ), a HALTx was performed after the native liver was handicapped in the same way as in group 2 (PCS and 68% PH). In group 5 ( $n = 12$ ), HALTx was performed after the handicap of the native liver as in group 3 (PCS, 68% PH and CBDL).

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### Experimental Animals

Male Wistar rats weighing 250-340g were purchased from Janssen Pharmaceutica N.V., Belgium. Donor and recipient rats were matched for size. Rats had free access to food and water prior to surgery. All animals received humane care as outlined in the *Guide for the Care and Use of Laboratory Animals* (NIH Publication No. 86-23, revised 1985).

### Surgical Techniques

All operations were carried out under ether anaesthesia with clean but non-sterile instruments and x 12.5 magnification.

- (a) *68% PH*: The median lobe and the left lobe of the liver were ligated and then excised according to the standard method introduced by Higgins and Anderson<sup>16</sup>.
- (b) *CBDL*: A segment of the common bile duct was removed after double ligation.
- (c) *PCS*: The standard method described by Lee<sup>17</sup> was modified using the cuff technique to perform a PCS. After a left nephrectomy, the left renal vein was freed up to the vena cava, which was then cross-clamped. An incision was made into the anterior wall of the stump of the left renal vein. The portal vein was divided from the pyloric vein to the liver hilus, tied, transected and cuffed with a 0.4 cm length polyethylene tube. The cuffed portal vein was then inserted into the left renal vein and secured with a 5-0 ligature.
- (d) *HALTx*: The standard technique described by Marni<sup>9</sup> was used with a slight modification. After the donor liver was perfused with heparinized 4°C physiological saline solution via the portal vein, 32% of the liver was used for transplantation. The native liver of the recipient rat was handicapped with a 68% PH or both a 68% PH and a CBDL according to the study group, while the hepatic artery remained intact. The engrafted liver was then implanted in the right paravertebral gutter under the native liver of the recipient. The cuff technique described by Kamada<sup>18</sup> was used for the anastomoses of both the vena cava and the portal vein. The engrafted liver received its blood supply from the portal vein of the recipient. The venous drainage was made through the right renal vein of the recipient to the vena cava. Restoration of the bile drainage was done by insertion of the intubated common bile duct of the donor liver into the recipient duodenum. No re-arterialisation was performed. To this standard procedure, we



added a fixation of the remaining donor diaphragm to the recipient lateral abdominal wall in order to avoid twisting and kinking of the donor vena cava.

The duration of the different operative phases (mainly the portal vein and the vena cava cross-clamping time) was comparable among group 1, 2 and 3. The cold preservation time of the grafts (starting from the saline perfusion via the portal vein ending at the portal re-vascularization of the engrafted liver) was kept between 72 and 79 minutes in the rats of HALTx group 4 and 5.

#### **Post-operative Care**

All rats were given 10 mg Na Cefazolin intramuscularly after the operation and this was repeated daily for three days. No immunosuppressive agent was administered to the rats. During the following twenty-four hours, rats were allowed to drink 5% glucose, after which the pre-operative diet of food pellets and water was resumed.

#### **Follow-up**

Body weight was recorded daily and survival time was determined.

For the biochemical analysis, a blood sample of 0.4 - 0.5 ml was collected from the tail vein. Serum total bilirubin concentration (TBil) and alanine aminotransferase activity (ALAT) were determined pre-operatively, on post-operative day 3, weekly for one month and monthly for 3 months.

An autopsy was performed whenever a rat died to identify possible complications. The livers were removed at sacrifice and fixed with 10% formalin. Four-micron sections were stained with hematoxylin and eosin (HE) or trichrome. Liver samples were studied by light microscopy.

#### **Statistical Analysis**

The data were expressed as the mean  $\pm$  standard deviation (SD). The Student's *t*-test was applied for the data analysis. *P* values equal to or less than 0.05 were considered statistically significant.

### **RESULTS**

#### **Body Weight and Animal Survival**

The body weight of all rats declined 24 hours after the operation with average  $10.00 \pm 6.47$  grams per rat in group 4 and  $18.33 \pm 14.03$  grams in group 5 (NS). Three weeks later, the rats of group 4 regained their pre-

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operative weight and even exceeded up to average  $24.07 \pm 3.40$  grams compared with  $1.67 \pm 11.16$  grams in group 5 (NS).

**Table 1.** Experimental groups and post-operative survival

Experimental groups	Survival time				
G1 ( <i>n</i> = 10): PCS	> 3 months (10/10)				
G2 ( <i>n</i> = 10): PCS + 68% PH	0.81 ± 0.56 days (mean ± <i>SD</i> )				
G3 ( <i>n</i> = 10): PCS + 68% PH + CBDL	0.51 ± 0.30 days (mean ± <i>SD</i> )				
	Survival rates				
	1 week months	2 weeks	3 weeks	1 month	2 months
G4 ( <i>n</i> = 12): HALTx + 68%	100%	100%	92%	92%	83%
PH	100%	83%	58%	50%	50%
G5 ( <i>n</i> = 12): HALTx + 68%					
PH + CBDL					

Notes: PCS, portacaval shunt; 68% PH, 68% partial hepatectomy; CBDL, common bile duct ligation; HALTx, heterotopic auxiliary liver transplantation.

As shown in the *Table 1*, the rats of group 1 could well tolerate a PCS whereas all rats in group 2 and 3 died of liver failure within two days after the surgery.

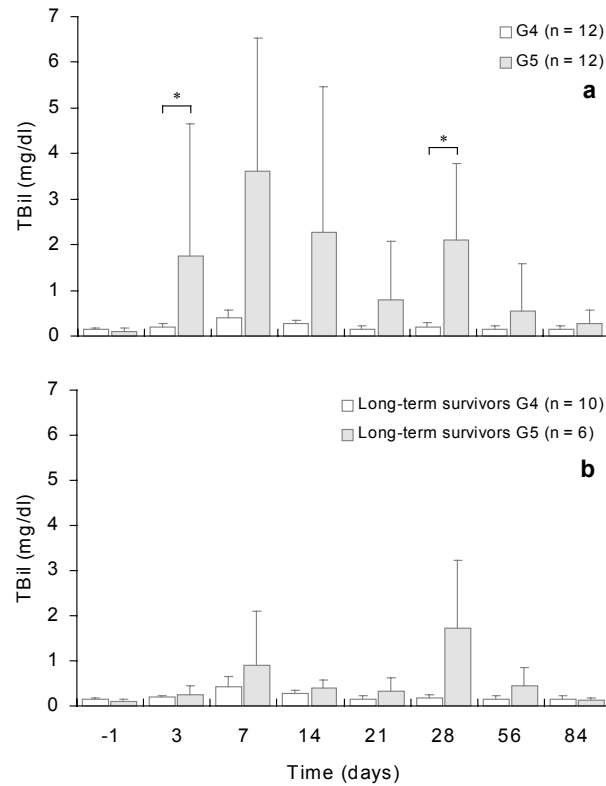
In group 4, the 1-month survival rate was as high as 92%. Only two rats died of abscess in the native liver or in both the native and the engrafted livers on the 17<sup>th</sup> and 34<sup>th</sup> post-operative day. In group 5, only 50% of the animal survived longer than 3 months. Except a rat that died of rejection on day 23, all others died of infectious complications (cholestasis in 1, abscess in 3 and peritonitis in 1).

### Serum Biochemical Parameters

Serum TBil of the rats in group 5 was significantly higher than those of group 4 on post-operative day 7 ( $p = 0.025$ ) and 28 ( $p = 0.02$ ) (*Figure 1a*). The rats (2<sup>#</sup>, 3<sup>#</sup>, 8<sup>#</sup> and 10<sup>#</sup>) of group 5, that had the highest values of TBil on day 7, developed infectious complications and died within the following weeks. By disregarding the values of those rats that died of complications, the differences between the two groups were still recorded but they were no longer statistically significant (*Figure 1b*). The high TBil peak of group 5 on

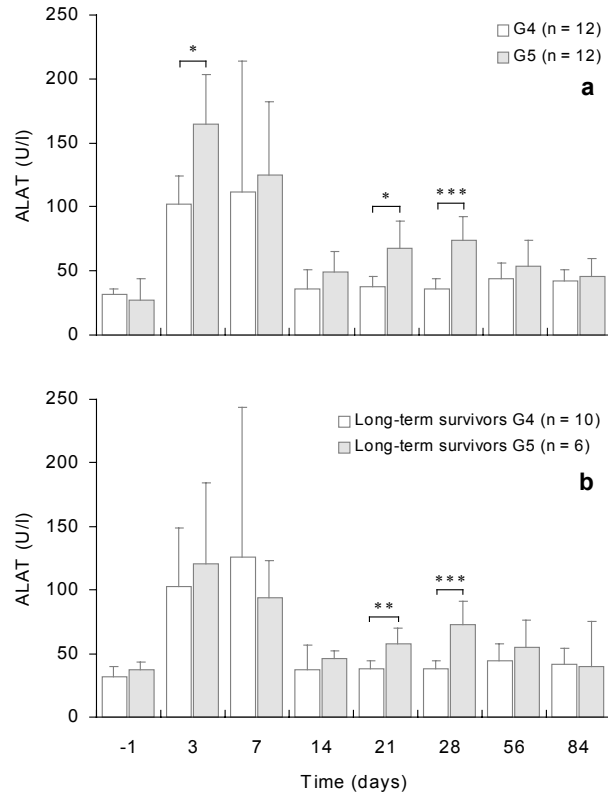
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day 28 was this time due to the two rats that developed secondary biliary cirrhosis of the engrafted livers found later at autopsy.



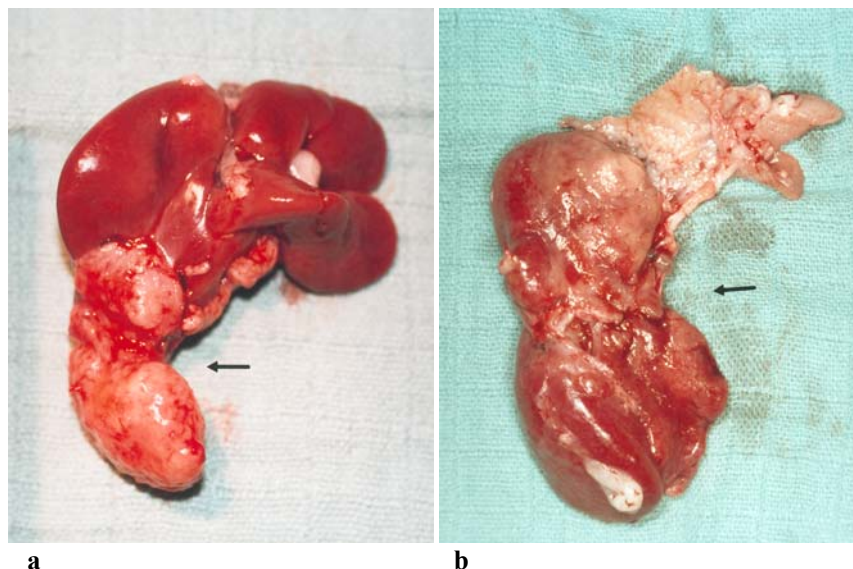
**Figure 1.** Comparison of the mean serum total bilirubin concentration (TBil) between group 4 and 5 post-HALTx: **(a)** Data of the whole group. Statistically significant differences were found on day 7 and day 28 ( $p < 0.05$ ); **(b)** Data of the long-term survivors. No significant differences between the two groups could be seen.

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**Figure 2.** Comparison of the mean serum alanine aminotransferase (ALAT) between group 4 and 5 post-HALTx: **(a)** Data of the whole group. The statistically significant differences were found on day 3 ( $p < 0.05$ ), day 21 ( $p < 0.05$ ) and day 28 ( $p < 0.001$ ); **(b)** Data of the long-term survivors. The differences between the two groups were not significant on day 3 and remained significant on day 21 ( $p < 0.01$ ) and 28 ( $p < 0.001$ ).

As showed in *Figure 2a*, significant differences of mean serum ALAT between the two groups were found on day 3 ( $p = 0.014$ ), 21 ( $p = 0.014$ ) and 28 ( $p < 0.001$ ). Similarly to serum TBil, the highest ALAT values of group 5 on day 3 were again due to the rats that later died of infectious complications. When those values were excluded, the differences were no longer significant on day 3 but remained significant on day 21 ( $p = 0.008$ ) and 28 ( $p < 0.001$ ).



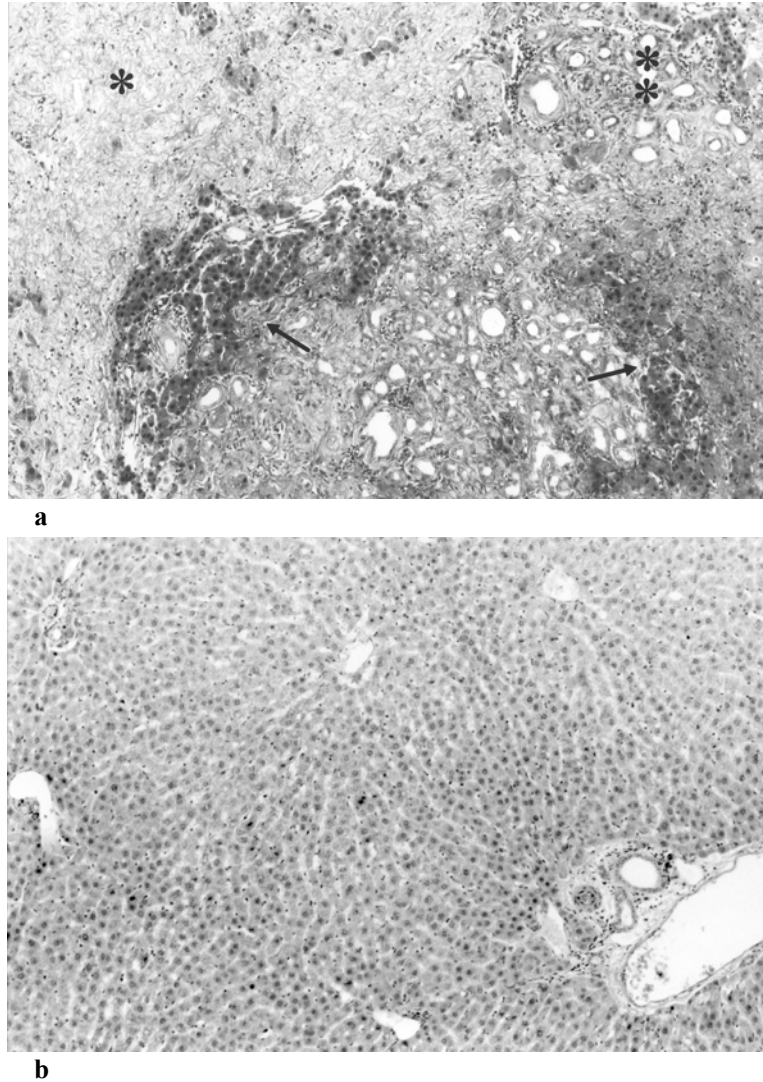
**Figure 3:** Autopsy. **(a)** a rat of group 4 (HALTx with a 68% PH to handicap the native liver) sacrificed 7 months after the operation: the engrafted liver (↗) atrophied whereas the native liver had normal appearance; **(b)** a rat of group 5 (HALTx with a 68% PH and a CBDL to handicap the native liver) sacrificed 12 months after the operation: the size of the engrafted liver (↗) enlarged and the native liver underwent complete atrophy.

### Autopsy and Histology

After the end of three months observation period, the long-term survival rats were killed at different time intervals. It was found that all the engrafted livers of the 10 survivors in group 4 underwent atrophy at sacrifice while their native livers showed normal appearance (*Figure 3a*). The general architecture of those engrafted livers was altered by fibrosis and ductular metaplasia. Only a few groups of hepatocytes were present (*Figure 4a*).

On the contrary, in group 5, the engrafted livers of all but 2 long term survivors appeared normal macroscopically whereas their native livers showed clear atrophy (*Figure 3b*). The general architecture of those engrafted livers was well preserved without major histological alterations (*Figure 4b*). The 2 remaining rats (1<sup>#</sup> and 6<sup>#</sup>) presented cirrhotic appearance of their engrafted livers and the histology revealed secondary biliary cirrhosis due to sub-obstruction of their hepaticoduodenostomy.

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**Figure 4:** Histological observations. **(a)** a rat of group 4 (HALTx with a 68% PH to handicap the native liver) sacrificed 7 months after the operation: the general architecture of the engrafted liver was altered by fibrosis (\*) and ductular metaplasia (\*\*). Only a few groups of hepatocytes were present (↗) (Trichrome, x 10); **(b)** a rat of group 5 (HALTx with a 68% PH and a CBDL to handicap the native liver) sacrificed 12 months after the operation: the general architecture of the engrafted liver was well preserved and no major alterations were present (HE, x 10).



## DISCUSSION

In the previous studies of rat HALTx, the post-operative animal survival and serum biochemistry were most commonly analysed without taking into account the role of the handicap of the native liver. In this study, we show for the first time how these handicaps can differently influence the post-transplant biochemical parameters.

The classical rat model of HALTx includes a complete shunting of the portal blood away from the native liver to re-vascularize the engrafted liver. In this case, the blood supply of the native liver is solely given by the hepatic artery, which corresponds effectively to a PCS of the liver. It appears that the blood flow of the hepatic artery can compensate the by-passed portal blood after a PCS and maintain a “functioning liver”, so that the effects of PCS do not significantly influence the animal survival. Without handicap of the native liver, the engrafted liver is therefore transplanted parallel with a normal functioning liver. The increased hepatic blood flow will not be sufficient to support proper hepatic function if a further handicap is carried out in the native liver. It is reported that in the case of a 68% PH added to an end-to-side PCS, the rat survival does not exceed 2 days <sup>11</sup>. The results of the present study confirm this data and show, as would have been anticipated, that the combination of a PCS, a 68% PH and a CBDL does even shorten this survival time. In both these circumstances, no animal can be kept alive unless an auxiliary liver graft is transplanted.

The long-term survivors in the HALTx groups make it possible to perform a comparative study of the serum biochemistry between the rats bearing different liver handicaps. When the rats were handicapped only with a 68% PH as in group 4, ALAT increased to a peak level 1 week post-transplant but then returned to normal values after 2 weeks reflecting transitory hepatocellular damage. Both ischemic injury of the graft caused by preservation and the surgical manipulations of the liver at the time of 68% PH are responsible for this initial elevation. On the other hand, TBil of this group only rose slightly after the surgery even in the rats that presented fatal complications. This demonstrates that in case of a sole 68% PH to handicap the native liver, either one of the two livers can still compensate for the functional deficiency of the other. An additional CBDL to the native liver will handicap this organ so profoundly that it can no longer compensate when the engrafted liver is affected by an acute infectious complication: TBil sharply increased in these cases followed by the animal deaths shortly later as shown in group 5. The “double handicap” of the native liver is also responsible for a second peak of TBil around 1 month after HALTx. The rats that developed secondary biliary cirrhosis of the graft presented the highest

#### 74 3.1. Handicap of Native Liver in Rat HALTx

values at this time point, when the exhaustion of the regeneration potential and thereby the functional failure of the native liver occurred <sup>19</sup>. The repermeabilisation of the extrahepatic biliary duct of the native liver may presumably explain the normalisation of serum TBil in the long-term survivors.

It has been found that long-standing bile duct obstruction of the native liver impairs the function of reticuloendothelial system, which might lead to serious post-operative complications <sup>20, 21</sup>. The rats handicapped with CBDL (group 5) had indeed a higher incidence of infection and more pronounced post-operative weight-loss. Cholestasis induced by CBDL also inhibits the metabolic functions of liver cells including mitochondrial function. As consequence, regeneration capacity of this organ is compromised <sup>22, 23</sup>. These may be the reasons why a CBDL together with a 68% PH can affect the native liver to such a degree that the post-HALTx animal survival and normal serum biochemical values depend totally upon the functionality of the engrafted liver.

HALTx has advantages over OLTx in treating acute liver failure, inborn errors of metabolism and, exceptionally, chronic liver diseases with high operative risk for OLTx. In the case of acute liver failure, the diseased native liver is expected to recover while the engrafted liver is providing functional support. The engrafted liver will then be removed or left in place to atrophy if the native liver resumes its proper function <sup>1-3</sup>. For patients with chronic liver diseases, regeneration of the engrafted liver and atrophy of the native liver is the rule after a HALTx <sup>24</sup>. Although up to now no clinical experience with HALTx to treat patients with inborn errors of metabolism has been reported, the long-term functionality of both the engrafted and the native livers should be looked for <sup>4</sup>. In view of these different situations, it is indeed important to have a better understanding about the mechanisms of inter-liver competition and the right approaches to control or to modify this process. It is for this purpose, the rat HALTx can be an interesting model to study the insight of this phenomenon, even though it does not replicate the exact clinical situation. It is generally accepted that proper regeneration of the liver can only be achieved when the portal vein is re-vascularized with venous blood or through arterialisation of the portal stump <sup>6, 15, 25</sup>. Interestingly, the partially resected native liver can still regain progressively its volume and function despite the lack of portal blood flow, whereas the engrafted liver eventually atrophies. In case of an additional handicap of the native liver, by means of a CBDL, the functional recuperation of the native liver is completely impaired while the engrafted liver takes over the function and keeps its normal macroscopic and microscopic aspects. These results suggest that besides the known hepatotrophic factors <sup>8, 26</sup>, the diseased state of the

native liver also influences the outcome of the inter-liver competition. Yu and co-workers have shown that the atrophy of the engrafted liver can be prevented by its re-arterialisation even when the native liver is not handicapped<sup>5</sup>. Re-arterialisation of the graft in the OLTx model reduces biliary complications, modifies the immunological response and improves micro-vascular perfusion of the graft<sup>27-29</sup>. In the rat model of HALTx, these beneficial effects certainly favour the competitive position of the engrafted liver.

The data of the present study show that in the rat model of HALTx the different manipulations on the native liver do influence not only the post-transplant animal survival but also the serum biochemical values of the liver. Furthermore, handicap of the native liver limited to a 68% PH will lead to the atrophy of the engrafted liver, whereas a 68% PH combined with a CBDL is a handicap that impairs the regeneration of the native liver. In the latter case, the post-HALTx survival and normalisation of the serum biochemistry depend solely upon the viability of the engrafted liver. Although this model has no direct similarity with clinical condition, it allows further approach and understanding to the concept of inter-liver competition.

## REFERENCES

1. Moritz, M. J., Jarred, B. E., Armenia, V., Radomski, J., Carabasi, R. A., Zeitoun, G., Columbus, K., Rubin, R., Smaddrey, W. (1990) Heterotopic liver transplantation for fulminant hepatic failure- a bridge to recovery. *Transplantation*, **50-3**, 524-526.
2. Van Hoek, B., Ringers, J., Kroes, A. C., van Krieken, J. H., van Schelven, W. D., Masclee, Ad., van Krieken-Hogenberk, L. G., Haak, H. R., Lamers, C. B., Terpstra, O. T. (1995) Temporary heterotopic auxiliary liver transplantation for fulminant hepatitis B. *Journal of Hepatology*, **23**, 109-118.
3. Boudjema, K., Cherqui, D., Jaeck, D., Chenard-Neu, M. P., Steib, A., Freis, G., Becmeur, F., Brunot, B., Simeoni, U., Bellocq, J. P., Tempe, J. D., Wolf, P., Cinqualbre, J. (1995) Auxiliary liver transplantation for fulminant and subfulminant hepatic failure. *Transplantation*, **59-2**, 218-223.
4. Madern, G. C., Terpstra, O. T., Sinaasappel, M., Provoost, A. P., Rothuizen, J., Molenaar, J. C. (1991) Heterotopic liver transplantation corrects the inborn error of hepatic metabolism in a dog model. *Transplantation Proceedings*, **23-1**, 716-717.
5. Yu, W. Y., Wan, X. Y., Wright, J. R., Coddington, D., Bitter-Suermann, H. (1994) Heterotopic liver transplantation in rats: Effects of intrahepatic islet isografts and split portal blood flow on liver integrity after auxiliary liver isotransplantation. *Surgery*, **115-1**, 108-117.
6. Lee, S., Edgington, T. S. (1966) Liver transplantation in the rat. *Surgical*

## 76 3.1. Handicap of Native Liver in Rat HALTx

*Forum*, **17**, 220-222.

7. Lee, S., Edgington, T. S. (1968) Heterotopic liver transplantation utilising inbred rat strains: I. Characterisation of allogeneic graft rejection and the effects of biliary obstruction and portal vein circulation on liver regeneration. *American Journal of Pathology*, **52-3**, 649-669.
8. Bestian, J. M., Janin, A., Zenner, L., Pruvot, F. R., Zelus, D., Courtade, A., Dessaint, J. P., Capron, A. (1995) Allogenic microchimerism following auxiliary heterotopic liver transplantation in rat and swine. *Transplantation Proceedings*, **27-2**, 1675.
9. Marni, A., Ferrero, M. E. (1985) Heterotopic liver grafting in the rat: A simplified method using cuff techniques. *Transplantation*, **39-3**, 329-331.
10. Kort, W. J., Wolff, E. D., Eastham, W. N. (1971) Heterotopic auxiliary liver transplantation in rats. *Transplantation*, **12-6**, 415-420.
11. Hess, F., Jerusalem, C., van der Heyde, M. N. (1972) Advantages of auxiliary liver homo-transplantation in rats. *Archives Surgery*, **104**, 76-80.
12. Müller, G. (1983) A simple technique for heterotopic auxiliary liver transplantation in the rat. *Transplantation*, **36-2**, 221-222.
13. Starzl, T. E., Marchioso, T. L., Huntley, R. T. (1964) Experimental and clinical homo-transplantation of the liver. *American New York Academic Science*, **120**, 739-765.
14. Marchioro, T. Z., Poster, K. A., Dickinson, T. C., Faris, T. D., Starzl, T. E. (1965) Physiologic requirements for auxiliary liver transplantation. *Surgery, Gynecology and Obstetrics*, **121**, 17-31.
15. Van der Heyde, M. N., Schalm, L., Vink, M. (1967) The role of functional competition in auxiliary liver transplantation. *Transplantation*, **5**, 78-80.
16. Higgins, G. M., Anderson, R. M. (1931) Experimental pathology of the liver: I. Restoration of the liver of the white rat following partial surgical removal. *Archives Pathology*, **12**, 186-202.
17. Lee S, Fisher B: Portacaval shunt in the rat. *Surgery* 1961; 50:668-672.
18. Kamada, N., Calne, R. Y. (1983) A surgical experience with five hundred thirty liver transplantation in the rat. *Surgery*, **93-1**, 64-69.
19. Gross, J. B., Reichen, J., Zeltner, T., Zimmermann, A. (1987) The evolution of changes in quantitative liver function tests in a rat model of cirrhosis: correlation with morphometric measurement of hepatocyte mass. *Hepatology*, **7**, 457-63.
20. Holman, J. M. Jr., Rikkers, L. F. (1982) Biliary obstruction and host defence failure. *Journal of Surgical Research*, **32**, 208-213.
21. Andersson, R., Foss, A. (1991) Abdominal sepsis following liver resection in the rat. *Hepatogastroenterology*, **6**, 547-549.
22. Koyaman, K., Takagi, Y., Ito, K., Sato, T. (1981) Experimental and clinical studies on the effect of biliary drainage in obstructive jaundice. *American Journal of Surgery*, **142**, 293-299.
23. Miyata, K. (1983) Delayed recovery of mitochondrial function in rat liver after

releasing biliary obstruction. *Nagoya Journal of Medical Science*, **45**, 97-105.

24. Terpstra, O. T., Schalm, S. W., Weimar, W., Willemse, P., Baumgartner, D., Groenland, T., ten Kate, F., Porte, R. J., De Rave, S., Reuvers, C. B., Stibbe, J., Terpstra, J. L. (1988) Auxiliary partial liver transplantation for end-stage chronic liver disease. *New England Journal of Medicine*, **319**, 1507-1511.
25. de Hemptinne, B. (1980) Acute effects of portacaval shunt and arterialization on rat hepatic regeneration. *European Surgical Research*, **12 (suppl 1)**, 3.
26. Terpstra, O. T., Reuvers, C. B., Schalm, S. W. (1988) Auxiliary heterotopic liver transplantation. *Transplantation*, **45**, 1003-1007.
27. Howden, B., Jablonski, P., Grossman, H., Marshall, V. C. (1989) The importance of the hepatic artery in rat liver transplantation. *Transplantation*, **47**, 428-431.
28. Engemann, R., Ulbrichs, K., Thiede, A., Müller-Ruchholtz, W., Hamelmann, H. (1982) Value of a physiological liver transplant model in rats. *Transplantation*, **33**, 566-568.
29. Post, S., Menger, M. D., Rentsch, M., Gonzalez, A. P., Herfarth, C., Messmer, K. (1992) The impact of arterialization on hepatic microcirculation and leukocyte accumulation after liver transplantation in the rat. *Transplantation*, **54**, 789-794.

### **3.2. Effects of Re-Arterialisation on Early Graft Function and Regeneration in the Rat Model of Heterotopic Auxiliary Liver Transplantation**

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#### **SUMMARY**

In the rat model of heterotopic auxiliary liver transplantation, graft re-arterialisation may influence the outcome of inter-liver competition. This was investigated in the current study using two transplanted groups with or without graft re-arterialisation. Immediately after re-perfusion, the re-arterialised grafts showed significantly higher bile flow rate and bilirubin excretion than the grafts without re-arterialisation. DNA synthesis rate was also increased more drastically in the re-arterialised group following the transplantation. Without re-arterialisation, the rats developed more pronounced cytolysis and cholestasis. Among the long-term survivors, all healthy re-arterialised grafts regenerated, whereas 5/6 non re-arterialised grafts atrophied. These data demonstrate that the re-arterialisation does increase graft survival by improving early hepatic function, enhancing regenerative response and preventing post-transplant biliary complications in this rat model.

CHAPTER 3 Study of Inter-liver Functional Competition **77**

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## **INTRODUCTION**

In the rat model of heterotopic auxiliary liver transplantation (HALTx), the coexistence of the graft and the recipient's native liver can induce functional inhibition and lead to atrophy of the newly engrafted liver [1]. Besides the role of hepatotrophic factors in maintaining hepatocyte integrity [2], other mechanisms may as well have an influence on the inter-liver competition. In a previous study, we showed that a certain degree of handicap on the native liver was needed to weaken the functional position of the native liver and to prevent atrophy of the graft [3]. It was evidenced that if the native liver was kept intact, the engrafted liver did eventually atrophy and was replaced by connective tissue [4]. By adding a hepatic re-arterialisation to the graft, atrophy could be avoided and regeneration of the engrafted liver was achieved [5].

The studies of orthotopic liver transplantation have demonstrated the beneficial effects of the graft re-arterialisation on improving micro-vascular perfusion [6], decreasing the expression of class I and II MHC antigens [7], maintaining an adequate oxygen supply and tissue ATP [8], reducing the post-transplant biliary complications [9, 10] and increasing animal survival rate [11]. Whether equal mechanisms might be implicated in HALTx remained to be analysed, as the interaction between the graft and the native liver added an important element to this specific rat model.

The current study was conducted to investigate the possible impact of the re-arterialisation on immediate graft function, regenerative response of the engrafted liver and post-operative graft survival and long-term outcome of the recipient animal in the rat model of HALTx.

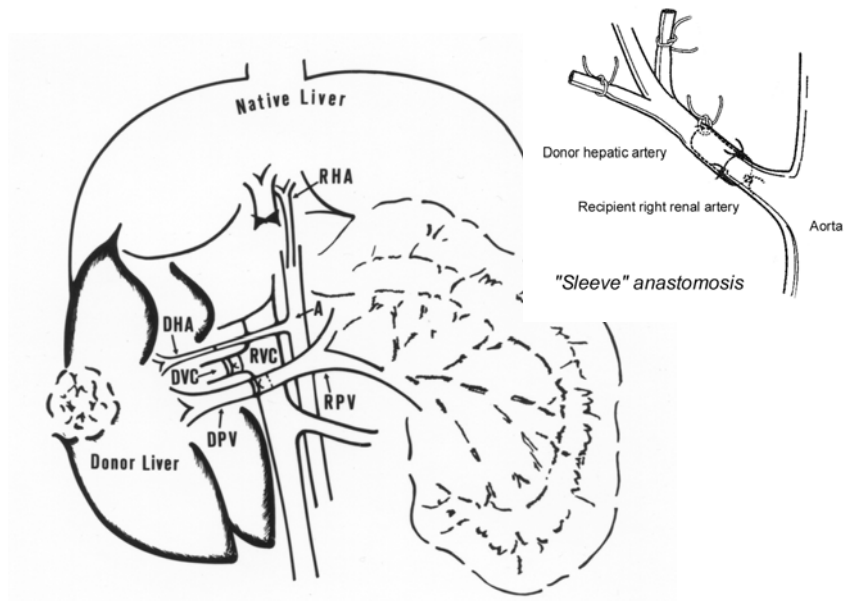
## **MATERIALS AND METHODS**

### **Animals and Experimental Groups**

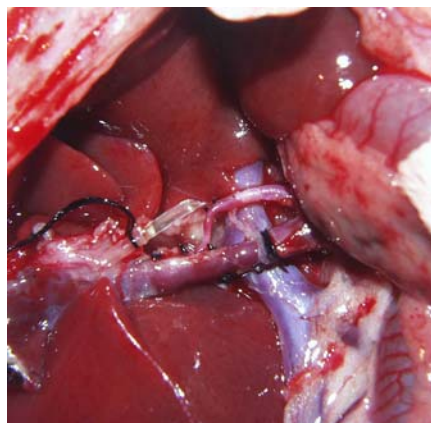
Male inbred Lewis rats (Harlan Netherlands B. V., Zeist, the Netherlands) weighing 240-300 g were used in this study. The rats had free access to water and standard pellet food. Housing, humidity and day-night circle were strict according to European guidelines for animal care. Before surgery, food but not water was withdrawn from the recipient rats for 12 hours. Donors and recipients were matched for size.

Two experimental groups were studied: HALTx-A group, HALTx with re-arterialisation of the engrafted liver ( $n = 58$ ); HALTx-nonA group, HALTx without re-arterialisation of the engrafted liver ( $n = 58$ ).





**Figure 1.** Technique of heterotopic auxiliary liver transplantation with graft re-arterialisation. The portal vein of the donor liver (DPV) was anastomosed with the portal vein of the recipient (RPV). Using "sleeve" technique, the hepatic artery of the donor liver (DHA) was re-connected to the recipient's aorta (A) through the right renal artery. The venous drainage of the donor liver was made between the donor vena cava (DVC) and the recipient vena cava (RVC) via the right renal vein. The recipient native liver received sole hepatic artery supply (RHA).



## **80** 3.2. Graft Re-arterialisation in Rat HALTx

### **Surgical Procedure**

All operations were carried out with ether anaesthesia, clean but non-sterile instruments and x 12.5 magnification. Body temperature of the rats was maintained at approximately 36°C by using heating pads.

Using 32% of the donor liver, HALTx was performed according to the technique as previously described [12]. The donor liver was perfused with cold ( $\pm 4^{\circ}\text{C}$ ) heparinized Ringer's solution: 5 ml via the portal vein and 3 ml via the hepatico-aortic segment. Following a right nephrectomy, the liver graft was implanted in the right paravertebral gutter under the native liver of the recipient. The cuff technique was used for the anastomoses of both the vena cava and the portal vein. The engrafted liver received its portal blood from the portal vein of the recipient. The venous drainage was made through the right renal vein of the recipient to the vena cava. In the HALTx-A group, the hepatic artery of the engrafted liver was re-connected to the recipient's right renal artery with a 10-0 monofilament nylon suture (Tyco Health Care, Belgium) using the "sleeve" technique [13]. The engrafted liver was then re-perfused simultaneously by portal and arterial blood [14]. As the total amount of portal flow went to the engrafted liver, the vascularization of the native liver was limited to sole hepatic artery supply (*Figure 1*). Restoration of the bile drainage was done by insertion of the intubated common bile duct of the donor liver into the recipient's duodenum.

The duration (mean  $\pm$  SD) of cold ischemia of the liver grafts was  $56 \pm 7$  minutes in the HALTx-A group and  $55 \pm 8$  minutes in the HALTx-nonA group. The duration of warm ischemia of the liver grafts (starting from implantation and ending at blood re-circulation of the engrafted liver) was  $25 \pm 3$  minutes in the HALTx-A group and  $25 \pm 1$  minutes in the HALTx-nonA group.

### **Analysis of Bile Production**

Six HALTx recipients in each experimental group were used to evaluate intra-operative bile secretion. Immediately after the blood re-circulation of the engrafted liver, a tubing of PE-50 was cannulated into the graft common bile duct. The bile production of the initial 15 minutes after re-perfusion was disregarded to obtain a stabilised outflow. Thereafter, the bile was collected at 15-minute intervals up to 60 minutes. All the samples were then frozen for the analysis of bilirubin concentration, which was expressed as bilirubin excretion ( $\mu\text{g}/\text{min}/\text{g}$  liver).

### **Measurement of DNA Synthesis Rate**

DNA synthesis rate was measured as an index of liver regeneration response

on post-operative day 1, 2, 3, 7 and 14. At each time point, 8 HALTx recipients of each experimental group were sacrificed under ether anaesthesia.  $^3\text{H}$  thymidine (specific activity 26 Ci/mmol, Nycomed Amersham plc, England) of 0.2 mCi/g body weight was injected intravenously 2 hours before sacrifice. The patency of the hepatic arterial anastomosis, whenever relevant, was examined. The liver was then flushed with cold physiological saline via the portal vein. The right lobes of the engrafted liver were frozen in liquid nitrogen for determining the DNA synthesis rate by the incorporation rate of  $^3\text{H}$  thymidine into nuclear DNA, while the left lobes and the recipient's native livers were kept in formalin for histological evaluation. The DNA fraction was extracted following the procedures described elsewhere [15]. The radioactivity of  $^3\text{H}$  thymidine was measured in a liquid scintillation counter and the DNA concentration of the same homogenate was determined by the standard method of colorimetric readings at 700 nm, using calf thymus DNA (Boehringer Mannheim GmbH, Germany) as the standard. The DNA synthesis rate was expressed as d.p.m.  $\times 10^{-3}/\mu\text{g}$  DNA.

#### **Post-operative Follow-ups**

Twelve rats in each group were used to evaluate post-operative liver biology, graft and animal survival. Blood samples were taken on post-operative day 1, 2, 3, 7 and weekly thereafter. Serum alanine aminotransferase activity (ALAT), total bilirubin concentration (TBil) and gamma glutamyl transferase activity (GGT) were measured using routine photometric tests on a Hitachi 747 analyser. An autopsy was performed whenever a rat died and all the long-term survivors were sacrificed at the end of the third post-transplant month for histological examination. The liver samples were fixed in 10% formalin. Four-micron thick sections were stained with hematoxylin and eosin (HE) or trichrome and examined under light microscope.

#### **Statistics**

The data were expressed as the mean  $\pm$  standard deviation (SD). The distribution of the data was checked by Kolmogorov test. The Student's *t*-test or the Mann-Whitney rank sum test was applied for normally or not-normally distributed data, respectively. *P* values equal to or less than 0.05 were considered statistically significant.

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**Table 1.** Intra-operative bile flow rate and bilirubin excretion of the engrafted liver after reperfusion in the HALTx-A and HALTx-nonA groups

	HALTx-A ( <i>n</i> = 6)	HALTx-nonA ( <i>n</i> = 6)	<i>p</i>
Bile flow rate (μl/min/g liver)			
15 min	1.65 ± 0.09	0.97 ± 0.13	< 0.001
30 min	1.78 ± 0.27	0.97 ± 0.10	< 0.001
45 min	1.84 ± 0.30	0.99 ± 0.09	< 0.001
60 min	1.89 ± 0.39	0.97 ± 0.03	< 0.001
Bilirubin excretion (μg/min/g liver)	0.06 ± 0.02	0.02 ± 0.01	0.007

Note: Data are expressed as mean ± SD.

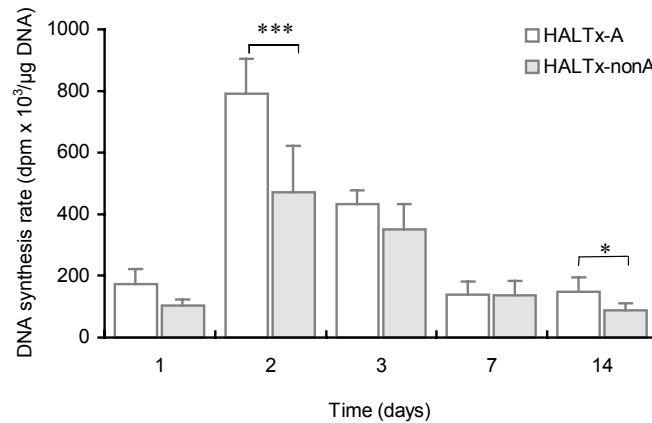
## RESULTS

### Intra-operative Bile Production

As shown in the *Table 1*, immediately after the blood re-circulation through the engrafted liver, the bile flow rate of the grafts with re-arterialisation was significantly higher than that of the non re-arterialised grafts ( $1.65 \pm 0.09$  vs.  $0.97 \pm 0.13$  μl/min/g liver,  $p < 0.001$ ). This remained throughout the observation period of 60 minutes ( $p < 0.001$ ). During this period, the mean bilirubin excretion of the HALTx-A group was more than 3 times higher than that of the HALTx-nonA group ( $0.06 \pm 0.02$  vs.  $0.02 \pm 0.01$  μg/min/g liver,  $p = 0.007$ ).

### DNA Synthesis Rate

In the two experimental groups, the incorporation rate of  $^3\text{[H]}$  thymidine into nuclear DNA increased after the transplantation and showed a peak value on the post-operative day 2. The mean value of DNA synthesis rate in the HALTx-A group was significantly higher than that in the HALTx-nonA group ( $791 \pm 113$  vs.  $471 \pm 151$  d.p.m.  $\times 10^{-3}/\mu\text{g DNA}$ ,  $p < 0.001$ ). These peak values fell sharply after day 2 and reached residual low levels on day 7. DNA synthesis rates remained steady thereafter with however progressive decline in the non re-arterialised group. On day 14, the values of the 2 groups were  $149 \pm 45$  (HALTx-A) and  $87 \pm 24$  (HALTx-nonA) d.p.m.  $\times 10^{-3}$ .



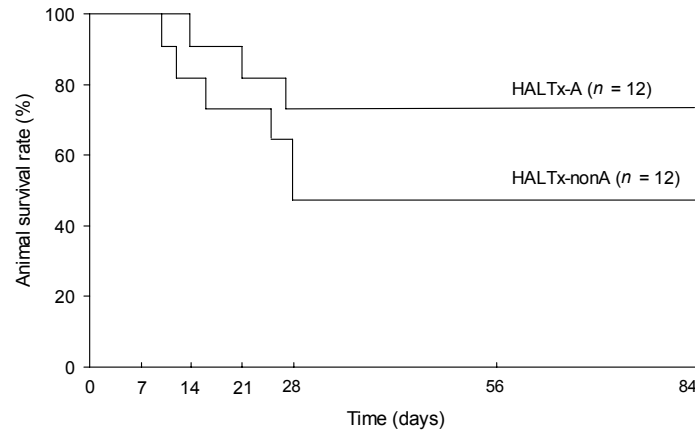
<sup>3</sup>/μg DNA ( $p < 0.05$ ) **Figure 2.** Post-HALTx course of DNA synthesis rate of the engrafted livers (mean  $\pm$  SD) in the re-arterialised group (HALTx-A) and the non re-arterialised group (HALTx-nonA). \*  $p < 0.05$ , \*\*\*  $p < 0.001$ .

(Figure 2).

### Post-operative Follow-ups

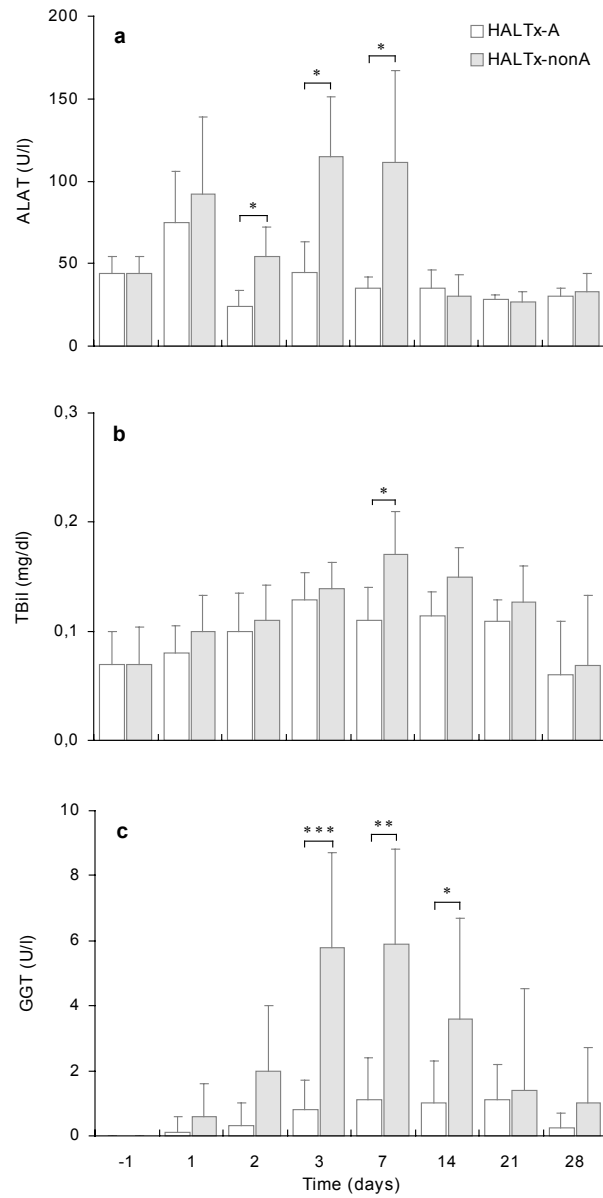
In the HALTx-A group, 9 out of 12 rats survived the observation period of three months. One rat died of cholangitis, and another 2 died of abscesses and parenchymal necrosis in the engrafted liver, the latter of which was due to thrombosis of the portal vein and the vena cava. Among the 12 HALTx-nonA rats, 6 died during the follow-up period. Post-mortem examination revealed cholangitis in 3 rats and complete necrosis of the engrafted livers in another 3. This resulted in a 3-month survival rate of 75% in the HALTx-A group compared to 50% in the HALTx-nonA group (Figure 3).

### 84 3.2. Graft Re-arterialisation in Rat HALTx



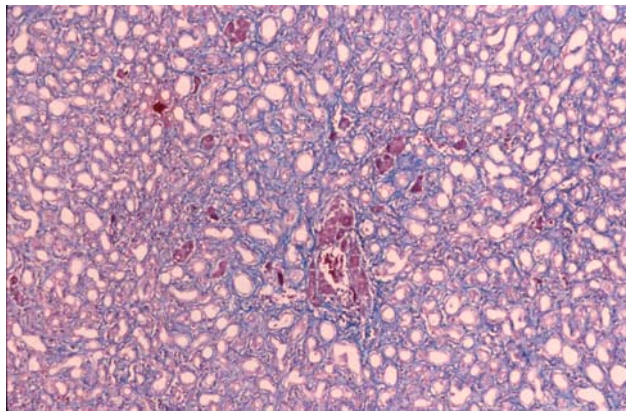
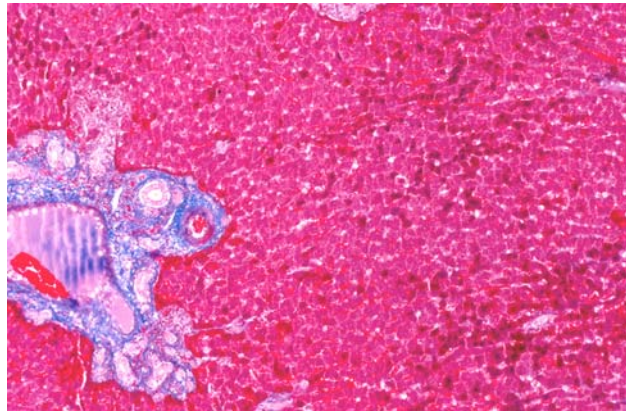
**Figure 3.** Three-month survival rate after HALTx with graft re-arterialisation (HALTx-A) and without graft re-arterialisation (HALTx-nonA).

After the transplantation, the levels of serum ALAT activity were elevated in the rats of both groups on day 1 and declined on day 2. Thereafter, the mean values of the re-arterialised rats remained in the normal range, while the rats of the HALTx-nonA group showed drastically increased values. The differences between the HALTx-A and HALTx-nonA group were significant on post-operative day 2 ( $24 \pm 10$  vs.  $54 \pm 18$  U/l,  $p < 0.05$ ), day 3 ( $45 \pm 18$  vs.  $115 \pm 36$  U/l,  $p < 0.05$ ) and day 7 ( $35 \pm 7$  vs.  $111 \pm 56$  U/l,  $p = 0.01$ ) (Figure 4a). The post-operative serum TBil concentration increased in the rats of both groups and, however, the re-arterialised rats had lower mean TBil values and sooner normalisation compared with that of the rats of the other group. On day 7, the re-arterialised rats had a mean TBil value of  $0.11 \pm 0.03$  mg/dl, which was significantly lower than  $0.17 \pm 0.04$  mg/dl of the non re-arterialised rats ( $p = 0.01$ ) (Figure 4b). Serum GGT activity showed even more pronounced differences: mean GGT activity of the rats in the HALTx-nonA group was 5 times higher than that of the re-arterialised rats on the post-operative day 3 and 7. GGT activity of the re-arterialised group was only slightly elevated after transplantation, while this parameter was profoundly altered in the other group (day 3,  $0.8 \pm 0.9$  vs.  $5.8 \pm 2.9$  U/l,  $p < 0.001$ ; day 7,  $1.1 \pm 1.3$  vs.  $5.9 \pm 2.9$  U/l,  $p = 0.002$  and day 14,  $1.0 \pm 1.3$  vs.  $3.6 \pm 3.1$  U/l,  $p = 0.04$ ) (Figure 4c).



**Figure 4.** Post-HALTx evolution of serum ALAT (mean  $\pm$  SD; **a**), TBil (**b**) and GGT (**c**) in the HALTx-A group and the HALTx-nonA group. \*  $p \leq 0.05$ , \*\*  $p < 0.01$  and \*\*\*  $p < 0.001$ .

**86** 3.2. Graft Re-arterialisation in Rat HALTx



**a.**  
**b.**

**Figure 5.** Histological observations. **(a)** A rat of HALTx-A group sacrificed 3 months after the transplantation: the general lobular architecture of the engrafted liver was well preserved with a patent hepatic artery (Trichrome staining, original magnification x 100). **(b)** A rat of HALTx-nonA group sacrificed at the end of 3<sup>rd</sup> post-transplant months: the general architecture of the engrafted liver was completely destroyed and replaced by massive ductal metaplasia. Only remnants of hepatocytes were rarely seen around the portal tract (Trichrome staining, original magnification x 100).

At the end of the 3-month follow-up period, amongst 9 long-term survivors of the HALTx-A group, five showed enlarged grafts with well-preserved general lobular architecture (*Figure 5a*), while their native livers were



atrophic. The engrafted livers of the other 4 rats were remained unchanged in size or smaller. Portal vein thrombosis was found in one case and intra-hepatic or intra-peritoneum abscesses were present in the other 3 rats. In contrast, the atrophy of the engrafted liver was found in 5 out of 6 long-term survivors in the HALTx-nonA group, while their native livers appeared normal. Histology revealed that massive ductular metaplasia altered the general architecture of those engrafted livers (*Figure 5b*). Only one graft did not show complete atrophy (engrafted liver: 4.1 g; native liver: 6.7 g) and in this case the engrafted liver had contracted extensive adhesions with the omentum.

## DISCUSSION

After a long-lasting debate, the importance of re-arterialisation for the rat model of orthotopic liver transplantation has generally been accepted. The possible impact of graft re-arterialisation on the rat model of HALTx was investigated in the present study with special focus on immediate graft function and post-transplant regenerative response.

The influence of re-arterialisation on early graft function was assessed by the initial intra-operative bile production after re-perfusion of the engrafted liver. This parameter is known to be more indicative for early post-operative hepatic function than standard liver biology [16]. Since bile production is an ATP and an O<sub>2</sub>-dependent process [17], it is expected that the hepatic artery should play an important role by maintaining an appropriate intra-hepatic pO<sub>2</sub>. The data of this study show that indeed the re-arterialisation of the engrafted liver leads to a significantly higher quantity and quality of bile production of the graft indicating an improved early graft function.

Post-operative biliary complications following non re-arterialised orthotopic liver transplantation were previously attributed to poor surgical technique rather than the lack of the hepatic artery re-construction [18]. Later on, it appeared that it was spontaneous post-transplant graft re-arterialisation that compensated the absence of initial artery supply, which might be accounted for the reported high success rates and an almost normal histology. The development of spontaneous graft re-arterialisation through adhesions of the omentum and the surrounding organs is however a slow process. The earliest sign of collateral arterial blood flow recorded by microspheres as well as by angiography in non-arterialised liver grafts was not seen before the third post-transplant week [19]. Only until the 8<sup>th</sup> week, significant re-arterialisation could be evidenced by histology, while the regression of bile duct damage and the regression of bile duct proliferation were accomplished

### 88 3.2. Graft Re-arterialisation in Rat HALTx

6 months after the transplantation [20]. Using magnetic resonance imaging, we observed that in the rat HALTx, the healthy engrafted livers could already be distinguished from those that would develop post-operative complications (including biliary obstruction) and eventually atrophy as early as in the first post-transplant week (un-published data). This indicates that in the HALTx model the early graft function is crucial for long-term graft survival, since the possible recuperation of the engrafted liver is conditioned not only by the degree of initial hepatic damage but also by proper balance in the inter-liver competition. If a functional advantage is even temporarily given to the native liver, spontaneous graft re-arterialisation, which may occur at a later stage, will not be able to reverse the ongoing graft atrophy. As for the orthotopic transplantation [6, 9], the current results emphasize the importance of re-arterialisation for reducing biliary complications and consequent graft failure in the HALTx model.

The data of the present study also showed that, with or without hepatic artery supply, regeneration was transiently stimulated in the engrafted livers following a HALTx. This increase in DNA synthesis rate was presumably due to a “first pass” of portal hepatotrophic factors flowing through the partial hepatectomized graft. The 24-hour delay of the initial peak compared with that after a standard two-third partial hepatectomy [21] could be due to ischemic damage and operative stress [22]. Remarkably, the re-arterialised rats revealed a more pronounced regenerative response than the non re-arterialised rats despite equivalent portal blood supply. As the donor and recipient rats were syngeneic and the ischemic time was comparable between the two experimental groups in this study, neither acute rejection [23] nor ischemic damage [24] could explain these recorded differences. Thus, it was the re-arterialisation that directly enhanced the first regeneration boost in the engrafted livers. It is known that warm ischemia during the transplantation procedures causes depletion of ATP, which directly induces damage to the hepatocytes [25] and impairs the general hepatic functions such as protein synthesis and bile secretion [26]. The initial regeneration response is also depressed [22]. Re-arterialisation does improve the hepatic oxygen supply needed for an increased ATP synthesis in the liver graft [8], so that the significantly high DNA synthesis rate shown in the re-arterialised rats of this study may directly be attributed to reduced warm ischemic injury of the engrafted liver.

The elevated regenerative response however did not persist as seen in the rats subjected to a two third partial hepatectomy or as recorded in the orthotopic liver transplantation model [27] and fell to low levels immediately after the initial peak. In orthotopic liver transplantation, a loss of the functional hepatocytes caused by cellular necrosis does continue to stimulate

regeneration, whereas in HALTx, the remaining recipient's native liver may compensate in case of a deficient graft function. In addition, this model is remarkable by the fact that after the transplantation the total amount of hepatic mass exceeds the pre-transplant normal liver volume. This could explain the "switch-off" phenomenon of proceeding regenerative process in the engrafted liver. The present results suggest that it takes more than a "first pass" of portal stimulating factors to maintain regeneration. Whether the "switch-off" is due to an increase of specific inhibitory factors or a reduction of stimulating factors circulating through the graft remains to be clarified. The rat model of HALTx may serve as a useful tool to study this phenomenon regularly seen in clinical HALTx and oversized liver transplantation. This oversized liver model may thus provide new paths to gain further insight into the complicated mechanisms of liver regeneration.

Atrophy of the engrafted liver in the HALTx was first recognised as such by Starzl and his co-workers [1]. As this atrophy could be prevented by portal vascularization of the graft [2], the maintenance of portal supply is considered the prime factor for the preservation of graft integrity. In turn, the hormonal hepatotrophic factors are regarded as the key elements in the inter-liver competition of HALTx. This study shows that in this oversized HALTx model, without re-arterialisation, graft survival can not be guaranteed by a sole portal blood supply. The functional position of the engrafted liver in the balance of inter-liver competition is another important factor determining graft regeneration. In addition, the current study demonstrates that re-arterialisation of the engrafted liver can not only significantly enhance the general function of the engrafted liver such as improved bile production and reduce post-operative biliary complications, but also induce a strong post-transplant regenerative response. A better graft and animal survival could then be obtained as compared with non re-arterialised model. We may thus conclude that the re-arterialisation of the engrafted liver indeed plays an important role to prevent graft atrophy and should be considered as necessity in this rat model of HALTx.

## REFERENCES

1. Marchioro TL, Porter KA, Dickins TC, Faris TD, Starzl TE: Physiologic requirements for auxiliary liver transplantation. *Surg Gynecol Obstet* 1965; 121: 17-31.
2. Starzl TE, Halgrimson CG, Francavilla FR, Porter KA, Brown TH, Putnam CW: The origin, hormonal nature and action of hepatotrophic substances in portal venous blood. *Surg Gynecol Obstet* 1973; 137: 179-199.
3. Fan Y-D, Praet M, de Hemptinne B: The need to handicap the recipient's native liver in the rat model of heterotopic auxiliary liver transplantation. *HPB*

### 90 3.2. Graft Re-arterialisation in Rat HALTx

- Surgery 1999; 11:225-234.
4. Hess F, Jerusalem C, van der Heyde MN: Advantages of auxiliary liver homotransplantation in rats. Arch Surg 1972; 104: 76-80.
  5. Yu W, Wan X, Wright-JR J, Coddington D, Bitter SH: Heterotopic liver transplantation in rats: effect of intrahepatic islet isografts and split portal blood flow on liver integrity after auxiliary liver isotransplantation. Surgery 1994; 115: 108-117.
  6. Post S, Menger MD, Rentsch M, Gonzalez AP, Herfarth C, Messmer K: The impact of arterialization on hepatic microcirculation and leukocyte accumulation after liver transplantation in the rat. Transplantation 1992; 54: 789-794.
  7. Sumimoto R, Shinomiya T, Yamaguchi A: Influence of hepatic arterial blood flow in rats with liver transplants. Examination of donor liver-derived serum class I MHC antigen in rats with liver transplants with or without hepatic arterial reconstruction. Transplantation 1991; 51: 1138-1139.
  8. Reck T, Steinbauer F, Steinbauer M, et al: Impact of arterialization on hepatic oxygen supply, tissue energy phosphates, and outcome after liver transplantation in the rat. Transplantation 1996; 62: 582-587.
  9. Howden B, Jablonski P, Grossman H, Marshall VC: The importance of the hepatic artery in rat liver transplantation. Transplantation 1989; 47: 428-431.
  10. Zhao D, Zimmermann A, Wheatley AM: Morphometry of the liver after liver transplantation in the rat: significance of an intact arterial supply. Hepatology 1993; 17: 310-317.
  11. Gao W, Lemasters JJ, Thurman RG: Development of a new method for hepatic rearterialization in rat orthotopic liver transplantation. Reduction of liver injury and improvement of surgical outcome by arterialisation. Transplantation 1993; 56: 19-24.
  12. Fan Y-D, Praet M, de Hemptinne B: Inbred PVG rats are not suitable for the model of heterotopic auxiliary liver transplantation (HALTx). Euro Surg Res 1996; 28: 33.
  13. Sato Y, Farges O, Akpinar E, Yunming S, Yunming B, Bismuth H: An easy and physiologic arterial reconstruction method (sleeve technique) for orthotopic rat liver transplantation. Transplant Proc 1996; 28: 3649-3651.
  14. Post S, Palma P, Gonzalez AP, Rentsch M, Menger MD: Timing of arterialization in liver transplantation. Ann Surg 1994; 220: 691-698.
  15. Besse T, de Hemptinne B, Kabeya V, Lambotte L: Stimulation of liver regeneration by prostacyclin. Transplant Proc 1991; 23: 542-544.
  16. Bowers BA, Branum GD, Rotolo FS, Watters CR, Meyers WC: Bile flow--an index of ischemic injury. J Surg Res 1987; 42: 565-569.
  17. Karwinski W, Husoy AM, Farstad M, Soreide O: Sixty minutes of normothermic ischemia in the rat liver: correlation between adenine nucleotides and bile excretion. J Surg Res 1989; 46: 99-103.
  18. Kamada N, Calne RY: A surgical experience with five hundred thirty liver transplants in the rat. Surgery 1983; 93: 64-69.
  19. Svensson G, Naredi P, Hafstrom L, Tufveson G: Quantitative measurements of collateral arterial blood flow in nonarterialized rat liver grafts. Transpl Int 1994; 7: 136-139.

20. Zhao D, Zimmermann A, Kuznetsova LV, Wheatley AM: Regression of bile duct damage and bile duct proliferation in the non-rearterialized transplanted rat liver is associated with spontaneous graft rearterialization. *Hepatology* 1995; 21: 1353-1360.
21. de Hemptinne B, Leffert HL: Selective effects of portal blood diversion and glucagon on rat hepatocyte rates of S-phase entry and deoxyribonucleic acid synthesis. *Endocrinology* 1983; 112: 1224-1232.
22. Bolitho G, Engelbrecht G, Lotz Z, et al: Liver regeneration after hepatic ischemia and reduced liver autotransplantation in the rat. *Hepatology* 1993; 17: 273-279.
23. Teramoto K, Shimizu K, Tsukada K, Kamada N: DNA synthesis in hepatocytes during liver allograft rejection in rats. *Transplantation* 1990; 50: 199-201.
24. Foschi D, Castoldi L, Lesma A, Musazzi M, Benevento A, Trabucchi E: Effects of ischaemia and reperfusion on liver regeneration in rats. *Eur J Surg* 1993; 159: 393-398.
25. Morimoto T, Kusumoto K, Isselhard W: Impairment of grafts by short-term warm ischemia in rat liver transplantation. *Transplantation* 1991; 52: 424-431.
26. Gonzalez FX, Rimola A, Grande L, et al: Predictive factors of early postoperative graft function in human liver transplantation. *Hepatology* 1994; 20: 565-573.
27. Engelbrecht GH, Mcleod H, Tyler M, Lotz Z, Jaskiewicz K, Hickman R: Does liver transplantation in the rat cause a regenerative response. The effect of arterialisation of the graft. *HPB Surg* 1994; 7: 211-217.

### 3.3. Effects of Portal Vein Arterialisation on Liver Regeneration after Partial Hepatectomy in the Rat

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#### SUMMARY

Although portal venous supply is considered essential to preserve hepatic integrity, in this study, effects of portal arterialisation on liver regeneration were evaluated in a rat model of partial hepatectomy. Ninety-six Lewis rats were randomly assigned to four groups of twenty-four each: partial hepatectomy only (group 1), partial hepatectomy with either venous or arterialised portal supply (groups 2 and 3, respectively), and partial hepatectomy without portal supply (group 4). Liver regeneration rate, 5-bromo-2-deoxyuridine (BrdU) labelling index, and liver biological characteristics were assessed on days 1, 2, 3 and 7. Compared with group 1, all the tested rats had a marked body weight loss after surgery, and only the rats in group 4 showed no signs of recovery on day 7. With maintained portal inflow (groups 1, 2, and 3), liver regeneration rate increased steadily to day-7 values of  $89.2 \pm 11.8\%$ ,  $81.4 \pm 8\%$  and  $77.4 \pm 9.4\%$ , respectively ( $p = \text{not significant}$ ), and 24-hour peak values of BrdU labelling index were  $159 \pm 26$ ,  $157 \pm 42$ , and  $149 \pm 48$ , respectively ( $p = \text{not significant}$ ). Conversely, the rats deprived of portal supply (group 4) showed profound inhibition of these two parameters ( $14 \pm 13$ ,  $p < 0.01$ ;  $32.1 \pm 7.7\%$ ,  $p < 0.001$ , respectively). These results indicate that proper portal blood supply is essential to initiate and maintain liver regeneration after partial hepatectomy. With an equivalent portal inflow rate of either venous or arterial source, the hepatic regeneration response can be sustained.

## **INTRODUCTION**

The concept of arterialisation of the portal vein has drawn a constant attention since the 1960s because of its various clinical applications. In an attempt to reduce post-shunt encephalopathy, portal arterialisation was first used clinically in combination with an end-to-side portosystemic shunt to treat patients with liver cirrhosis.<sup>1-3</sup> Because the increased oxygen supply after portal arterialisation can improve hepatic energy metabolism and liver regeneration, a technique of partial arterialisation has also been favoured in extended hepatectomy for patients with advanced hepatobiliary malignancy.<sup>4</sup> Temporary use of portal arterialisation during liver transplantation surgery has shown beneficial effects on shortening warm ischemia time of the donor liver and consequently reducing the incidence of immediate post-operative graft failure.<sup>5, 6</sup> In the case of extensive portal and splanchnic vein thrombosis, or absence of the portal and mesenteric vein because of anatomic variations, portal arterialisation is an option to preserve sufficient hepatic blood supply in orthotopic liver transplantation.<sup>7-11</sup> More recently, encouraging results have been reported with the use of this procedure in heterotopic auxiliary liver transplantation to treat patients with acute liver failure.<sup>12</sup>

The quality of the portal venous inflow has generally been regarded as the key elements for the preservation of hepatic integrity.<sup>13-15</sup> With a relative low concentration of hormonal hepatotrophic factors, an arterialised portal supply would hamper the regeneration capacity of the liver. In the present study, liver growth and DNA synthesis rate were compared among rats with either venous or arterialised portal inflow, or without portal supply after partial hepatectomy. The post-operative evolution of body weight and liver biological characteristics was also analysed. A modified technique of arterial-venous anastomosis was used to obtain an arterialised portal supply with an inflow rate similar to that of portal venous supply following partial hepatectomy.

## **MATERIALS AND METHODS**

### **Experimental Animals**

Male inbred Lewis rats (IFFA CREDO Belgium sa, Brussels, Belgium), weighing 240-300 g, were used in this study. Animals received humane care according to European guidelines (Het Belgisch Staatsblad, Feb. 29<sup>th</sup>, 1992). All animals were kept in a temperature-controlled environment with a 12-hour light-dark cycle and had free access to food and water. After an

acclimation period of one week, all rats were fasted for 12 hours before surgery.

Ninety-six rats were randomly assigned into four groups of twenty-four rats each. A standard 68% partial hepatectomy was performed in the rats of all four experimental groups to induce liver regeneration. In group 1, only partial hepatectomy was performed. In group 2, with the exception of a period of portal vein occlusion to reproduce the transitory splanchnic congestion similar to group 3, portal venous supply of the liver remained intact. In group 3, portal supply of the liver was arterialised through an arterial-venous anastomosis. In group 4, portal supply of the liver was completely by-passed by a portacaval shunt.

### **Surgical Procedure**

Under ether anaesthesia, all the operations were performed using clean but non-sterile instruments. A heating pad was used to maintain a stable body temperature at approximately 36°C throughout the surgery.

A 68% partial hepatectomy (68% PH) was performed according to the technique described by Higgins and Anderson.<sup>16</sup> Through a midline incision, the median and the left lobes of the liver were excised after placement of a 5-0 suture ligature on the corresponding pedicle. Rats of all groups were also subjected to a right nephrectomy, which was needed for portal arterialisation in group 3. In the rats of group 2, after a 68% PH, the portal vein was cross-clamped for a period of ten minutes (equivalent to the average time needed to perform portal vein arterialisation in the rats of group 3). A portacaval shunt (PCS) was placed in the rats of group 3 and 4 using the technique introduced by Lee and Fisher:<sup>17</sup> after dividing the pyloric vein, the portal vein was ligated and transected. Using hand-sutured technique, the distal part of the portal vein was anastomosed to the vena cava in an end-to-side manner. Arterialisation of the portal vein (PVA) in the rats of group 3 was performed with the stent technique: after a right nephrectomy, a polyethylene tubing (24 Ga) of 0.6 cm was interposed between the right renal artery and the proximal part of the portal stump, and then secured with a 5-0 suture. This technique had the advantage of allowing arterial-venous anastomosis to be conducted with a stable inflow rate by using a stent with a standard inner diameter. It was measured in our laboratory that, through a stent of 24 Ga (0.5 mm inner diameter), the average flow rate of an arterialised portal supply was between  $8 \pm 3$  ml/min and  $9 \pm 3$  ml/min, which was similar to the recorded portal venous inflow after a 68% PH ( $7 \pm 2$  ml/min, minimum;  $8 \pm 2$  ml/min maximum). The peritoneal cavity was closed in two layers.



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At the end of operation, 1.5 cc of heparinized physiological saline solution was transfused intravenously and 10 mg Na Cefazolin was injected intramuscularly. The pre-operative diet of food pellets were resumed two hours after operation.

#### Assessment of Liver Regeneration

Six rats from each group were sacrificed on the post-operative day 1, 2, 3 and 7. One hour before sacrifice, all examined rats were weighted and 5-bromo-2-deoxyuridine (BrdU, 50 mg/Kg body weight) was injected intraperitoneally. Each liver was removed and immediately weighed. The ratio of liver weight to body weight (LW-BW) was calculated. From the resected liver weight at surgery and the remnant liver weight at autopsy, the liver regeneration rate (LRR) was determined according to the formula of Child:<sup>18</sup>

$$\frac{(\text{liver weight at autopsy} - \text{estimated residual liver weight at the time of surgery})}{\text{resected liver weight}} \times 100\%$$

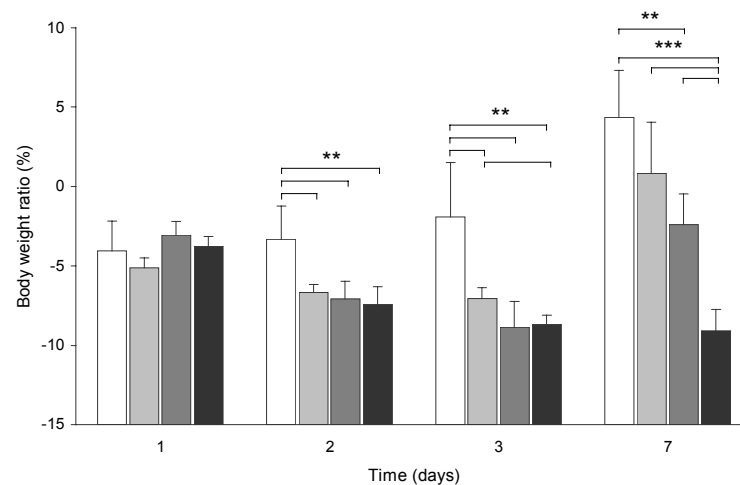
The resected liver specimens were fixed in 4% formalin. Five-micron thick sections were stained with haematoxylin and eosin for light microscopic examination. To allow calculation of BrdU labelling index, an indicator for DNA synthesis in hepatocytes, immunohistochemical staining of incorporated BrdU was obtained using Ventana system (Ventana Medical Systems, Inc. Tucson, Arizona, USA). After deparaffinization, the sections were sequentially treated with inhibitor for four minutes and protease 1 for six minutes. The sections were then incubated with monoclonal IgG1 anti-BrdU antibodies (Medical & biological laboratories co., Naka-ku Nagoya, Japan) for thirty-two minutes, after which they were incubated consequently with amplifier A, amplifier B, biotinylated Ig, avidin-hrpo, AEC and AEC H2O2 for eight minutes each. The sections were counterstained using haematoxylin for six minutes and bluing reagent for two minutes. All steps were carried out at the temperature of 37°C. Hepatocytes containing nuclei with brown staining were considered labelled. The number of positive stained cells was counted in randomly selected fields of ten each in periportal, midzonal and centrolobular areas using high-power fields (original magnification x 320). The number of BrdU-labelled nuclei per 1000 hepatocytes was designated as the BrdU labelling index.

#### Liver Biochemical Tests

At sacrifice, blood samples were drawn from the inferior vena cava. Serum alanine aminotransferase activity (ALAT) and albumin concentration (Alb) were analysed for subsequent assessment of hepatic injury and liver function, respectively.

### Statistics

Data were expressed as mean  $\pm$  standard deviation (SD). Data distribution was checked by the Kolmogorov test. The Student's *t*-test or the Mann-Whitney rank sum test was applied for normally or not-normally distributed data, respectively. *P* value of 0.05 or less was considered statistically significant.



**Figure 1.** Body weight ratio (BWR) 1, 2, 3 and 7 days after 68% partial hepatectomy. Compared with the control rats (group 1, □), on day 2 and 3, the rats either with portal inflow (venous, group 2, ■; arterialised, group 3, ■) or without portal inflow (group 4, ■) had marked body weight loss. On day 7, only the rats deprived of portal supply (group 4, ■) showed no signs of recovery. (*n* = 6 each group). Values were expressed as mean  $\pm$  SD. Asterisks indicated statistically significant differences between the experimental groups. \*\* *p* < 0.01; \*\*\* *p* < 0.001

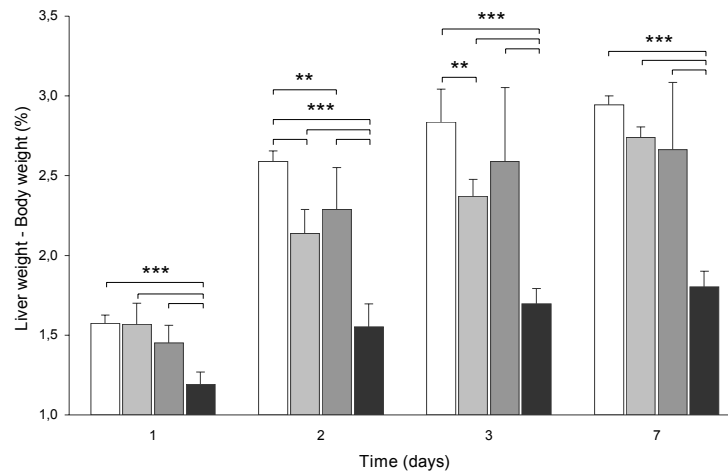
## RESULTS

### Body Weight

Post-operative changes in body weight were expressed as the ratio to the initial value immediately after surgery (day 0) (*BWR*). As shown in *Figure 1*, the mean *BWR* of the rats subjected to partial hepatectomy (group 1) increased steadily from  $-4.06 \pm 1.89\%$  on day 1 to  $4.35 \pm 2.97\%$  on day 7. The different surgical manipulations in the rats of group 2 and 3 showed similar effects on *BWR* evolution: *BWR* values of these two groups declined during the first three days after surgery. They were significantly lower than that of group 1 on the second ( $p < 0.01$ , group 2 and 3) and the third post-operative day ( $p < 0.01$ ;  $p = 0.001$ , respectively). On day 7, though the values of group 2 and 3 were still lower than that of group 1 ( $p < 0.01$ , group 3), they nearly recovered from the initial loss, and the difference between these two groups was not significant. Without portal blood supply, however, the mean *BWR* value of the rats in group 4 remained at a low level even seven days after surgery ( $-9.09 \pm 1.35\%$ ), which was significantly lower than the three other groups ( $4.35 \pm 2.97\%$ ,  $0.83 \pm 3.37\%$ ,  $-2.42 \pm 1.93\%$ , respectively;  $p < 0.001$ ).

### Ratio of Liver Weight to Body Weight (*LW-BW*)

After the operation, the mean *LW-BW* value of the rats in group 1 rose from  $1.57 \pm 0.1\%$  on day 1 to  $2.93 \pm 0.18\%$  on day 7, an increase of 213% from its initial value of  $0.94 \pm 0.08\%$  (day 0). A sharp increase of *LW-BW* was also observed in the rats of both group 2 and 3: from the day-1 value of  $1.57 \pm 0.13\%$  (group 2) and  $1.50 \pm 0.13\%$  (group 3), they reached  $2.73 \pm 0.19\%$  and  $2.69 \pm 0.20\%$  on day 7, which corresponded to a 180% and 177% increase from the initial values ( $0.98 \pm 0.06\%$ ,  $0.97 \pm 0.04\%$ , respectively). Compared with those of group 1, the values of group 2 and 3 were significantly lower on day 2 ( $p < 0.001$ , group 2;  $p < 0.01$ , group 3) and day 3 ( $p = 0.001$  group 2), whereas the differences were no longer significant on day 7. No marked differences could be recorded between the values of group 2 and 3. In the absence of portal supply, the rats of group 4 showed the lowest increase rate of *LW-BW* among the four groups. Throughout the one-week observation period, the *LW-BW* values of group 4 were markedly lower than those of the other three groups ( $p < 0.001$ ). On day 7, the value of  $1.80 \pm 0.2\%$  was only an increase of 94% from its initial value of  $0.93 \pm 0.03\%$  (*Figure 2*).

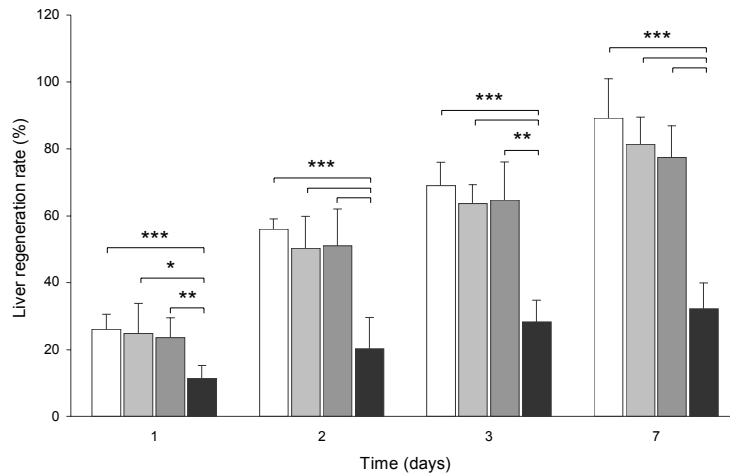


**Figure 2.** Ratio of liver weight to body weight (*LW-BW*) 1, 2, 3 and 7 days after 68% partial hepatectomy. Compared with the control rats (group 1, □), the increase of *LW-BW* was slower in the rats with either venous portal inflow (group 2, ■; day 2 and 3) or arterialised portal inflow (group 3, ■; day 2). Throughout the one-week observation period, *LW-BW* in the rats deprived of portal supply (group 4, ■) was significantly lower than those in the rats of other groups. ( $n = 6$  each group). Values were expressed as mean  $\pm$  SD. Asterisk indicated statistically significant differences between the experimental groups. \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ .

### Liver Regeneration Rate (*LRR*)

As shown in *Figure 3*, hepatectomised livers regenerated at a similar rate in the rats of group 1, 2 and 3 during the first post-operative week. From the day-1 values of  $26.3 \pm 4.8\%$  (group 1),  $24.8 \pm 8.9\%$  (group 2) and  $23.7 \pm 5.8\%$  (group 3), *LRR* values increased to  $89.2 \pm 11.8\%$ ,  $81.4 \pm 8\%$  and  $77.4 \pm 9.4\%$  on day 7, respectively. No significant differences were found among the values of these three groups. Complete portal by-pass in the rats of group 4 hampered liver regeneration profoundly: compared with those of group 1, 2 and 3, the respective *LRR* values of group 4 were significantly lower on day 1 ( $p \leq 0.01$ ), day 2 ( $p < 0.001$ ) and day 3 ( $p \leq 0.001$ ). On day 7, the value of  $32.1 \pm 7.7\%$  in the rats of group 4 was 64%, 61% and 59% less than those of group 1, 2 and 3, respectively ( $p < 0.001$ ).

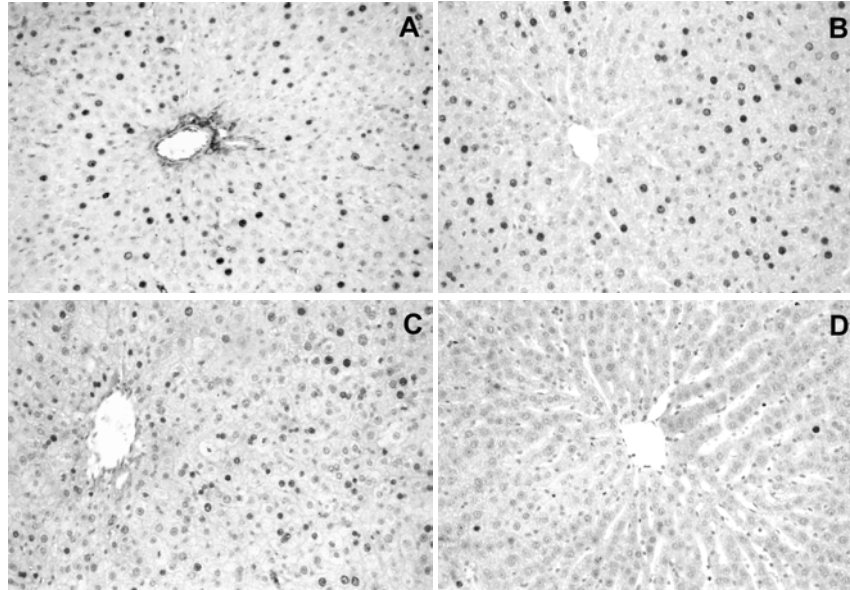
### 98 3.3. Portal Arterialisation on Liver Regeneration



**Figure 3.** Liver regeneration rate (LRR) 1, 2, 3 and 7 days after 68% partial hepatectomy. Similar to the control rats (group 1, □), the rats with persistent portal inflow (venous, group 2, ■; arterialised, group 3, ■) had significantly higher liver growth than the rats deprived of portal supply (group 4, ■) during the first post-operative week. ( $n = 6$  each group). Values were expressed as mean  $\pm$  SD. Asterisk indicated statistically significant differences between the experimental groups. \*  $p \leq 0.05$ ; \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ .

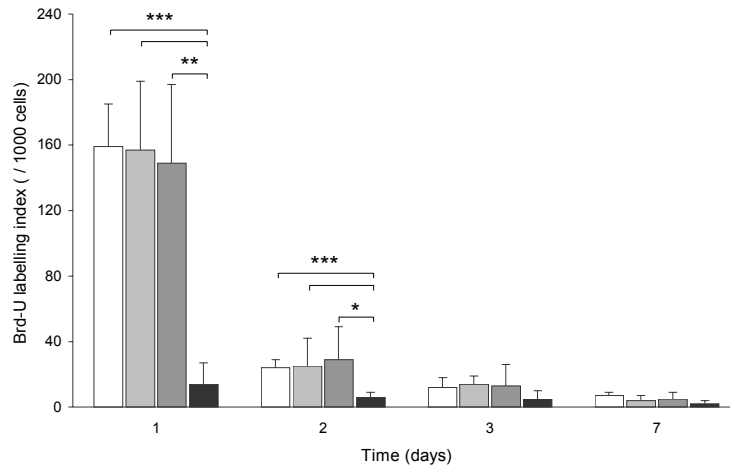
#### BrdU Labelling Index

Immunohistochemical staining for BrdU in livers of the four experimental groups is shown in *Figure 4*. The post-operative evolution of BrdU labelling index was similar in the rats of group 1, 2 and 3: the number of positively stained cells peaked twenty-four hours after PH ( $159 \pm 26$ ,  $157 \pm 42$  and  $149 \pm 48$ , respectively) and declined thereafter. Differences among these three groups were not significant. In contrast, the BrdU index was drastically suppressed in the rats without portal blood supply. The recorded 24-hour value of  $14 \pm 13$  in the rats of group 4 was less than one tenth of those in the rats of the other three groups ( $p < 0.001$ , compared with group 1 and 2;  $p < 0.01$ , with group 3). On day 2, it was still significantly lower than those of other groups ( $p < 0.001$ , compared with group 1 and 2;  $p < 0.05$ , with group 3) (*Figure 5*).



**Figure 4.** Immunohistochemical staining for 5-bromo-2-deoxyuridine (BrdU) 24 hours after 68% partial hepatectomy. Similar to the control rats (group 1, **A**), the rats with persistent portal inflow (venous, group 2, **B**; arterialised, group 3, **C**) had a significantly higher percentage of BrdU-positive staining hepatocytes (nuclei with deposition of brownish pigment) than the rats deprived of portal supply (group 4, **D**). (original magnification x 400).

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**Figure 5.** 5-bromo-2-deoxyuridine (BrdU) labelling index on 1, 2, 3 and 7 days after 68% partial hepatectomy. On day 1 and 2, similar to the control rats (group 1, □), the rats with persistent portal inflow (venous, group 2, ■; arterialised, group 3, ■) had a significantly higher percentage of BrdU-positive staining hepatocytes than the rats deprived of portal supply (group 4, ■). (n = 6 each group). Values were expressed as mean ± SD. Asterisk indicated statistically significant differences between the experimental groups. \*  $p \leq 0.05$ ; \*\*  $p < 0.01$ ; \*\*\*  $p < 0.001$ .

#### Liver Biochemical Tests

The mean values of serum albumin concentration decreased in the rats of all four groups after surgery (*Table 1*). Serum albumin concentration (Alb) of the rats in groups 1, 2 and 3 recovered one week later and no marked differences were found among the values of these three groups. Without portal supply, the rats of group 4 showed a prolonged decrease in serum Alb levels, and the value of  $2.7 \pm 0.3$  on day 7 was significantly lower than the level of  $3.2 \pm 0.3$  of group 1 ( $p = 0.01$ ). The mean levels of serum alanine aminotransferase activity (ALAT) peaked one day after surgery in all rats of the four groups and declined thereafter. Compared with the value of group 1, significantly higher ALAT levels were found only in group 3 on day 2 ( $p < 0.05$ ) and in group 4 on day 1 ( $p < 0.01$ ).

**Table 1.** Post-operative evolution of serum albumin concentration (Alb) and alanine aminotransferase activity (ALAT)

	Day -1	Day 1	Day 2	Day 3	Day 7
Alb (g/dl)	3.3 ± 0.2				
group 1 (n = 6)		2.9 ± 0.2	3.0 ± 0.2	3.0 ± 0.5	3.2 ± 0.3
group 2 (n = 6)		2.7 ± 0.2	2.8 ± 0.3	3.0 ± 0.3	3.0 ± 0.3
group 3 (n = 6)		2.7 ± 0.1	2.8 ± 0.2	2.7 ± 0.2	2.9 ± 0.3
group 4 (n = 6)		2.9 ± 0.2	2.8 ± 0.2	2.7 ± 0.3	2.7 ± 0.3*
ALAT (U/l)	41 ± 9				
group 1 (n = 6)		292 ± 62	109 ± 37	68 ± 17	56 ± 7
group 2 (n = 6)		360 ± 188	130 ± 87	65 ± 11	49 ± 9
group 3 (n = 6)		357 ± 59	156 ± 47*	62 ± 19	54 ± 11
group 4 (n = 6)		454 ± 70**	131 ± 24	54 ± 7	50 ± 26

Note: Values were expressed as mean ± SD. Asterisk indicated statistically significant differences versus the values of group 1. \*  $p \leq 0.05$ ; \*\*  $p < 0.01$ .

## DISCUSSION

The importance of portal blood supply to regeneration of the liver has been noted since the 1920s, based upon the observation that total liver mass and hepatocyte number are larger with intact portal inflow and atrophy occurs in liver deprived of portal venous blood.<sup>19</sup> It is further evidenced as such by the discovery that a liver lobe transplanted in an auxiliary position will regenerate, if the engrafted liver instead of the native liver receives the portal blood.<sup>20, 21</sup> The data from the current study clearly show that adequate portal inflow is essential for liver regeneration after a partial hepatectomy. Corresponding well with previous reports,<sup>22, 23</sup> complete diversion of portal blood realised by a portacaval shunt not only results in prolonged post-operative body weight loss, but also delays or abolishes the expected sharp increase in DNA synthesis induced by a standard 68% partial hepatectomy and impairs the restoration of the liver mass. Absence of portal flow induces hepatocyte degeneration characterized by cell volume decrease and cell death,<sup>24</sup> and deprives of the portal hepatotrophic factors.<sup>25, 26</sup> These may be responsible for the hampered regeneration capacity. On the other hand, a significant decrease in total portal inflow is known to lead to a rapid increase



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in the blood flow in the hepatic artery, through which general liver function can be sustained.<sup>11, 27</sup> However, the data from this study reveal that, deprived of portal supply, the peak value of DNA synthesis rate is reduced to one tenth of the expected normal value, and liver regeneration rate one week after partial hepatectomy is only one third of that recorded in the rats with normal portal supply. This finding may indicate that the degree of the compensation from the hepatic artery is insufficient to initiate a proper hepatic regeneration response and maintain the regenerative process after partial hepatectomy, which would confirm the theory that this compensatory mechanism of the so-called hepatic artery buffer response is only partial.<sup>28, 29</sup>

To date, much speculation exists, in particular, on whether the quality of the portal blood (contents) or the quantity of the portal blood (inflow rate) is of major importance for liver regeneration. It is suggested that the diversion of hormonal hepatotrophic factors, such as endogenous insulin, from the portal tract to the systemic circulation, is responsible for the atrophy seen after portal shunt.<sup>14, 30</sup> Conversely, other investigators have shown that when the portal tract is perfused by systemic venous blood rather than the blood flowing from the intestines and pancreas, liver size can be maintained<sup>18, 31</sup> and regenerative hyperplasia is within the normal range.<sup>32</sup> The evidence in this study that hepatic regeneration response can be initiated, even with an arterialised portal supply after a partial hepatectomy, further supports the later argument. Moreover, when the inflow rates are equivalent, the hepatectomised livers with an arterialised portal supply can regain their weight as fast as those with a portal venous supply. By separately measuring hepatic venous and arterial inflow (per gram of tissue mass) after a left portal vein stenosis, Rocheleau et al. recently demonstrated that either portal venous or hepatic arterial flow could maintain viable liver mass in proportion to the amount of blood flow delivered.<sup>33</sup> The data from the present study may therefore indicate that the initiation and maintenance of liver regeneration induced by a partial hepatectomy do not necessarily require highly concentrated hepatotrophic factors, or that elements other than those classical splanchnic factors may be the initial trigger of the regenerative process.

Based on the discoveries of the last decades, the phenomenon of liver regeneration has been defined as a precise and highly orchestrated response induced by specific external stimuli, which involves sequential changes in morphologic structure, gene expression and growth factor production. Although many growth factors and cytokines, such as hepatocyte growth factor, epidermal growth factor, transforming growth factor- $\alpha$ , interleukin-6, tumour necrosis factor- $\alpha$  insulin, and norepinephrine, appear to play an important role in this process, none of these substances seem to be a primary

mitogen for hepatocytes.<sup>34</sup> It has been realised that before replication, hepatocytes have to be sensitised in order to be competent and fully responsive to the trophic action of growth factors.<sup>35, 36</sup> It is hypothesized that this so-called “priming” phase, meaning the transition of hepatocyte from a  $G_0$  resting state to a  $G_1$  replicative state, needs gene activation, and that more than one pathway may be involved, which may require growth factors and cytokines such as tumour necrosis factor and interleukin 6.<sup>37</sup> As the priming begins simultaneously in all hepatocytes almost immediately after partial hepatectomy,<sup>38</sup> one can assume that it may be triggered either by hormonal factors already present in the blood or liver, or by hemodynamic factors. In parallel to an early and persistent increase in portal pressure,<sup>39, 40</sup> researchers have found that changes in hepatocyte membrane potential can be recorded as early as within five minutes following partial hepatectomy.<sup>41</sup> This has been linked to a series of alterations that include increases in sodium-potassium adenosine triphosphatase pump activity, sodium fluxes into hepatocytes, and sodium-dependent amino acid transport.<sup>38, 42-44</sup> A recent report showed that the blood-flow-activated sodium and potassium current in cardiac microvascular endothelial cells is flow-rate dependent.<sup>45</sup> In addition, an increase in portal pressure is also related to such ultra-structural changes in the liver as sinusoid dilatation and disappearance of the sieve-plate arrangement of small endothelial pores, thus leaving the surface of the parenchymal liver cell directly exposed to any circulating hepatotrophic substances present in the portal blood.<sup>40</sup> We speculate that a significant increase in relative blood flow and consequently an even slight increase in portal pressure induced by partial hepatectomy may itself be one of the first factors triggering the entire process of liver regeneration. Further clarification of the exact role of changing portal inflow rate and pressure in the priming phase may sort out the earliest signals associated with the initiation of the hepatic regeneration response.

The data from the present study demonstrate that proper blood perfusion in the portal tract is essential for initiating and maintaining liver regeneration after a partial hepatectomy in the rat. In case of an equivalent inflow rate, the immediate hepatic regeneration response can be initiated and the one-week liver regeneration rate can be sustained by an arterialed portal supply at the same level as by a portal venous inflow.

## REFERENCES

1. Hulten O. Arterialization of the liver in cirrhosis. *Scand J Clin Lab Invest Suppl* 1966; 18:43-3.

### 104 3.3. Portal Arterialisation on Liver Regeneration

2. Maillard JN, Benhamou JP, Rueff B. Arterialization of the liver with portacaval shunt in the treatment of portal hypertension due to intrahepatic block. *Surgery* 1970; 67(6):883-890.
3. Otte JB, Reynaert M, de Hemptinne B, Geubel A, Carlier M, Jamart J, Lambotte L et al. Arterialization of the portal vein in conjunction with a therapeutic portacaval shunt. Hemodynamic investigations and results in 75 patients. *Ann Surg* 1982; 196(6):656-663.
4. Shimizu Y, Miyazaki M, Shimizu H, Ito H, Nakagawa K, Ambiru S, Yoshidome H et al. Beneficial effects of arterialization of the portal vein on extended hepatectomy. *Br J Surg* 2000; 87(6):784-789.
5. Sheil AG, Thompson JF, Stephen MS, Graham JC, Evers AA, Bookallil M, Kalpokas M et al. Liver graft revascularization by donor portal vein arterialization following "no touch" donor hepatectomy. *HPB Surg* 1988; 1(1):57-66.
6. Morimoto T, Terasaki M, Higashiyama H, Tanaka K, Uemoto S, Tanaka A, Shimahara Y et al. Clinical application of arterialization of portal vein in living related donor partial liver transplantation. *Transpl Int* 1992; 5(3):151-154.
7. Pichlmayr R, Gubernatis G, Grosse H, Seitz W, Mauz S, Ennker I, Mei M et al. Liver transplantation in low portal vein flow: separation of portal vein areas with divided portal-venous and arterialized caval-venous liver perfusion. 1. Clinical case report. *Langenbecks Arch Chir* 1989; 374(4):232-239.
8. Neuhaus P, Bechstein WO, Blumhardt G, Steffen R. Management of portal venous thrombosis in hepatic transplant recipients. *Surg Gynecol Obstet* 1990; 171(3):251-252.
9. Stieber AC, Zetti G, Todo S, Tzakis AG, Fung JJ, Marino I, Casavilla A et al. The spectrum of portal vein thrombosis in liver transplantation. *Ann Surg* 1991; 213(3):199-206.
10. Erhard J, Lange R, Giebler R, Rauen U, de Groot H, Eigler FW. Arterialization of the portal vein in orthotopic and auxiliary liver transplantation. A report of three cases. *Transplantation* 1995; 60(8):877-879.
11. Troisi R, Kerremans I, Mortier E, Defreyne L, Hesse UJ, de Hemptinne B. Arterialization of the portal vein in pediatric liver transplantation. A report of two cases. *Transpl Int* 1998; 11(2):147-151.
12. Erhard J, Lange R, Rauen U, Scherer R, Friedrich J, Pietsch M, de Groot H et al. Auxiliary liver transplantation with arterialization of the portal vein for acute hepatic failure. *Transpl Int* 1998; 11(4):266-271.
13. Fisher B, Fisher ER, Safffer E. Investigations concerning the role of a humoral factor in liver regeneration. *Cancer Res* 1963; 23:914-920.
14. Starzl TE, Francavilla A, Halgrimson CG, Francavilla FR, Porter KA, Brown TH, Putnam CW. The origin, hormonal nature, and action of hepatotrophic substances in portal venous blood. *Surg Gynecol Obstet* 1973; 137(2):179-199.
15. Cohen DM, Jaspan JB, Polonsky KS, Lever EG, Moossa AR. Pancreatic hormone profiles and metabolism posthepatectomy in the dog. Evidence for a hepatotrophic role of insulin, glucagon, and pancreatic polypeptide. *Gastroenterology* 1984; 87(3):679-687.

16. Higgins GM, Anderson RM. Experimental pathology of the liver: I. Restoration of the liver of the white rat following partial surgical removal. *Arch Pathol* 1931; 12:186-202.
17. Lee S, Fisher B. Portacaval shunt in the rat. *Surgery* 1961; 50:668-672.
18. Child CG, Barr D, Halswarde GK. Liver regeneration following portal-caval transposition in dogs. *Ann Surg* 1953; 138:600-608.
19. Rous P, Larimore LD. Relation of the portal blood to liver maintenance: a demonstration of liver atrophy conditional on compensation. *J Exp Med* 1920; 31:609-632.
20. Sigel B, Acevedo FJ, Dunn MR. The effect of partial hepatectomy on autotransplanted liver tissue. *Surg Gynecol Obstet* 1963; 117:29-36.
21. Marchioro TL, Porter KA, Dickins TC, Faris TD, Starzl TE. Physiologic requirements for auxiliary liver transplantation. *Surg Gynecol Obstet* 1965; 121:17-31.
22. Whittemore AD, Kasuya M, Voorhees ABJ, Price JBJ. Hepatic regeneration in the absence of portal viscera. *Surgery* 1975; 77(3):419-426.
23. Rokicki M, Rokicki W. Liver regeneration in rats after complete and partial occlusion of the portal blood influx. *Res Exp Med (Berl)* 1993; 193(5):305-313.
24. Bilodeau M, Aubry MC, Houle R, Burnes PN, Ethier C. Evaluation of hepatocyte injury following partial ligation of the left portal vein. *J Hepatol* 1999; 30(1):29-37.
25. Fisher B, Szuch P, Levine M, Fisher ER. A portal blood factor as the humoral agent in liver regeneration. *Science* 1971; 171(971):575-577.
26. Francavilla A, Starzl TE, Barone M, Zeng QH, Porter KA, Zeevi A, Markus PM et al. Studies on mechanisms of augmentation of liver regeneration by cyclosporine and FK 506. *Hepatology* 1991; 14(1):140-143.
27. Lautt WW, Greenway CV. Conceptual review of the hepatic vascular bed. *Hepatology* 1987; 7(5):952-963.
28. Greenway CV, Oshiro G. Comparison of the effects of hepatic nerve stimulation on arterial flow, distribution of arterial and portal flows and blood content in the livers of anaesthetized cats and dogs. *J Physiol (Lond)* 1972; 227(2):487-501.
29. Lautt WW. The 1995 Ciba-Geigy Award Lecture. Intrinsic regulation of hepatic blood flow. *Can J Physiol Pharmacol* 1996; 74(3):223-233.
30. Rozga J, Jeppsson B, Bengmark S. Hepatotrophic factors in liver growth and atrophy. *Br J Exp Pathol* 1985; 66(6):669-678.
31. de Jonge J, Madern GC, Terpstra OT, Sinaasappel M, Molenaar JC, Provoost AP, Tilanus HW. Directing portal flow is essential for graft survival in auxiliary partial heterotopic liver transplantation in the dog. *J Pediatr Surg* 1999; 34(8):1265-1268.
32. Lee S, Broelsch CE, Flamant YM, Chandler JG, Charters AC, Orloff MJ. Liver regeneration after portacaval transportation in rats. *Surgery* 1975; 77(1):144-149.

### 106 3.3. Portal Arterialisation on Liver Regeneration

33. Rocheleau B, Ethier C, Houle R, Huet PM, Bilodeau M. Hepatic artery buffer response following left portal vein ligation: its role in liver tissue homeostasis. *Am J Physiol* 1999; 277(5 Pt 1):G1000-G1007.
34. Michalopoulos GK, DeFrances MC. Liver regeneration. *Science* 1997; 276(5309):60-66.
35. Moolten FL, Oakman NJ, Bucher NL. Accelerated response of hepatic DNA synthesis to partial hepatectomy in rats pretreated with growth hormone or surgical stress. *Cancer Res* 1970; 30(9):2353-2357.
36. Bucher NL. Liver regeneration then and now. In: Jirtle RL, editor. *Liver Regeneration and Carcinogenesis: Molecular and Cellular Mechanisms*. San Diego: Academic Press, 1995: 1-25.
37. Fausto N. Liver regeneration. *J Hepatol* 2000; 32(1 Suppl):19-31.
38. Kren BT, Trembley JH, Fan G, Steer CJ. Molecular regulation of liver regeneration. *Ann N Y Acad Sci* 1997; 831:361-381.
39. Fogli L, Gorini P, Cappellari L, Morsiani E. Effect of partial hepatectomy and liver regeneration on portal pressure in rats. *Surgical Research Communications* 1990; 6:159-166.
40. Morsiani E, Aleotti A, Ricci D. Haemodynamic and ultrastructural observations on the rat liver after two-thirds partial hepatectomy. *J Anat* 1998; 192(Pt 4):507-515.
41. Zhang XK, Gauthier T, Buczynski FJ, Wang GQ, Gong YW, Minuk GY. Changes in liver membrane potentials after partial hepatectomy in rats. *Hepatology* 1996; 23(3):549-551.
42. Wondergem R, Potter VR. A new protocol for studying the early events during liver regeneration. *Life Sci* 1978; 23(15):1565-1575.
43. Schenk DB, Hubert JJ, Leffert HL. Use of a monoclonal antibody to quantify (Na<sup>+</sup>,K<sup>+</sup>)-ATPase activity and sites in normal and regenerating rat liver. *J Biol Chem* 1984; 259(23):14941-14951.
44. Moule SK, McGivan JD. Regulation of the plasma membrane potential in hepatocytes--mechanism and physiological significance. *Biochim Biophys Acta* 1990; 1031(3):383-397.
45. Moccia F, Villa A, Tanzi F. Flow-activated Na<sup>(+)</sup> and K<sup>(+)</sup> Current in cardiac microvascular endothelial cells. *J Mol Cell Cardiol* 2000 Aug ;32 (8 ):1589 - 93 2000; 32(8):1589-1593.

## **CHAPTER 4**

### **GENERAL DISCUSSION**

### **Evaluation of graft viability following heterotopic auxiliary liver transplantation**

As assessed in the present work, various surgical manipulations on the recipient's native liver in the rat model of heterotopic auxiliary liver transplantation (HALTx) do affect post-transplant animal survival, graft survival and liver biology differently.

Although a normal rat can tolerate a 68% partial hepatectomy (68% PH) without major complications, this manipulation becomes a significant functional handicap to the liver whose portal supply is completely deprived by a portacaval shunt (PCS).<sup>1, 2</sup> The partial hepatectomy greatly increases the metabolic load at the hepatocellular level,<sup>3</sup> while the portal influx of nutritive elements needed to response to the high energetic metabolism is totally bypassed by a PCS.<sup>4, 5</sup> This results in an energetic deficit,<sup>6</sup> and, consequently, hepatic metabolic activities and active biosynthesis of cellular components are drastically hampered.<sup>2</sup> However, as shown in the current work, with the support of an auxiliary liver, the portal deprived native liver can recover from this surgical handicap and even take over the engrafted liver to regenerate. This indicates that functional reduction of the portal deprived liver induced by a 68% PH is still reversible.

The combined handicap of a 68% PH and a common bile duct ligation (CBDL) on a portal deprived liver is so severe not only that hepatic regeneration capacity is completely impaired leading to native liver atrophy in all long-term HALTx survivors, but also that the rats can not survive this handicap without the presence of a functioning auxiliary liver. In this case, the post-transplant animal survival relies solely upon the viability of the engrafted liver, and the values of serum liver biochemistry therefore reflect directly the functionality of the engrafted organ.<sup>7</sup> It is recognized that, in cholestasis, the intracellular retention of biliary constituents and the diminution of bile acids excretion inevitably leads to the imbalance between hydrophobic and hydrophilic bile salts in the biliary system.<sup>8-10</sup> The known hepatotoxic potential of the hydrophobic bile salts, which play a central role in the pathogenesis of bile-salt-induced liver injury, is then unmasked.<sup>8-12</sup> Along with high biliary pressure,<sup>13, 14</sup> hydrophobic bile salts cause hepatocyte death mainly through cytolysis.<sup>8, 10</sup> This is characterised by morphologic alterations including membrane disruption and cytoplasmic enlargement accompanied with increased hepatocellular enzyme release.<sup>15-17</sup> Apoptosis, a form of programmed cell death, has recently been found to be the predominant mechanism of hepatocyte death under moderate bile acid toxicity.<sup>17-19</sup> Hydrophobic bile salts also interfere with the mitochondrial function thereby decreasing ATP production,<sup>20-22</sup> and inhibit directly hepatocyte proliferation.<sup>23-25</sup> Furthermore, the activation of Kupffer cells

following extra- or intra-hepatic bile duct obstruction is linked to increased generation of reactive oxygen species and free-radical formation, which may indirectly injure the hepatocytes.<sup>26-32</sup> The subsequent activation of hepatic stellate cells by Kupffer cells induces the net deposition of collagen and other components of the extra-cellular matrix perpetuating liver necrosis, fibrosis and cirrhosis.<sup>29, 33-37</sup>

Although the damaging effects of this combined handicap of 68% PH and CBDL on the native liver is apparently fatal and irreversible, the data of post-transplant monitoring by magnetic resonance imaging (MRI) from the current work show that, in the presence of a viable auxiliary liver, the expected drastic functional decline of the native liver (reflected by  $T_1$  relaxation times) becomes obvious only four weeks after transplantation. This new evidence demonstrates that beyond its life-supporting role, a well-functioning auxiliary liver also reduces the mechanical and the biochemical aggression of the handicap imposed on the native liver. This slows down the process of the physiological deterioration of the handicapped organ. Liver biology, especially during the initial phase after a HALTx, is then a global representation of the viability of both the engrafted and the native livers. Extra caution should thus be taken when this parameter is used to assess graft viability in the rat model of HALTx. Obviously, this specific approach cannot be applied in the experimental studies where the survival of the native liver is expected, as the imposed inflicted handicap will completely abolish its regenerative potential.

The possible advantages of using MRI to monitor post-HALTx hepatic viability are for the first time evaluated and clearly demonstrated in the current work. It allows a separate identification of post-transplant complications such as abscess and necrosis in the two individual livers. The analysis of the relation between signal intensity and hepatic viability brings further interesting information. Corresponding well with a previous clinical report,<sup>38</sup> it is found that the higher the signal intensity on a  $T_1$  weighted image, the better the hepatic viability. The viability of each individual liver can thus be assessed by its signal intensity at any given time. In the current work, the post-transplant evolution of  $T_1$  relaxation times is compared between the healthy engrafted livers and the livers presenting infectious and ischemic complications. It reveals that the eventual outcome of the engrafted and the native liver, either regeneration or atrophy, can already be predicted at an early post-operative stage by examining the pattern of their  $T_1$  evolution. A non-invasive measurement as such can facilitate systematic post-transplant follow-up of the evolution of the viability of both livers. MRI is hence a very promising tool for the studies on inter-liver functional competition. In the situations as pre-surgical evaluation of transplant candidates in fulminant or sub-fulminant hepatic failure, and post-transplant



or post-hepatectomy assessment of liver viability, the clinical potential of MRI could be proved beneficial and indeed warrants further prospective human studies.

### **Study of inter-liver functional competition following heterotopic auxiliary liver transplantation**

After a clinical auxiliary liver transplantation, portal blood supply is shared between the engrafted liver and the recipient's native liver. It has generally been accepted that most of the common post-transplant events, such as reperfusion injury, allograft rejection, allograft viral infection and native liver regeneration, cause changes in intrahepatic portal venous resistance modifying the amount of portal blood delivered to the liver. The liver with less portal resistance receives more portal blood and consequently hepatotrophic factors, and therefore holds a more favourable position in the inter-liver competition. In this way, an alteration of the relative portal blood distribution will eventually lead to regeneration imbalance between the two livers.<sup>39-42</sup>

Notably, in the rat model of HALTx used in the current work, the recipient's portal supply is completely shunted towards the engrafted liver, while the native liver viability is only sustained by hepatic artery supply. Despite the fact that there is no sharing of portal blood between the two livers, a change in the degree of surgical handicap on the native liver can still alter the outcome of the engrafted organ from regeneration to atrophy; vice versa, an improvement of the physiological status of the engrafted liver, as obtained by re-establishing its hepatic arterial supply, does result in more pronounced native liver atrophy. Unlike in orthotopic liver transplantation (OLTx) where the progress of hepatic regeneration relays solely upon the number of viable hepatocytes in the newly implanted liver graft, after an auxiliary liver transplantation, the regeneration starts immediately in both coexisting livers to fulfil the body demand for hepatic metabolic function. If the liver is already in a more favourable physiological condition and in addition receives optimal portal supply, its regeneration will be advantaged preferably by "the first wave" of stimulating growth factors influx delivered via the portal tract.<sup>41, 43</sup> In parallel to the progressing regeneration of the one liver, decreased concentration of those stimulating factors in the blood circulation will further bring the other liver into a "dormant" state with eventual atrophy. On the other hand, a proper regenerative response cannot occur in a liver when its general metabolic function is profoundly hampered by either surgical trauma or reperfusion injury, despite proper portal blood supply. In this case, regeneration of the other liver, even deprived of portal supply, can still be stimulated by the "second wave" of growth factors re-

circulating through the hepatic artery.<sup>44</sup>

In contrary to what is seen in human liver transplantation, re-construction of the graft hepatic artery in the rat appears not to be needed for graft and animal survival.<sup>45, 46</sup> Microscopic studies of the non re-arterialised grafts will however reveal a high incidence of bile duct proliferation and biliary complications.<sup>46-48</sup> Regression of bile duct proliferation and bile duct damage is often observed in long-term survival rats, which eventually leads to almost normal liver histology.<sup>49</sup> The mechanism of this compensation to the lack of initial artery supply is mainly attributed to spontaneous graft re-arterialisation,<sup>46, 48-50</sup> which can be seen after the third post-transplant week.<sup>49, 50</sup> In HALTx, the possible recuperation of the engrafted liver is conditioned not only by the degree of initial hepatic damage but also by the balance of the inter-liver competition. Good early graft function is hence crucial for long-term graft survival in this model. Though the absence of arterial supply in the engrafted liver gives, in principle, only temporarily functional advantage to the native liver, spontaneous progressive graft re-arterialisation, which may occur at a later stage, will not be able to reverse the ongoing graft atrophy. In addition to the beneficial effects of graft re-arterialisation reported in previous studies,<sup>51-56</sup> the data of the current work further reveal that re-arterialisation improves early hepatic function, induces a more marked early regenerative response and reduces post-transplant biliary complications. The engrafted liver is thus functionally advantaged in the inter-liver competition with better graft survival compared with the non re-arterialised model.

Besides stressing the importance of the accessibility of the portal supply with its stimulating growth factors and nutritive elements, these results emphasize that the coexisting two livers after a HALTx are also competing for the regenerative capability based on their physiological status. The balance between the supply of regeneration stimulating factors and the hepatic capability to response to those factors will determine the final fate of the two livers. Up to date, little is known about whether any specific signals are transmitted between the two livers to regulate the post-HALTx process. If so, further exploration of the nature of those signals will bring more detailed insights about the mechanisms of the inter-liver competition phenomenon and may also have important clinical implications, in case of acute liver failure where temporary functional support of the auxiliary graft is required,<sup>57</sup> or in case of metabolic diseases where coexistence of two well-functioning livers is needed.<sup>58</sup>

**P**ortal blood supply has been proved to play a vital role in liver regeneration. As presented in this work, complete lack of portal supply will lead to a significantly impaired regenerative response and restoration of liver

mass. When the liver is deprived of its portal vascular blood, hormonal growth factors may only reach the liver through the arterial circulation.<sup>44</sup> Through vasodilation, hepatic arterial inflow increases instantly after a PCS to compensate the reduction of total liver blood supply. However, the data of this work show that the degree of the compensation from this so-called “hepatic artery buffer response” is insufficient to restore a proper hepatic regeneration response and delays the regenerative process after a standard partial hepatectomy. This might be due to the lower concentration of the hepatotrophic factors in the arterial blood. Interestingly, it is observed in the current work, that with a re-established same amount of blood inflow, the hepatic regeneration rate in the hepatectomized rats is equal, irrespective of the portal supply of either venous or arterial nature. These data suggest that the “required concentration” of hepatotrophic factors may not be essential, or rather that the presence of blood in the portal tract itself is a prerequisite of initiating and maintaining a normal liver regeneration. This may explain why, in ALTx, a liver graft with an arterIALIZED portal supply is sufficient to sustain the life of the patient with acute liver failure, while, with an uninterrupted portal venous supply, the recovery and regeneration of the diseased native liver can go on unaffected.<sup>59</sup>

**L**iver regeneration can be induced by a great variety of stimuli ranging from dietary alteration, to toxic damage and to loss of liver mass as after partial hepatectomy.<sup>60</sup> Determining what factors are responsible for initiating the regenerative process is one of the core issues in understanding the molecular bases of liver regeneration.<sup>61</sup> Amongst the so-called “hepatotrophic factors” discovered during the last three decades, some growth factors and cytokines were hypothesized to be the “initiators”.<sup>62-64</sup> Although the role of most of those growth regulators has been identified as crucial for a specific step of hepatocyte proliferation either as a stimulator or as an inhibitor, none of them seems to be a primary mitogen.<sup>64, 65</sup> Moreover, it has recently been shown that hepatocytes need to be “sensitized” before replicating, which means that they have to transit from a  $G_0$  resting state to a  $G_1$  replicative state, in order to be competent and fully responsive to the trophic action of the growth factors.<sup>66, 67</sup> It has been found that this so-called “priming phase” begins simultaneously in all resting hepatocytes almost immediately after partial hepatectomy.<sup>68</sup> As the production of growth factors and cytokines is not an immediate event, early hepatic regenerative response following partial hepatectomy is more likely to be activated either by hormonal factors already present in the blood or in the liver, or by changing hemodynamic factors. The most remarkable event instantly occurring after partial hepatectomy is the substantial increase of the portal blood flow to the residual liver mass with the consequent rise of portal pressure.<sup>69, 70</sup> This significant hemodynamic change could be one of the first triggers that

initiate the regeneration cascade in the hepatectomised liver. Studies have shown that, in parallel to an early and persistent increase in portal pressure, changes in hepatocyte membrane potential can be recorded as early as within the first five minutes after liver resection.<sup>71</sup> This is linked to a series of events essential for hepatocytes replication.<sup>68, 72-74</sup> In addition, following partial hepatectomy, consequent alterations in hepatic ultra-structure have been documented involving sinusoid dilatation and disappearance of the sieve-plate arrangement of small endothelial pores. This leaves the surface of the parenchymal liver cell directly exposed to the circulating hepatotrophic substances in the portal blood.<sup>70</sup> Moreover, an increased portal pressure reflecting wall shear stress of sinusoids causes nitric oxide release.<sup>75</sup> This leads to an increased mRNA expression of *c-fos*, an immediate-early gene,<sup>76, 77</sup> the protein product of which is known to participate in the stimulation of cell growth.<sup>78</sup> It has however been shown that, in a portal branch ligation model, *c-fos* as well as other factors such as *c-myc*, *c-jun*, NF- $\kappa$ B, STAT3 and IL-6 are similarly induced in both proliferating and atrophying liver lobes, and even in the livers of sham operated animals. Only the actual liver mass present during the period corresponding to mid- to late G<sub>1</sub> phase seems, in this case, to determine the magnitude of the proliferative response.<sup>79-82</sup> This could suggest that the observed early regenerative response after liver resection is not specifically related to either increased portal flow or loss of liver mass; or that still other not yet identified factors are the true regenerative triggers. Further clarification on how the hemodynamic changes and / or the degree of liver mass reduction modulates the extent of hepatic regeneration after partial hepatectomy may elucidate another piece of the complex puzzles in the liver regeneration process, which has still kept some of its intriguing secrets.

## REFERENCES

1. Hess F, Jerusalem C, Heyde MN. Advantages of auxiliary liver homotransplantation in rats. Arch Surg 1972; 104 (1):76-80.
2. Rokicki M, Rokicki W. Liver regeneration in rats after complete and partial occlusion of the portal blood influx. Res Exp Med (Berl ) 1993; 193(5):305-313.
3. Nagino M, Tanaka M, Nishikimi M, Nimura Y, Kubota H, Kanai M, Kato T et al. Stimulated rat liver mitochondrial biogenesis after partial hepatectomy. Cancer Res 1989; 49(17):4913-4918.
4. Starzl TE, Porter KA, Francavilla A. The Eck fistula in animals and humans. Curr Probl Surg 1983; 20(11):687-752.
5. Callery MP, Ricordi C, Scharp DW, Kamei T, Swanson CJ, Lacy PE, Flye MW. Hepatic insufficiency after portacaval shunting is prevented by prior intraportal pancreatic islet autotransplantation. Surgery 1989; 106(2):257-265.

## 114 CHAPTER 4 General Discussion

6. Rokicki W, Rokicki M, Czyzewski K. The effect of "end to side" portacaval anastomosis on regeneration ability of white rat liver. *Z Exp Chir Transplant Kunstliche Organe* 1990; 23(3):175-179.
7. Hess F, Willemen A, Jerusalem C. Auxiliary liver transplantation in the rat, influence of the condition of the recipient's liver on the fate of the graft. *Eur Surg Res* 1977; 9(4):270-279.
8. Greim H, Trulzsch D, Czygan P, Rudick J, Hutterer F, Schaffner F, Popper H. Mechanism of cholestasis. 6. Bile acids in human livers with or without biliary obstruction. *Gastroenterology* 1972; 63(5):846-850.
9. Campbell CB, Cowen AE. Bile salt metabolism. II. Bile salts and disease. *Aust N Z J Med* 1977; 7(6):587-595.
10. Sagawa H, Tazuma S, Kajiyama G. Protection against hydrophobic bile salt-induced cell membrane damage by liposomes and hydrophilic bile salts. *Am J Physiol* 1993; 264(5 Pt 1):G835-G839.
11. Stiehl A. [Pathophysiology of cholestasis: correlation between bile acid metabolism and liver damage]. *Z Gastroenterol* 1992; 30 Suppl 1:46-48.
12. Kullak-Ublick GA, Meier PJ. Mechanisms of cholestasis. *Clin Liver Dis* 2000; 4(2):357-385.
13. Hardison WG, Weiner RG, Hatoff DE, Miyai K. Similarities and differences between models of extrahepatic biliary obstruction and complete biliary retention without obstruction in the rat. *Hepatology* 1983; 3(3):383-390.
14. Toyota N, Miyai K, Hardison WG. Effect of biliary pressure versus high bile acid flux on the permeability of hepatocellular tight junction. *Lab Invest* 1984; 50(5):536-542.
15. Billington D, Evans CE, Godfrey PP, Coleman R. Effects of bile salts on the plasma membranes of isolated rat hepatocytes. *Biochem J* 1980; 188(2):321-327.
16. Stiehl A, Walker S, Stiehl L, Rudolph G, Hofmann WJ, Theilmann L. Effect of ursodeoxycholic acid on liver and bile duct disease in primary sclerosing cholangitis. A 3-year pilot study with a placebo-controlled study period. *J Hepatol* 1994; 20(1):57-64.
17. Benz C, Angermuller S, Tox U, Kloters-Plachky P, Riedel HD, Sauer P, Stremmel W et al. Effect of tauroursodeoxycholic acid on bile-acid-induced apoptosis and cytolysis in rat hepatocytes. *J Hepatol* 1998; 28(1):99-106.
18. Patel T, Bronk SF, Gores GJ. Increases of intracellular magnesium promote glycodeoxycholate-induced apoptosis in rat hepatocytes. *J Clin Invest* 1994; 94(6):2183-2192.
19. Patel T, Gores GJ. Apoptosis and hepatobiliary disease. *Hepatology* 1995; 21(6):1725-1741.
20. Kanai M, Tanaka M, Nimura Y, Nagino M, Katoh T, Ozawa T. Mitochondrial dysfunction in the non-obstructed lobe of rat liver after selective biliary obstruction. *Hepatogastroenterology* 1992; 39(5):385-391.
21. Spivey JR, Bronk SF, Gores GJ. Glycochenodeoxycholate-induced lethal hepatocellular injury in rat hepatocytes. Role of ATP depletion and cytosolic free calcium. *J Clin Invest* 1993; 92(1):17-24.
22. Sokol RJ, Straka MS, Dahl R, Devereaux MW, Yerushalmi B, Gumprich E, Elkins N et al. Role of oxidant stress in the permeability transition induced in

- rat hepatic mitochondria by hydrophobic bile acids. *Pediatr Res* 2001; 49(4):519-531.
23. Tracy TFJ, Bailey PV, Goerke ME, Sotelo-Avila C, Weber TR. Cholestasis without cirrhosis alters regulatory liver gene expression and inhibits hepatic regeneration. *Surgery* 1991; 110(2):176-182.
  24. Foss A, Andersson R, Ding JW, Hochbergs P, Paulsen JE, Bengmark S, Ahren B. Effect of bile obstruction on liver regeneration following major hepatectomy: an experimental study in the rat. *Eur Surg Res* 1995; 27(2):127-133.
  25. Danchenko E, Petermann H, Chirkin A, Dargel R. Effect of bile acids on the proliferative activity and apoptosis of rat hepatocytes. *Exp Toxicol Pathol* 2001; 53(2-):227-233.
  26. Poli G. Liver damage due to free radicals. *Br Med Bull* 1993; 49(3):604-620.
  27. Nakazawa H, Genka C, Fujishima M. Pathological aspects of active oxygens/free radicals. *Jpn J Physiol* 1996; 46(1):15-32.
  28. Neuschwander-Tetri BA, Nicholson C, Wells LD, Tracy TFJ. Cholestatic liver injury down-regulates hepatic glutathione synthesis. *J Surg Res* 1996; 63(2):447-451.
  29. Sokol RJ, Devereaux MW, Khandwala R. Effect of oxypurinol, a xanthine oxidase inhibitor, on hepatic injury in the bile duct-ligated rat. *Pediatr Res* 1998; 44(3):397-401.
  30. Purucker E, Winograd R, Roeb E, Matern S. Glutathione status in liver and plasma during development of biliary cirrhosis after bile duct ligation. *Res Exp Med (Berl)* 1998; 198(4):167-174.
  31. Sung JJ, Go MY. Reversible Kupffer cell suppression in biliary obstruction is caused by hydrophobic bile acids. *J Hepatol* 1999; 30(3):413-418.
  32. Aldana PR, Goerke ME, Carr SC, Tracy TFJ. The expression of regenerative growth factors in chronic liver injury and repair. *J Surg Res* 1994; 57(6):711-717.
  33. Poli G, Albano E, Dianzani MU. The role of lipid peroxidation in liver damage. *Chem Phys Lipids* 1987; 45(2-4):117-142.
  34. Sokol RJ, Devereaux M, Khandwala RA. Effect of dietary lipid and vitamin E on mitochondrial lipid peroxidation and hepatic injury in the bile duct-ligated rat. *J Lipid Res* 1991; 32(8):1349-1357.
  35. Friedman SL. Seminars in medicine of the Beth Israel Hospital, Boston. The cellular basis of hepatic fibrosis. Mechanisms and treatment strategies. *N Engl J Med* 1993; 328(25):1828-1835.
  36. Tracy TFJ, Goerke ME, Bailey PV, Sotelo-Avila C, Weber TR. Growth-related gene expression in early cholestatic liver injury. *Surgery* 1993; 114(3):532-537.
  37. Roggin KK, Papa EF, Kurkchubasche AG, Tracy TFJ. Kupffer cell inactivation delays repair in a rat model of reversible biliary obstruction. *J Surg Res* 2000; 90(2):166-173.
  38. Siegelman ES, Mitchell DG, Rubin R, Moritz MJ, Munoz SJ, Palazzo JP, Rifkin MD. Recovery of native liver after heterotopic liver transplantation for fulminant hepatic failure: MR studies. *J Comput Assist Tomogr* 1992; 16(1):152-154.

## 116 CHAPTER 4 General Discussion

39. Marchioro TL, Porter KA, Dickins TC, Faris TD, Starzl TE. Physiologic requirements for auxiliary liver transplantation. *Surg Gynecol Obstet* 1965; 121:17-31.
40. Nagashima I, Bergmann L, Schweizer R. How can we share the portal blood inflow in auxiliary partial heterotopic liver transplantation without portal hypertension? *Surgery* 1994; 116(1):101-106.
41. Chandler JG, Lee S, Krubel R, Rosen H, Nakaji NT, Orloff MJ. The inter-liver competition and portal blood in regeneration of auxiliary liver transplants. *Surg Forum* 1971; 22:341-343.
42. Shaw BWJ. Auxiliary liver transplantation for acute liver failure. *Liver Transpl Surg* 1995; 1(3):194-200.
43. Fisher B, Szuch P, Fisher ER. Evaluation of a humoral factor in liver regeneration utilizing liver transplants. *Cancer Res* 1971; 31(3):322-331.
44. Rozga J, Jeppsson B, Bengmark S. Hepatotrophic effect of portal blood during hepatic arterial recirculation. *Eur Surg Res* 1986; 18(5):302-311.
45. Kamada N, Calne RY. A surgical experience with five hundred thirty liver transplants in the rat. *Surgery* 1983; 93(1 Pt 1):64-69.
46. Chan FK, Zhang Y, Shaffer EA. Bile secretory function of the arterialized versus nonarterialized rat liver allograft. *Transplantation* 1996; 62(11):1657-1663.
47. Kamada N, Sumimoto R, Kaneda K. The value of hepatic artery reconstruction as a technique in rat liver transplantation [see comments]. *Surgery* 1992; 111(2):195-200.
48. Imamura H, Rocheleau B, Cote J, Huet PM. Long-term consequence of rat orthotopic liver transplantation with and without hepatic arterial reconstruction: a clinical, pathological, and hemodynamic study. *Hepatology* 1997; 26(1):198-205.
49. Zhao D, Zimmermann A, Kuznetsova LV, Wheatley AM. Regression of bile duct damage and bile duct proliferation in the non-rearterialized transplanted rat liver is associated with spontaneous graft rearterialization. *Hepatology* 1995; 21(5):1353-1360.
50. Svensson G, Naredi P, Hafstrom L, Tufveson G. Quantitative measurements of collateral arterial blood flow in nonarterialized rat liver grafts. *Transpl Int* 1994; 7(2):136-139.
51. Howden B, Jablonski P, Grossman H, Marshall VC. The importance of the hepatic artery in rat liver transplantation. *Transplantation* 1989; 47(3):428-431.
52. Sumimoto R, Shinomiya T, Yamaguchi A. Influence of hepatic arterial blood flow in rats with liver transplants. Examination of donor liver-derived serum class I MHC antigen in rats with liver transplants with or without hepatic arterial reconstruction. *Transplantation* 1991; 51(5):1138-1139.
53. Post S, Menger MD, Rentsch M, Gonzalez AP, Herfarth C, Messmer K. The impact of arterialization on hepatic microcirculation and leukocyte accumulation after liver transplantation in the rat. *Transplantation* 1992; 54(5):789-794.
54. Zhao D, Zimmermann A, Wheatley AM. Morphometry of the liver after liver transplantation in the rat: significance of an intact arterial supply. *Hepatology* 1993; 17(2):310-317.

55. Gao W, Lemasters JJ, Thurman RG. Development of a new method for hepatic rearterialization in rat orthotopic liver transplantation. Reduction of liver injury and improvement of surgical outcome by arterialization. *Transplantation* 1993; 56(1):19-24.
56. Reck T, Steinbauer F, Steinbauer M, Schwille PO, Wittekind C, Hohenberger W, Kockerling F. Impact of arterialization on hepatic oxygen supply, tissue energy phosphates, and outcome after liver transplantation in the rat. *Transplantation* 1996; 62(5):582-587.
57. Slapak M. Fulminant liver failure: clinical and experimental study. *Ann R Coll Surg Engl* 1975; 57(5):234-247.
58. Burdelski M, Rogiers X. Liver transplantation in metabolic disorders. *Acta Gastroenterol Belg* 1999; 62(3):300-305.
59. Schleimer K, Lange R, Rauhen U, Nowak B, Brandt-Mainz K, De Groot H, Erhard J. Auxiliary rat liver transplantation with portal vein arterialization in acute hepatic failure. *Transplantation* 2000; 70(1):73-78.
60. Fausto N, Mead JE. Regulation of liver growth: protooncogenes and transforming growth factors. *Lab Invest* 1989; 60(1):4-13.
61. Taub R. Liver regeneration 4: transcriptional control of liver regeneration. *FASEB J* 1996; 10(4):413-427.
62. Gentile JM, Grace JTJ. A cell growth stimulating factor in partially hepatectomized rat serum. *Surg Forum* 1968; 19:62-63.
63. Morley CG, Kingdon HS. The regulation of cell growth. I. Identification and partial characterization of a DNA synthesis stimulating factor from the serum of partially hepatectomized rats. *Biochim Biophys Acta* 1973; 308(2):260-275.
64. Michalopoulos GK, DeFrances MC. Liver regeneration. *Science* 1997; 276(5309):60-66.
65. LaBrecque D. Liver regeneration: a picture emerges from the puzzle. *Am J Gastroenterol* 1994; 89(8 Suppl):S86-S96.
66. Moolten FL, Oakman NJ, Bucher NL. Accelerated response of hepatic DNA synthesis to partial hepatectomy in rats pretreated with growth hormone or surgical stress. *Cancer Res* 1970; 30(9):2353-2357.
67. Bucher NL. Liver regeneration then and now. In: Jirtle RL, editor. *Liver Regeneration and Carcinogenesis: Molecular and Cellular Mechanisms*. San Diego: Academic Press, 1995: 1-25.
68. Kren BT, Trembley JH, Fan G, Steer CJ. Molecular regulation of liver regeneration. *Ann N Y Acad Sci* 1997; 831:361-381.
69. Fogli L, Gorini P, Cappellari L, Morsiani E. Effect of partial hepatectomy and liver regeneration on portal pressure in rats. *Surgical Research Communications* 1990; 6:159-166.
70. Morsiani E, Aleotti A, Ricci D. Haemodynamic and ultrastructural observations on the rat liver after two-thirds partial hepatectomy. *J Anat* 1998; 192(Pt 4):507-515.
71. Zhang XK, Gauthier T, Burczynski FJ, Wang GQ, Gong YW, Minuk GY. Changes in liver membrane potentials after partial hepatectomy in rats. *Hepatology* 1996; 23(3):549-551.
72. Wondergem R, Potter VR. A new protocol for studying the early events during liver regeneration. *Life Sci* 1978; 23(15):1565-1575.



## 118 CHAPTER 4 General Discussion

73. Schenk DB, Hubert JJ, Leffert HL. Use of a monoclonal antibody to quantify (Na<sup>+</sup>,K<sup>+</sup>)-ATPase activity and sites in normal and regenerating rat liver. *J Biol Chem* 1984; 259(23):14941-14951.
74. Moule SK, McGivan JD. Regulation of the plasma membrane potential in hepatocytes--mechanism and physiological significance. *Biochim Biophys Acta* 1990; 1031(3):383-397.
75. Kelm M, Feelisch M, Deussen A, Strauer BE, Schrader J. Release of endothelium derived nitric oxide in relation to pressure and flow. *Cardiovasc Res* 1991; 25(10):831-836.
76. Feelisch M, Ostrowski J, Noack E. On the mechanism of NO release from sydnonimines. *J Cardiovasc Pharmacol* 1989; 14 Suppl 11:S13-S22.
77. Schoen JM, Wang HH, Minuk GY, Lautt WW. Shear stress-induced nitric oxide release triggers the liver regeneration cascade. *Nitric Oxide* 2001; 5(5):453-464.
78. Angel P, Karin M. The role of Jun, Fos and the AP-1 complex in cell-proliferation and transformation. *Biochim Biophys Acta* 1991; 1072(2-3):129-157.
79. Lambotte L, Saliez A, Triest S, Tagliaferri EM, Barker AP, Baranski AG. Control of rate and extent of the proliferative response after partial hepatectomy. *Am J Physiol* 1997; 273(4 Pt 1):G905-G912.
80. Starkel P, Horsmans Y, Sempoux C, De Saeger C, Wary J, Lause P, Maiter D et al. After portal branch ligation in rat, nuclear factor kappaB, interleukin-6, signal transducers and activators of transcription 3, c-fos, c-myc, and c-jun are similarly induced in the ligated and nonligated lobes. *Hepatology* 1999; 29(5):1463-1470.
81. Lambotte L, Li B, Leclercq I, Sempoux C, Saliez A, Horsmans Y. The compensatory hyperplasia (liver regeneration) following ligation of a portal branch is initiated before the atrophy of the deprived lobes. *J Hepatol* 2000; 32(6):940-945.
82. Laurent S, Otsuka M, De Saeger C, Maiter D, Lambotte L, Horsmans Y. Expression of presumed specific early and late factors associated with liver regeneration in different rat surgical models. *Lab Invest* 2001; 81(9):1299-1307.

## SUMMARY

Although auxiliary liver transplantation does offer advantages over orthotopic liver transplantation, in particular in treating patients with acute hepatic failure and children with hepatic-based inborn errors of metabolism, its results are often compromised by the interactions between the coexisting engrafted liver and native liver. To date, it is generally accepted that the process and final outcome of the so-called inter-liver competition is governed by the supply of portal blood and, most crucial of all, the distribution of hormonal “hepatotrophic factors” between the two livers. However, other factors such as the physiological status of each individual liver might also play an important role in this mechanism. It was, therefore, the purpose of the current work, to investigate whether a change in the functional condition of each individual liver would influence the fate of the two livers using a rat model of heterotopic auxiliary liver transplantation (HALTx), and further to re-examine the controversy on whether changing the content of the portal blood would alter the hepatic regeneration capacity after a standard partial hepatectomy.

To define indicative biological parameters for post-HALTx evaluation of graft viability, a 68% partial hepatectomy (68% PH) and a common bile duct ligation (CBDL) were performed to hamper the native liver’s functionality. Commonly used serum biochemical liver parameters were analysed. It was found that the surgical handicap on the native liver did efficiently induce lethal hepatic failure, as, in this case, animal survival relied solely upon the viability of the engrafted liver. The liver biology therefore reflected directly the graft functionality. The post-operative values of serum bilirubin concentration, gamma glutamyl transferase activity, alkaline phosphatase, alanine aminotransferase activity and aspartate aminotransferase activity were significantly lower in the transplanted rats than in the control rats. These data reveal that a viable engrafted liver can indeed reduce the deleterious effect of cholestasis caused by the surgical handicap on the native liver (Chapter 2.1.).

In a further study, for the first time, magnetic resonance imaging (MRI) was tested for its potential to monitor post-transplant hepatic viability in this rat model of HALTx. The results showed, firstly, that post-transplant hepatic complications as abscess and necrosis could be depicted separately in the coexisting two livers on MR image. Secondly, it was found that the better the hepatic viability, the higher the signal intensity on  $T_1$  weighted image. The viability of each individual liver could thus be assessed by its signal intensity at any given time. Thirdly, in the presence of a viable engrafted liver, the expected drastic functional decline of the native liver reflected by

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its  $T_1$  relaxation times became obvious only 4 weeks after transplantation. This showed clearly the role of the well-functioning auxiliary liver in reducing the mechanical and biochemical aggression of the inflicted handicap on the native liver. Fourthly, as soon as after the first post-transplant week, the healthy engrafted livers could already be distinguished from those that would develop complications by a significant decrease in  $T_1$  relaxation times. This non-invasive measurement does facilitate systematic follow-up of the evolution of the two livers after HALTx. MRI is thus a very promising tool for the studies on inter-liver functional competition. Prospective human studies based on these new discoveries may further demonstrate the beneficial potential of MRI in clinical situations such as pre-surgical evaluation of transplant candidates in fulminant or sub-fulminant hepatic failure and post-transplant or post-hepatectomy assessment of liver viability (Chapter 2.2.).

In order to study specifically the effects of various hepatic physiological conditions on the inter-liver competition, the possible influence of portal blood distribution to the native liver was abolished by completely shunting it to the engrafted liver. This allowed focusing on how the different handicaps of the native liver could affect animal survival and the regeneration capacity of both the engrafted and the residual native livers. It was observed that a viable auxiliary liver was able to prevent high animal mortality caused by a 68% PH on a portal deprived native liver. The native liver could recover from this functional reduction and even regenerate, while the engrafted liver atrophied despite its proper portal blood supply. An additional handicap on the native liver, through a combined 68% PH and CBDL, resulted this time in atrophy of the native liver and regeneration of the engrafted organ (Chapter 3.1.). In another experimental setting, the effects of graft re-arterialisation on liver regeneration were evaluated. It was shown that, the re-arterialised grafts had significantly higher bile flow rate and bilirubin excretion immediately after re-perfusion compared with that of the grafts without re-arterialisation. Re-arterialisation also increased DNA synthesis rate of the engrafted liver, and the rat developed less cytolysis and less cholestasis. This led to a significantly improved graft growth, which was paralleled to a more pronounced native liver atrophy. Re-arterialisation could indeed enhance the functional advantage of the engrafted liver (Chapter 3.2.).

In the last part of this work, it was found that complete absence of portal blood supply to a partial resected liver caused a marked inhibition of early regenerative response and liver growth rate. The degree of the compensation from the “hepatic artery buffer response” was insufficient to restore normal hepatic regeneration after a standard partial hepatectomy. Notably, if the amount of portal blood inflow was re-established by arterialising the portal

stump, the DNA synthesis rate as well as liver growth could be sustained at a level equivalent to that observed in the rats of control group (Chapter 3.3.).

The data of the current work emphasize that, besides the portal blood supply, the change of the physiological condition of each individual liver does affect the balance of the inter-liver competition after a HALTx. Rather than the high concentration of the hepatotrophic factors, the presence of sufficient blood flow in the portal tract, of either venous or arterial origin, appears a prerequisite to maintain the normal chronology and the intensity of the regeneration process.

## SAMENVATTING

Een auxilliaire levertransplantatie biedt duidelijke voordelen aan bij de behandeling van patiënten lijdende aan acuut leverfalen, alsook bij kinderen met congenitale metabole ziekten. Ondanks de conceptuele aantrekkelijkheid van deze type transplantatie, zijn de resultaten hiervan niet steeds éénduidig en dit ondermeer omwille van het ontstaan van een functionele competitie tussen de eigen of natieve lever en de getransplanteerde lever. Vandaag wordt het algemeen aanvaard dat de portale bevoeiing van de lever met zijn hepatotropische factoren de functionele status van de 2 levers in grote mate beïnvloedt. Buiten deze extrinsieke factoren moet men nochtans ook rekening houden met levergebonden intrinsieke factoren wanneer gesproken wordt over de functionele competitie tussen de 2 levers. De fysiologische status van de 2 levers speelt inderdaad een belangrijke rol in het min of meer receptief staan van het orgaan ten opzichte van externe stimulatiefactoren. Het was dan ook het doel van ons onderzoek om op een rattenmodel van heterotopie auxilliaire levertransplantatie (HALTx) verschillende functionele beperkingen van donor- en recipiëntlevers te testen en na te gaan in welke mate deze factoren de competitie tussen de recipiëntlever en de auxilliaire lever zouden kunnen beïnvloeden. Hiernaast hebben wij ons eveneens de vraag gesteld of extrinsieke factoren met dan hier meer expliciet hemodynamische factoren implicaties zouden kunnen hebben op de lever regeneratie capaciteit na een standaard partiële hepatectomie.

Om met routine biologische parameters de post-HALTx viabiliteit en functie van het getransplanteerd orgaan te kunnen volgen, werd de natieve lever met een 68% partiële hepatectomie en galwegen ligatuur belast. Deze heeldkundige manipulatie van de natieve lever veroorzaakt een dodelijk leverfalen in de afwezigheid van een getransplanteerde auxilliaire leverent. Indien de overleving van de ratten rechtstreeks afhankelijk is van de auxilliaire lever, zal de biologie grotendeels de functie van deze ent weerspiegelen. De serumconcentraties van bilirubine, gamma GT, alkalische fosfatase, gamma glutamyl transferase, alanine aminotransferase activiteit en aspartaat aminotransferase activiteit van de getransplanteerde groep zijn inderdaad duidelijk lager dan deze van de controlegroep. Deze gegevens tonen aan dat in dit model de functionele levertransplant het toxisch effect van de cholestase op de natieve lever kan tegenwerken (deel 2.1).

Om de evolutie en de viabiliteit van de 2 levers na auxilliaire levertransplantatie snel en herhaaldelijk te kunnen volgen werd voor de eerste keer nucleair resonantie-onderzoek gebruikt (MRI). Het onderzoek maakte het mogelijk om in eerste instantie de postoperatieve levercomplicaties zoals abcessen en parenchymateuze aantastingen van elke

individuele lever vroegtijdig te kunnen diagnosticeren. Ten tweede kon men via de signaalintensiteit op de  $T_1$  gewogen beelden de viabiliteit van elke lever beoordelen, waarbij hogere intensiteitsbeelden werden bekomen bij betere leverfunctie. Eindelijk maakte deze studie het mogelijk om de evolutie van de  $T_1$  relaxatietijden van de 2 levers te vergelijken in een normale toestand en in geval van levercomplicaties. De finale evolutie van de 2 levers kon op deze manier reeds vanaf de eerste postoperatieve week voorspeld worden. Deze gevoelige en niet-invasieve meting vergemakkelijkt duidelijk de follow-up van de 2 levers na HALTx. MRI is dus een veelbelovend werkinstrument in de studie van de functionele competitie tussen 2 levers. Prospectieve studies bij de mens zouden op dit vlak zeker interessante resultaten kunnen leveren bij de selectie van patiënten lijdend aan fulminant of subfulminant leverfalen en voor de bepaling van de leverfunctionaliteit na transplantatie of na partiële leverresectie (deel 2.2.).

In het gebruikte model van HALTx was de portale bloedtoevoer van de recipiënt lever volledige geshunt naar de getransplanteerde lever, terwijl de viabiliteit van de natieve lever enkel verzekerd werd door bevoeiing via de arteria hepatica. Ondanks het feit dat het portaal bloed exclusief naar de transplantlever geshunt wordt, blijkt de evolutie van deze ent naar regeneratie of atrofie rechtstreeks gebonden aan de graad van de handicap aangebracht aan de eigen lever. En een 68% PH, en een galwegen ligatuur van de eigen lever is nodig om atrophie van de ent te voorkomen (deel 3.1.). Anderzijds, konden wij tonen dat een rearterialisatie van de getransplanteerde lever toegevoegd een de portale revascularisatie een duidelijk meer uitgesproken stimulatie van de galexcretie na reperfusie met zich meebrengt; rearterialisatie gaf ook een hogere DNA synthese, minder uitgesproken cytolyse en cholestase. Met rearterialisatie kan men dan ook een significant betere entoverleving bekomen. De functionele optimalisatie van de ent heeft een rechtstreekse repercussie op de functionele competitie tussen de 2 levers (deel 3.2.).

In het laatste deel van het werk (deel 3.3) konden we aantonen dat in een klassiek model van partiële leverresectie een volledige portale derivatie de regeneratieve capaciteit van de lever significant verminderd en aldus het herstel van de levermassa na leverresectie verhinderd. De spontane arteriële compensatie na de portale shunt was onvoldoende om een normale regeneratieve respons na partiële hepatectomie te verzekeren. Restitutio van het totale leverdebiet via areterialisatie van de portale stomp kon anderzijds, merkwaardig genoeg, wel de normale respons herstellen. Deze resultaten suggereren dat hoge concentraties van hepatotrofische factoren niet essentieel zijn bij de initiële fase van leverregeneratie.

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In conclusie, het zijn dus niet enkel de portale stimulerende groeifactoren en nutritieve elementen, maar het is ook en vooral de algemene functionele status van elke lever welke een rechtstreekse invloed heeft op de regeneratieve capaciteit van elk individueel organ. Een voldoende debiet, dat het weze met arterieel of veneus bloed, via het portaal systeem naar de lever moet verzekerd worden om een normale regeneratiechronologie na partiële leverresectie te bekomen.

## RESUME

Si la transplantation auxiliaire du foie présente des avantages théoriques évidents chez les patients souffrants d'insuffisance hépatique aigue ainsi que chez les enfants atteints de maladie métabolique congénitale, les résultats après ce type de transplantation ne sont pas toujours aussi clairement positifs. L'interaction qui a lieu entre le foie donneur et le foie receveur joue ici, parmi d'autres, un rôle important. Il est généralement admis que l'apport de sang portal contenant les facteurs hormonaux hépatotrophiques influence directement le devenir des foies mis en compétition fonctionnelle. D'autres facteurs tel que l'état physiologique propre de chaque foie individuel joue également un rôle dans le devenir des organes mis en compétition. Ce fut donc le but du présent travail d'étudier l'influence de l'état physiologique et de l'état fonctionnel des foies en présence sur la balance compétitive interhépatique, et ce, à l'aide du modèle de transplantation auxiliaire hétérotopique chez le rat (HALTx). En annexe nous avons tenté de réexaminer la controverse au sujet de la question concernant le rôle prépondérant, soit qualitatif, soit quantitatif de l'apport portal du sang sur la régénération hépatique après hépatectomie partielle standard.

Afin de suivre dans le modèle de transplantation auxiliaire la fonction et la viabilité du greffon à l'aide de paramètres biologiques courants, le foie hôte fut soumis à une hépatectomie de 68% et une ligature de la voie biliaire principale. Ces manipulations chirurgicales du foie hôte sont responsables d'une insuffisance hépatique mortelle en l'absence d'un greffon auxiliaire. Si la survie des rats est dans ce cas directement dépendant du foie greffé, la biologie de l'animal sera un reflet direct de sa fonction. Comparés aux groupes contrôles, les groupes transplantés ont des concentrations sériques inférieures en bilirubine, gamma GT, phosphatase alcaline, d'activité de gamma glutamyl transférase et alanine aminotransférase. Ces résultats montrent que dans ce modèle, un greffon fonctionnel peut prévenir l'effet toxique de la cholestase du foie hôte.

Pour l'étude suivante la Résonance Magnétique Nucléaire (RMN) fut utilisée pour la première fois afin de suivre la viabilité postopératoire hépatique dans le modèle de HALTx. Ce travail a tout d'abord pu mettre en évidence pour chaque foie individuel, l'apparition de complications postopératoires tels qu'abcès hépatique et nécrose parenchymateuse. Cet examen a montré en deuxième lieu que, plus la fonctionnalité hépatique est conservée plus le degré d'intensité du signal des images pesées en  $T_1$  est prononcé. L'évolution de la viabilité des 2 foies pouvait ainsi être documentée et comparée à tout moment du suivi postopératoire. En comparant l'évolution du temps de



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relaxation en  $T_1$  des 2 foies, l'RMN peut en outre prédire dès la première semaine postopératoire le devenir des foies qui resteront sains et ceux qui présenteront des complications. L'RMN apparait comme un outil non-invasif fort utile pour le suivi de l'évolution des 2 foies après transplantation auxiliaire et est prometteur dans l'étude de la compétition fonctionnelle interhépatique. Des études humaines prospectives en RMN pourraient s'avérer dès lors fort intéressantes non seulement pour prédire le devenir d'un foie atteint d'hépatite fulminante ou subfulminante mais aussi de pronostiquer la viabilité d'un greffon hépatique ou d'un foie restant après résection hépatique étendue (partie 2.2.).

Dans le modèle de HALTx utilisé dans ce travail, l'apport sanguin portal fut totalement shunté vers le greffon alors que le foie natif n'était irrigué que par l'artère hépatique. Malgré le fait que la totalité du sang portal fut dérivé en direction du foie transplanté, l'évolution de celui-ci vers la régénération ou l'atrophie dépendait directement du degré d'handicap du foie hôte (partie 3.1.). Par ailleurs, la réartérialisation du greffon ajouté a sa vascularisation portale influence directement et de façon significative la quantité de production biliaire posttransplantation et celui de cytolysse et de cholestase ainsi que le degré de synthèse d'ADN du foie greffé. La réartérialisation du greffon a permis d'obtenir une survie significativement meilleure des animaux ainsi traités. La meilleure fonctionnalité du foie greffé après artérialisation a logiquement un effet direct sur la compétition interhépatique (partie 3.2.).

La dernière partie du travail (partie 3.3.) montre que l'absence totale de l'apport de sang portal affecte significativement la régénération du foie et la restauration de la masse hépatique après hépatectomie partielle. La compensation vasodilatatrice de l'artère hépatique suite à la réduction du débit hépatique total après shunt porto-cave est insuffisante pour maintenir une régénération normale après hépatectomie partielle. Toutefois, la régénération est normalisée si le moignon portal est artérialisé et le débit hépatique total restauré à des valeurs équivalentes à celles des groupes contrôles. Ces résultats suggèrent que de fortes concentrations de facteurs hépatotroques ne sont pas indispensables dans la phase initiale de régénération hépatique.

En conclusion, non seulement l'apport portal et son contenu en hormones de croissances stimulantes ainsi que d'éléments nutritifs, mais aussi et surtout l'état physiologique du foie greffé influence la capacité régénérative des 2 foies dans le modèle de HALTx. Une quantité suffisante de sang artériel ou portal doit être véhiculé par la veine porte aux hépatocytes pour assurer une chronologie et intensité normale de la régénération après hépatectomie partielle.

## ABSTRACT

This work was conducted in rat models to investigate whether the factors other than known portal hormonal elements could also influence the inter-liver competition in heterotopic auxiliary liver transplantation (HALTx), and whether the quality or the quantity of portal supply was more important for hepatic regeneration.

The results showed that the handicap of a 68% partial hepatectomy (68% PH) plus a common bile duct ligation (CBDL) could drastically hamper native liver's functionality and post-HALTx animal survival relied solely upon graft's viability. Thus, the significantly lower values of serum biochemistry in the transplanted rats indicated directly that the graft was well functioning.

On a  $T_1$  weighted magnetic resonance image, as observed, hepatic complications could be depicted individually in the coexisting two livers and hepatic viability could be assessed by its signal intensity. The healthy grafts could be distinguished from those that later developed complications by a significant decrease in  $T_1$  relaxation times at an early post-transplant stage.

Despite with proper portal supply, the engrafted liver would atrophy if the native liver was handicapped with a 68% PH alone, whereas graft regeneration was assured by a more severe handicap (adding a CBDL). Furthermore, when the graft's physiological status was improved through re-establishing its arterial supply, it had not only a better early function (immediate bile flow rate and bilirubin excretion) and less biliary complications (cytolysis and cholestasis), but also more pronounced liver regeneration (DNA synthesis rate and graft growth) compared with that of control group.

Complete absence of portal supply to a partial resected liver caused a marked inhibition of early regenerative response and liver growth. When an equivalent amount of portal blood inflow was re-established by arterialising the portal stump, DNA synthesis rate and liver growth could be sustained at a level comparable to that of control rats.

These data emphasize that, besides portal supply, a change of the physiological condition of each individual liver does affect the balance of the inter-liver competition after HALTx. Rather than a high concentration of hormonal factors, the presence of sufficient blood flow in the portal tract, of either venous or arterial origin, appears a prerequisite to maintain a normal chronology and intensity of hepatic regeneration.